Ovarian Cancer

Second Edition

For further volumes: http://www.springer.com/series/5808

## **Cancer Treatment and Research** Steven T. Rosen, MD, *Series Editor*

M. Sharon Stack • David A. Fishman Editors

# **Ovarian Cancer**

Second Edition



*Editors* M. Sharon Stack, PhD Professor & Vice Chair for Research Mulligan Professor of Cancer Research Department of Pathology and Anatomical Sciences 1 Hospital Dr., Room M214C Medical Sciences Bldg Columbia, MO 65212 USA stackm@health.missouri.edu; stackm@missouri.edu

David A. Fishman, MD Professor and Director Department of Obstetrics and Gynecology New York University School of Medicine New York, New York 10016 fishmd01@med.nyu.edu

Series Editor Steven T. Rosen, MD Robert H. Lurie Comprehensive Cancer Center Northwestern University Chicago, Illinois USA

ISSN 0927-3042 ISBN 978-0-387-98093-5 DOI 10.1007/978-0-387-98094-2 Springer New York Dordrecht Heidelberg London

Library of Congress Control Number: 2008943691

#### © Springer Science+Business Media, LLC 2001, 2009

All rights reserved. This work may not be translated or copied in whole or in part without the written permission of the publisher (Springer Science+Business Media, LLC, 233 Spring Street, New York, NY 10013, USA), except for brief excerpts in connection with reviews or scholarly analysis. Use in connection with any form of information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed is forbidden. The use in this publication of trade names, trademarks, service marks, and similar terms, even if they are not identified as such, is not to be taken as an expression of opinion as to whether or not they are subject to proprietary rights.

While the advice and information in this book are believed to be true and accurate at the date of going to press, neither the authors nor the editors nor the publisher can accept any legal responsibility for any errors or omissions that may be made. The publisher makes no warranty, express or implied, with respect to the material contained herein.

Printed on acid-free paper

Springer is part of Springer Science+Business Media (www.springer.com)

As we struggle to understand, treat, and cure this deadly disease, this book is dedicated to the women who suffer with ovarian cancer and those who love them.

## Preface

This volume provides a set of comprehensive reviews from experts in the field on key clinical, translational, and basic research issues in ovarian cancer for clinicians and scientists. Chemoprevention, staging, and novel therapeutic targets are addressed in Part I with a series of reviews highlighting prevention strategies, surgical treatments, and translation of novel targets into clinical practice. Part II is focused on tumorigenesis and biomarkers. Reviews highlight genetic and epigenetic changes active in transformation of ovarian surface epithelium and biomarkers currently under investigation as diagnostic/ prognostic indicators or therapeutic targets. Part III includes comprehensive overviews of tumor progression, metastasis, and translational research models. These reviews evaluate key signal transduction pathways in ovarian cancer, describe the novel adhesive microenvironment unique to ovarian tumors, and provide a comprehensive description of in vitro organotypic and in vivo murine models used to study ovarian cancer onset, progression, and metastasis.

Columbia, Missouri New York, NY M.S. Stack D.A. Fishman

## Contents

Par	t I Chemoprevention, Staging, and Novel Therapeutic Targets	
1	Chemoprevention of Ovarian Cancer Anna Hoekstra and Gustavo C. Rodriguez	3
2	Staging and Surgical Treatment Mario M. Leitao Jr., and Richard R. Barakat	35
3	Novel Therapeutic Targets	63
4	Biomarker Targets and Novel Therapeutics	85
Par	t II Tumorigenesis and Biomarkers	
5	Tumor Suppressor Genes Zhen Lu and Robert C. Bast, Jr.	109
6	<b>Epigenetics and Ovarian Cancer</b> Kenneth P. Nephew, Curt Balch, Shu Zhang, and Tim H-M. Huang	131
7	Aberrant Epithelial Differentiation in Ovarian Cancer Elizabeth R. Smith, Kathy Qi Cai, Callinice D. Capo-chichi, and Xiang-Xi Xu	147
8	The Human Kallikrein Gene Family: New Biomarkers for Ovarian Cancer	165

