References

- de Lacey G, Record C, Wade J. How accurate are quotations and references in medical journals? BMJ 1985;291:884-6.
- Holt S, Siebers R, Suter A, Loan R, Jefferey O. The accuracy of references in Australian and New Zealand medical journals. N Z Med J 2000;113:416–7.
- Roach VJ, Lau TK, Kee WD. The quality of citations in major international obstetrics and gynecology journals. Am J Obstet Gynecol 1997;177:973–5.
- Evans JT, Nadjari HI, Burchell SA. Quotational and reference accuracy in surgical journals. A continuing peer review problem. JAMA 1990; 263:1353–4.
- Siebers R. The accuracy of references of three allergy journals. J Allergy Clin Immunol 2000; 105:837–8.
- Siebers R. Accuracy of references in the New Zealand Journal of Medical Laboratory Science. N Z J Med Lab Science 1999;53:46–8.
- International Committee of Medical Journal Editors. Uniform requirements for manuscripts submitted to biomedical journals. Ann Intern Med 1997;126:36–47.
- Pitkin RM, Branagan MA, Burmeister LF. Accuracy of data in abstracts of published research articles. JAMA 1999;281:1110–1.
- **9.** Cartwright CP. Synthetic viral particles promise to be valuable in the standardization of molecular diagnostic assays for hepatitis C virus [Editorial]. Clin Chem 1999;45:2057–9.
- Toennes SW, Maurer HH. Efficient cleavage of conjugates of drugs or poisons by immobilized β-glucuronidase and arylsulfatase in columns. Clin Chem 1999;45:2173–82.
- Meijer WG, Swaanenburg JCJM, van Veldhuisen DJ, Kema IP, Willemse PHB, de Vries EGE. Troponin I, troponin T, and creatine kinase-MB mass in patients with the carcinoid syndrome with and without heart failure [Letter]. Clin Chem 1999;45:2296–7.
- Datta P, Foster K, Dasgupta A. Comparison of immunoreactivity of five human cardiac troponin I assays toward free and complexed forms of the antigen: implications for assay discordance [Technical Brief]. Clin Chem 1999;45: 2266–9.
- 13. Batstra MR, van Driel A, Peterson JS, van Donselaar CA, van Tol MJ, Bruining GJ, et al. Glutamic acid decarboxylase antibodies in screening for autoimmune diabetes: influence of comorbidity, age, and sex on specificity and threshold values [Technical Brief]. Clin Chem 1999;45:2269–72.
- Bollhalder M, Mura C, Landt O, Maly FE. Light-Cycler PCR assay for simultaneous detection of the H63D and S65C mutations in the *HFE* hemochromatosis gene based on opposite melting temperature shifts [Technical Brief]. Clin Chem 1999;45:2275–8.
- Horn PS, Pesce AJ, Copeland BE. Reference interval computation using robust vs parametric and nonparametric analyses [Technical Brief]. Clin Chem 1999;45:2284–5.

Robert Siebers

Department of Medicine Wellington School of Medicine PO Box 7343 Wellington South, New Zealand Fax 64-4-389-5427 E-mail rob@wnmeds.ac.nz *Editor's Note:* The accuracy of reference listings is important for investigators and clinicians, and no less so in the online era. In *Clinical Chemistry Online*, references are linked to the full text of cited articles or to their abstracts at Medline. This linking requires accurate citations.

I examined the reference linking in the first 21 pieces in the December 1999 issue of *Clinical Chemistry* (the issue studied by Siebers) at www. clinchem.org. Among 440 references to articles in journals that are indexed at Medline, 409 (93%) were linked to full text of the articles or to Medline entries. The remaining 7% that were not linked presumably represent a subset of the 25% of articles in which Siebers found some errors in the citation.

The author of the Letter above examined the same online issue for us. He reports that references were not linked when they had errors in the year, volume number, first page number, journal name (or its abbreviation), or name of the first author. References that were linked included references with errors in co-authors' names or ending page numbers, spelling errors in the title, simple spelling errors of the first author's name, transpositions of authors' names, and omissions of authors' initials. This information sheds additional light on the types of errors that were the most common.