Contents

9	Soluble Epidermal Growth Factor Receptor: A Biomarker of Epithelial Ovarian Cancer Andre T. Baron, Jacqueline M. Lafky, Cecelia H. Boardman, Elsa M. Cora, Marites C. Buenafe, Dachao Liu, Alfred Rademaker, David A. Fishman, Karl C. Podratz, Jill L. Reiter, and Nita J. Maihle	189
10	Activated Epidermal Growth Factor Receptor in Ovarian Cancer Laurie G. Hudson, Reema Zeineldin, Melina Silberberg, and M. Sharon Stack	203
Part	t III Tumor Progression, Metastasis, Research Models	
11	<b>Ras-Superfamily GTP-ases in Ovarian Cancer</b>	229
12	Lipid Generation and Signaling in Ovarian Cancer Yan Xu, Dongmei Wang, and Zeneng Wang	241
13	Lysophosphatidic Acid and Invasion Fengqiang Wang and David A. Fishman	269
14	<b>Cell Adhesion in Ovarian Cancer</b>	297
15	Microenvironmental Regulation of Ovarian Cancer Metastasis Maria V. Barbolina, Natalie M. Moss, Suzanne D. Westfall, Yueying Liu, Rebecca J. Burkhalter, Francoise Marga, Gabor Forgacs, Laurie G. Hudson, and M. Sharon Stack	319
16	Organotypic Models of Metastasis: A Three-dimensional Culture Mimicking the Human Peritoneum and Omentum for the Study of the Early Steps of Ovarian Cancer Metastasis	335
17	Animal Models of Ovarian Cancer Denise C. Connolly	353
Inde	x	393

х

## Contributors

**Roshan Agarwal** Department of System Biology, M.D. Anderson Cancer Center, University of Texas, Houston, TX, USA

**Curt Balch** Medical Sciences, Indiana University, Bloomington; Indiana University Cancer Center, Indianapolis, IN 46202, USA

Richard R. Barakat Department of Surgery, Division of Gynecology, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10065, USA

Maria V. Barbolina Department of Biopharmaceutical Sciences, University of Illinois, Chicago, IL, USA

Andre T. Baron Division of Hematology/Oncology, Department of Internal Medicine, Lucille P. Markey Cancer Center, University of Kentucky, 800 Rose Street, Lexington, KY, 40536-0093, USA, a.baron@uky.edu

**Robert C. Bast** Department of Experimental Therapeutics, Harry Carrothers Wiess Distinguished University Chair in Cancer Research, M.D. Anderson Cancer Center, University of Texas, Houston, TX 77030-4009, USA, rbast@mdanderson.org

Michael J. Birrer Professor of Medicine, Harvard Medical School, Director, Gynecologic Cancer Research Program, Harvard Cancer Center, Director, Gynecologic Medical Oncology, Yawkey 9, Massachusetts General Hospital, 55 Fruit Street, Boston MA 02114, mbirrer@partners.org

**Cecelia H. Boardman** Department of Obstetrics and Gynecology, Medical College of Virginia, Virginia Commonwealth University Health System, 1101 East Marshall Street, Richmond, VA 23298-0034, USA

Marites C. Buenafe Department of Family Practice, Lexington Clinic, 1221 South Broadway, Lexington, KY 40504, USA

**Rebecca J. Burkhalter** Departments of Pathology & Anatomical Sciences, University of Missouri School of Medicine, Columbia, MO, USA

Kathy Qi Cai Ovarian Cancer Programs, Fox Chase Cancer Center, Philadelphia, PA 19111, USA

**Callinice D. Capo-chichi** Department of Medicine and Department of Obstetrics and Gynecology, Sylvester Comprehensive Cancer Center, University of Miami School of Medicine, Miami, FL 33136, USA

Giovanna Casagrande Medical Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD 20892, USA

Setsuko K. Chambers Division of Women's Cancers, Arizona Cancer Center, The University of Arizona, Tucson, AZ, USA, Schambers@azcc.arizona.edu

**Kwai Wa Cheng** Department of System Biology, M.D. Anderson Cancer Center, University of Texas, Houston, TX, USA

Mary Clouser Arizona Cancer Center, The University of Arizona, Tucson, AZ, USA

Denise C. Connolly Fox Chase Cancer Center, Philadelphia, PA, USA

**Elsa M. Cora** Department of Biochemistry and Nutrition, University of Puerto Rico, Medical Sciences Campus, School of Medicine, San Juan, PR 00936-5067, USA

**Eleftherios P. Diamandis** Mount Sinai Hospital, University Health Network and Toronto Medical Laboratories; Division of Clinical Biochemistry, Department of Laboratory Medicine & Pathobiology, University of Toronto, ON, Canada, eddiamandis@mtsinai.on.ca

**Songuel Dogan** Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science, University of Chicago, Chicago, IL, 60637, USA

**Wafic M. ElMasri** Medical Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD 20892, USA

John Farley Department of Obstetrics and Gynecology, Uniformed Services University of the Health Sciences, 4301 Jones Bridge Road, Bethesda, MD 20814; Christina Annunziata Center for Cancer Research, National Cancer Institute, Associate Clinical Investigator, Medical Oncology Branch, Building 10, Room 12N226, 10 Center Drive, Bethesda, MD 20892-1906, USA

David A. Fishman Department of Obstetrics and Gynecology, New York University School of Medicine, 550 First Avenue, TH 528, New York, NY 10016, USA, david.fishman@nyumc.org

**Gabor Forgacs** Departments of Physics; Biological Sciences; Biomedical Engineering, University of Missouri School of Medicine, Columbia, MO, USA

Lisa M. Hess Science Officer, Arizona Cancer Center, The University of Arizona, Tucson, AZ, USA

Anna Hoekstra Director, Division of Gynecologic Oncology, Northshore University Health Systems, Suite 1507 Walgreen Building, Evanston Hospital, 2650 Ridge Ave., Evanston, IL 60201, USA

**Ebony Hoskins** Medical Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD 20892, USA

**Tim H-M. Huang** Division of Human Cancer Genetics, Comprehensive Cancer Center, The Ohio State University, Columbus, OH 43210, USA

Laurie G. Hudson Department of Pharmaceutical Sciences, College of Pharmacy, University of New Mexico, Albuquerque, NM 87131-0001, USA, lhudson@salud.unm.edu

Hilary A. Kenny Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science, University of Chicago, Chicago, IL, 60637, USA

**Daniel Kimm** Medical Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD 20892, USA

Elise C. Kohn Medical Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD 20892, USA

**Thomas Krausz** Department of Surgical Pathology, University of Chicago, Chicago, IL, 60637, USA

Jacqueline M. Lafky Tumor Biology Program, Mayo Clinic–Rochester, 200 First Street S.W., Rochester, MN 55905, USA

Mario M. Leitao Department of Surgery, Division of Gynecology, Memorial Sloan-Kettering Cancer Center, 1275 York Avenue, New York, NY 10065, USA

**Ernst Lengyel** Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science; Committee on Cancer Biology, University of Chicago, Chicago, IL 60637, USA, elengyel@uchicago.edu

**Dachao Liu** Department of Preventive Medicine, Robert H. Lurie Comprehensive Cancer Center, Northwestern University's Feinberg School of Medicine, 680 North Lake Shore Drive, Suite 1102, Chicago, IL 60611, USA

Yueying Liu Departments of Pathology & Anatomical Sciences, University of Missouri School of Medicine, Columbia, MO, USA

**Zhen Lu** Department of Experimental Therapeutics, M.D. Anderson Cancer Center, University of Texas, Houston, TX 77030-4009, USA

**Nita J. Maihle** Department of Obstetrics/Gynecology and Reproductive Sciences, Yale University School of Medicine, 300 George Street, Suite 8100, New Haven, CT 06511, USA **Francoise Marga** Departments of Physics, University of Missouri School of Medicine, Columbia, MO, USA