Authors' errors in references should become exceedingly rare with the current availability of programs that import citations directly from Medline. We encourage authors to avail themselves of these tools (which also save hours in manuscript preparation). Citation errors are glaringly obvious in the online journal. In the near future, we hope to provide additional electronic tools to help authors to identify errors in their reference listings. It will remain the author's responsibility to the community to check the accuracy of the references. We cannot know the references that the author has in mind.

-DB

Prostaglandin D Synthase Does Not Produce Prostate-specific Antigen Cross-Reactivity in Renal Cell Carcinoma

To the Editor:

Immunoreactive prostate-specific antigen (PSA) has been detected in the sera of female and male renal cell carcinoma (RCC) patients in several studies (1-3). These measurements were attributed to the tumor because PSA reverted to undetectable concentrations after nephrectomy. However, attempts to definitively ascribe this increase to PSA were not successful either by immunohistochemistry with PSA monoclonal antibodies (1, 2) or by amplification of PSA by reverse transcription-PCR (RT-PCR) (3), suggesting cross-reaction with a PSA-like protein. Prostaglandin D synthase (PGDS) in amniotic fluid has been found to cross-react with a PSA polyclonal antibody, but not with PSA monoclonal antibodies (4). PGDS is present in the kidney (5)and is increased in the serum of patients with renal failure (6, 7). Because the Chiron PSA immunoassay (ACS:180) used in our original study (3) utilizes a polyclonal antibody, albeit immunopurified, we investigated whether PGDS could be responsible for the increased concentrations of PSA detected in RCC patients.

RNA was extracted from six female and six male tumor samples and from nondiseased kidney tissue adjacent to the tumor. A 573-bp fragment of PGDS was amplified by RT-PCR. Southern blot hybridization and DNA sequencing of the PCR product confirmed the presence of PGDS. PGDS was expressed in both the nondiseased kidney samples adjacent to the tumor and in most tumor samples, but was not up-regulated in the tumor (Fig. 1).

Serum from the six female RCC patients was also assayed for PSA, using the ACS:180 assay and the ultrasensitive PSA immunofluorometric assay developed by Yu and Diamandis (8), and for PGDS, using the immunoassay developed by Melegos et al. (9). The results for these six

male

-ve

-ve

female

female

NK

Fig. 1. RT-PCR and Southern blot hybridization of PGDS expression in nondiseased kidney tissue adjacent to the tumor (NK; *top*) and RCC tissue from 6 female (5 NK and 6 RCC tissue samples, respectively) and 6 male patients with RCC (*bottom*).

The DNA markers (*lane 1*, marker IX; Roche) and the negative control (*lane -ve*, no cDNA) are also shown.

patients, respectively, were as follows: PSA (ACS:180), 0.89, 0.54, 0.39, 0.27, 0.26, and 0.15 µg/L (concentrations in healthy females were undetectable at <0.04 μ g/L); PSA (Yu assay), undetectable; PGDS, 798, 281, 294, 480, 366, and 705 mg/L. There was no correlation between the serum samples that exhibited the highest PSA concentrations and those with higher PGDS. In fact, the PGDS concentrations, although variable, were all within the reference interval. Of interest was the inability of the monoclonal-based Yu assay to detect PSA.

From these preliminary data, it appears that PGDS expression is not increased in RCC and that PGDS is unlikely to be the source of the crossreacting antigen detected previously in the serum of women with RCC. In keeping with these findings, we have determined that PGDS does not cross-react with several commercially available PSA antibodies and assay systems (data not shown). Although PSA could not be detected here with a more specific and sensitive PSA assay, PSA expression was detected recently in some RCC cell lines (10) and a cDNA library (11). These findings are yet to be extrapolated to human tissue. Other potential PSA-related antigens are currently being examined as candidates for the cross-reacting protein.