Gordon B. Mills Department of System Biology, M.D. Anderson Cancer Center, University of Texas, Houston, TX, USA, gmills @mdanderson.org

Anirban K. Mitra Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science, University of Chicago, Chicago, IL, 60637, USA

Natalie M. Moss Department of Cell & Molecular Biology, Northwestern University, Chicago, IL, USA

**Kenneth P. Nephew** Medical Sciences, Indiana University, Bloomington; Indiana University Cancer Center, Indianapolis, IN 46202, USA

**Karl C. Podratz** Department of Gynecologic Surgery/Oncology, Mayo Clinic–Rochester, 200 First Street S.W., Rochester, MN 55905, USA

Alfred Rademaker Department of Preventive Medicine, Robert H. Lurie Comprehensive Cancer Center, Northwestern University's Feinberg School of Medicine, 680 North Lake Shore Drive, Suite 1102, Chicago, IL 60611, USA

**Jill L. Reiter** Department of Obstetrics/Gynecology and Reproductive Sciences, Yale University School of Medicine, 300 George Street, Suite 8100, New Haven, CT 06511, USA

**Gustavo C. Rodriguez** Director, Division of Gynecologic Oncology, NorthShore University HealthSystem, Suite 1507 Walgreen Building, Evanston Hospital, 2650 Ridge Ave., Evanston, IL 60201, USA, grodriguez@northshore.org

**Melina Silberberg** Department of Pharmaceutical Sciences, College of Pharmacy, University of New Mexico, Albuquerque, NM, USA

**Elizabeth R. Smith** Department of Medicine, and Department of Obstetrics and Gynecology, Sylvester Comprehensive Cancer Center, University of Miami School of Medicine, Miami, FL 33136, USA

M. Sharon Stack Professor & Vice Chair for Research, Mulligan Professor of Cancer Research, Department of Pathology & Anatomical Sciences; 1 Hospital Dr., Room M214C Medical Sciences Bldg., Columbia, MO 65212, USA, stackm@missouri.edu; stackm@health.missouri.edu

**Dongmei Wang** Department of Obstetrics and Gynecology, Indiana University, 975 W. Walnut St. IB355A, Indianapolis, IN 46202, USA

Fengqiang Wang Department of Obstetrics Gynecology and Reproductive Sciences, Mount Sinai School of Medicine, New York University, 1176 fifth Avenue, Box 1170, New York, NY 10029, USA, Fengqiang.wang@mssm.edu **Zeneng Wang** Department of Cancer Biology, The Lerner Research Institute, Cleveland Clinic, 9500 Euclid Avenue, Cleveland, OH 44195, USA

Suzy D. Westfall Departments of Pathology & Anatomical Sciences, University of Missouri School of Medicine, Columbia, MO, USA

Xiang-Xi Xu Department of Medicine and Department of Obstetrics and Gynecology, UM/Sylvester Comprehensive Cancer Center, University of Miami School of Medicine, Miami, FL 33136, USA, Xxu2@med.miami.edu

**Yan Xu** Department of Obstetrics and Gynecology, Indiana University, 975 W. Walnut St. IB355A, Indianapolis, IN 46202, USA, xu2@iupui.com

**S. Diane Yamada** Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science, University of Chicago, Chicago, IL, 60637, USA

George M. Yousef Department of Laboratory Medicine and the Li Ka Shing Knowledge Institute, St. Michael's Hospital, 30 Bond Street, Toronto, ON, Canada, M5B 1W8, yousefg@smh.toronto.on.ca

**Reema Zeineldin** Department of Pharmaceutical Sciences, College of Pharmacy, University of New Mexico, Albuquerque, NM, USA

**Shu Zhang** Medical Sciences, Indiana University, Bloomington, IN, 47405, USA; Department of Obstetrics and Gynecology, Ren Ji Hospital, Shanghai JiaoTong, University School of Medicine, Shanghai, 200001, China

**Marion Zillhardt** Department of Obstetrics and Gynecology/Section of Gynecologic Oncology, Center for Integrative Science, University of Chicago, Chicago, IL, 60637, USA

## **Chapter 8 The Human Kallikrein Gene Family: New Biomarkers for Ovarian Cancer**

George M. Yousef and Eleftherios P. Diamandis

### The Human Kallikrein Gene Family

#### Structure and Genomic Organization

The term "kallikrein" (derived from the Greek *kallikreas*, for pancreas) was introduced in the 1930s to describe proteolytic enzymes that can release small vasoactive peptides from high-molecular-weight precursors. There are two categories of human kallikreins; the plasma and the tissue kallikreins. The plasma kallikrein is encoded by a single gene on chromosome 4. This enzyme (a serine protease) releases the vasoactive peptide bradykinin from a high-molecular-weight precursor synthesized in the liver.<sup>1</sup> The human tissue kallikrein family is localized on chromosome 19 and also encodes for serine protease enzymes.<sup>2–4</sup>

Recently, a new classification emerged for tissue kallikreins that is not based on the functional definition but rather on structural criteria and map location. Based on the newer definition, the number of genes that are included in this family increased to 15, a number that is comparable with that of homologous families found in rat and mouse.<sup>5,6</sup> Because all kallikreins (except KLK1) do not have classic "kallikrein" activity, they are better defined as "kallikreinrelated peptidases." A list of the official names of all kallikrein genes and proteins is included in Table 8.1, and a schematic diagram showing the human tissue kallikrein genes map within an approximately 300-kb region, and the lengths of the genes, the distances between them, as well as the direction of transcription have now been accurately defined.<sup>7,8</sup> The kallikrein family is bounded from the telomeric side by the Siglec family of genes<sup>9</sup> and

E.P. Diamandis  $(\boxtimes)$ 

Mount Sinai Hospital, University Health Network and Toronto Medical Laboratories; Division of Clinical Biochemistry, Department of Laboratory Medicine & Pathobiology, University of Toronto, ON, Canada

e-mail: eddiamandis@mtsinai.on.ca

Official Gene	Other Names/Symbols	GenBank Accession	UniGene	OMIM	SwissProt
Symbol	Other Names/Symbols	INUITOET	Cluster	ID	ID
KLK1	Pancreatic/renal	M25629	Hs.123107	147910	Q07276
	kallikrein, hPRK	M33105			
KLK2	Kallikrein-related	M18157	Hs.181350	147960	P20151
	peptidase 2				
	Human glandular				
	kallikrein I,				
VIV2	hGK-l Kallikasin salatad	V14010	Ha 171005	176920	D07200
KLKJ	pentidase 3	M24542	115.1/1995	1/0620	10/288
	Prostate specific antigen	M24343			
	PSA APS	1412/2/4			
KLK4	Kallikrein-related	AF113141	Hs.218366	603767	O9Y5K2
	peptidase 4	AF135023			
	Prostase, KLK-L1.	AF148532			
	EMSP1, PRSS17,				
	ARM1				
KLK5	Kallikrein-related	AF135028	Hs.50915	605643	Q9Y337
	peptidase 5	AF168768			
	KLK-L2, HSCTE				
KLK6	Kallikrein-related	AF013988	Hs.79361	602652	Q92876
	peptidase 6	AF149289			
	Zyme, Protease M,	U62801			
	Neurosin, PRSS9	D78203	11 151054	(0.1.120	<b>D</b> 40066
KLK/	Kallikrein-related	L33404	Hs.151254	604438	P49862
	peptidase /	AF166330			
VIVO	HSCCE, PRSS6	A D000840	Ha 104570	605614	060250
KLKð	nentidase 8	AB009849	HS.104570	003044	060239
	Nouropsin: Ovesin:	AF095745 A D010780			
	TADG-14 PRSS19	A E055982			
	HNP	111 055902			
KLK9	Kallikrein-related	AF135026	Hs.448942	605504	O9UKO9
	peptidase 9				
	KLK-L3				
KLK10	Kallikrein-related	AF055481	Hs.69423	602673	O43240
	peptidase 10	NM 002776			
	NES1, PSSSL1	-			
KLK11	Kallikrein-related	AB012917	Hs.57771	604434	Q9UBX7
	peptidase 11				
	TLSP/Hippostasin,				
	PRSS20				
KLK12	Kallikrein-related	AF135025	Hs.159679	605539	Q9UKR0
	peptidase 12				
VI V12	KLK-L5 Kallilansin nalatad	4 1125024	II- 1(520)	(05505	OOLUK D 2
KLK13	Kallikrein-related	AF135024	HS.165296	605505	Q9UKR3
VIVIA	KLK-L4 Kallikrain related	A E161221	Ha 282025	606125	000003
KLK14	pentidase 14	AI 101221	115.203723	000155	Q71003
	KIK-I6 protein				
KLK15	Kallikrein-related	AF303046	Hs 250770	610601	O9H2R5
	peptidase 15		10.200770	010001	27112103
	Prostinogen, HSRNASPH	[			