References

- Pummer K, Wirnsberger G, Pürstner P, Stettner H, Wandschneider G. False positive prostate specific antigen values in the sera of women with renal cell carcinoma. J Urol 1992;148: 21–3.
- 2. Geisler E, Andaz S, Nirmul P, Gheewala A,

Sehonanda A, Gerst P. False-positive prostatic-

male

Sehonanda A, Gerst P. False-positive prostaticspecific antigen in the serum of a man with renal cell carcinoma. Br J Urol 1997;79:299– 300.

- Clements J, Ward G, Kaushal A, Hii SI, Kennett C, Nicol D. A prostate specific antigen-like protein associated with renal cell carcinoma in women. Clin Cancer Res 1997;3:1427–31.
- Melegos DN, Yu H, Diamandis EP. Prostaglandin D₂ synthase: a component of human amniotic fluid and its association with fetal abnormalities. Clin Chem 1996;42:1042–50.
- Eguchi Y, Eguchi N, Oda H, Seiki K, Kijima Y, Matsu-ura Y, et al. Expression of lipocalin-type prostaglandin D synthase in human heart and its accumulation in the coronary circulation of angina patients. Proc Natl Acad Sci U S A 1997;94:14689–94.
- Hoffmann A, Nimtz M, Conradt HS. Molecular characterization of β trace protein in human serum and urine: a potential diagnostic marker for renal diseases. Glycobiology 1997;7:499– 506.
- Melegos D, Grass L, Pierratos A, Diamandis EP. Highly elevated levels of prostaglandin D synthase in the serum of patients with renal failure. Urology 1999;53:32–7.
- Yu H, Diamandis E. Ultrasensitive time-resolved immunofluorometric assay of prostatespecific antigen in serum and preliminary clinical studies. Clin Chem 1993;39:2108–14.
- Melegos DN, Diamandis EP, Oda H, Urade, Y, Hayaishi, O. Immunofluorometric assay of prostaglandin D synthase in human tissue extracts and fluids. Clin Chem 1996;42:1984–91.
- Takahashi T, Hoshi S, Satoh M, Kaneda T, Suzuki KI, Nakagawara KI, Orikasa S. The study of PSA gene expression on urogenital cell lines. Int J Urol 1999;6:526–531.
- Rae F, Bulmer B, Nicol D, Clements J. The human tissue kallikreins and a novel *KLK*1 mRNA transcript are expressed in a renal cell carcinoma cDNA library. Immunopharmacology 1999;45:83–8.

Bronwyn Bulmer¹ Greg Ward² Eleftherios Diamandis³ David Nicol⁴ Judith Clements^{1,5*}

 ¹ Co-operative Research Centre for Diagnostic Technologies and
⁵ Centre for Molecular Biotechnology School of Life Sciences Queensland University of Technology GPO Box 2434 Brisbane 4001 Queensland, Australia

Departments of ² Chemical Pathology and ⁴ Urology Princess Alexandra Hospital Woolloongabba 4102 Queensland, Australia

³ Department of Pathology and Laboratory Medicine Mount Sinai Hospital Toronto, Ontario, M5G 1X5 Canada

*Address correspondence to this author at: Center for Molecular Biotechnology, School of Life Sciences, Queensland University of Technology, GPO Box 2434, Brisbane 4001, Queensland, Australia. Fax 61-7-38641534; e-mail j.clements@ qut.edu.au.

Plasma Cardiac Troponin Concentrations after Extreme Exercise

To the Editor:

The New Zealand Ironman competition is an international ultradistance triathlon in which each athlete swims 3.8 km, cycles 180 km, and runs 42.2 km on the same day, completing the event in a time ranging from 9 to 16 h. In 1998, the race was held on March 15. A summary of the medical complications of the race and their treatment has been published separately (1). During and immediately after the race, 134 of the 650 starting athletes presented to the race medical facility for advice and treatment. Of these, 64 underwent venipuncture for measurement of plasma electrolytes because of clinical suspicion of acute hyponatremia (2). The residual blood from these tests was used in the study reported here.

Athletes withdrew from the race because of injury or exhaustion when necessary. Those who presented for medical treatment were asked for informed consent, either at presenta-

608