**Table 8.1** Official and other gene and protein names for members of the human kallikrein gene family

OMIM, Online Mendelian Inheritance in Man.



**Fig. 8.1** Schematic presentation of the human kallikrein locus on chromosome 19q13.4. Gene locations are indicated by arrowheads that show the direction of transcription. Gene lengths are shown in kilobases. Position of the location of expanded regions in the mouse and rat genome that contain closely related *KLK1* paralogs is indicated. Non-kallikrein genes are shaded in gray

centromerically by the testicular acid phosphatase gene (ACPT).<sup>10</sup> New, uniform nomenclatures are now established for the human tissue kallikreins and their rodent orthologs.<sup>11,12</sup> There are many common structural features of the human kallikrein genes and proteins.<sup>13</sup> All genes are formed of five coding exons, and most of them have one or more extra 5' untranslated exons. The first coding exon always contains a 5' untranslated region, followed by the methionine start codon, located  $\sim$  50 bp away from the end of the exon. The stop codon is always located  $\sim$ 156 bp from the beginning of the last coding exon. Moreover, exon sizes are nearly identical, and the positions of the residues of the catalytic triad of serine proteases are conserved. All kallikrein proteins are synthesized as a pre/pro peptides with a signal peptide of about 17–20 amino acids at the amino terminus, followed by an activation peptide of about 4-9 amino acids (with the exception of KLK5), followed by the mature (enzymatically active) protein. Finally, all proteins contain 10-12 cysteine residues that will form 5–6 disulfide bonds. The position of the cysteine residues is also fully conserved.

#### Tissue Expression and Hormonal Regulation

Many kallikreins are transcribed predominately in few tissues, as indicated by Northern blotting. By using the more sensitive RT-PCR technique, kallikreins are found to be expressed at lower amounts in several other tissues. The tissue expression of kallikrein mRNAs and proteins is summarized elsewhere.<sup>14</sup> Kallikrein abundance can be categorized as highly restricted (KLK2 and KLK3 in prostate), restricted (KLK5 in skin, salivary gland, breast, and esophagus; KLK6 in brain and central nervous system; KLK7 in esophagus, heart, liver, and skin; KLK8 in breast, esophagus, skin, and tonsil; KLK13 in esophagus and tonsil), or wide (KLK1, 4, 9–12, 14, and 15). Interestingly, many kallikreins are expressed in endocrine-related organs, including the prostate, testis, ovary, and breast.<sup>15</sup>

In the ovary, there is abundant expression of the mRNA of KLK6–8 and KLK10, followed by lower levels of KLK1, 9, 11, 14, and 15. At the protein level, KLK1, 6, 7, 10–11 show highest expression levels, followed by KLK8 and KLK14.<sup>14</sup> Given the coexpression of many kallikreins in the same tissue, it is possible that these kallikreins may act in concert in cascade pathways, reminiscent of the coagulation and apoptotic processes.

Several reports confirmed that many kallikreins are under steroid hormone regulation.<sup>16–18</sup> An interesting observation is the different patterns of hormonal regulation in different tissues (e.g., KLK4 is upregulated by androgen in prostate and breast cancer cell lines and by estrogen in endometrial cancer cell lines).

#### Kallikreins in Normal Physiology

From a functional point of view, kallikreins are serine proteases (SPs). SPs are peptidases with a uniquely activated serine residue in the substrate-binding pocket. They are involved in many vital functions such as digestion, blood clotting, fibrinolysis, fertilization, and complement activation and are related to many diseases including cancer, arthritis, and emphysema.<sup>19</sup>

Accumulating evidence indicates that kallikreins might have diverse functions in different tissues and developmental stages. KLK1 has a known role in blood pressure regulation by cleaving low-molecular-weight kininogen to produce vasoactive kinin peptides. Intact kinin binds to bradykinin B<sub>2</sub> receptor in target tissues and exerts a broad spectrum of biological effects including blood pressure reduction via vasodilation, smooth muscle relaxation or contraction, pain induction, and mediation of the inflammatory response.<sup>19</sup> Low renal synthesis and urinary excretion of tissue kallikreins have been linked to hypertension in animals and humans.<sup>20</sup> Apart from its kininogenase activity, KLK1 has been implicated in the processing of growth factors and peptide hormones in light of its presence in pituitary, pancreas, and other tissues. As summarized by Bhoola et al.,<sup>19</sup> KLK1 has been shown to cleave proinsulin, low-density lipoprotein, prorenin, angiotensinogen, vasoactive intestinal peptide, procollagenase, and the precursor of atrial natriuretic factor.

KLK3 (also known as prostate specific antigen; PSA) has been shown to rapidly hydrolyze semenogelin I and semenogelin II, as well as fibronectin, resulting in liquefaction of the seminal clot after ejaculation.<sup>21</sup> Several other potential substrates for KLK3 have been identified, including IGFBP-3, TGF- $\beta$ , parathyroid hormone-related peptide, and plasminogen.<sup>22</sup> KLK2 is found to be able to cleave semenogelin I and semenogelin II but at different cleavage sites and with lower efficiency than that of KLK3.<sup>23</sup> The mouse and porcine orthologs of KLK4 were originally designated "enamel matrix serine proteases" because of their predicted role in normal teeth development.<sup>24</sup> Recent evidence shows that a splice variant of kallikrein 4 is a predominately nuclear protein that might have a role in controlling gene expression.<sup>25</sup>

A few kallikreins, especially KLK5 and KLK7, are expressed in the stratum corneum of the skin and are known to be involved in desquamation of corneocytes.<sup>26</sup> Another group of kallikreins, KLK6, KLK8, and KLK11, are highly expressed in the central nervous system where they are thought to play a role in neural plasticity.<sup>27</sup> Another possible mechanism for kallikrein action is the activation of proteinase-activated receptors (PARs). Activation of these receptors elicits different responses in several tissues. In addition, they switch-on cell signaling pathways (e.g., the MAP kinase pathway), leading to cell growth and division.<sup>28</sup>

#### **Regulation of Kallikrein Activity**

Kallikrein activity is controlled at both the mRNA and protein levels. Besides KLK3 and KLK2, and more recently KLK10, no other kallikrein gene promoter has been functionally tested. TATA box variants are found in the three classic kallikreins (KLK1-3).<sup>29</sup> Also, androgen response elements have been identified and experimentally verified.<sup>30</sup> No obvious TATA boxes are found in the promoter of other kallikreins. At the protein level, there are different mechanisms for controlling serine protease activity by which unwanted activation is avoided and precise spatial and temporal regulation of the proteolytic activity is achieved. One important mechanism is by producing kallikreins in an inactive "proenzyme" (or zymogen) form, which is activated as necessary. The N-terminal extension of the mature enzyme, or the "prosegment," sterically blocks the active site and thus prevents binding of substrates. The activation of the zymogen can occur intracellularly (i.e., in the trans-Golgi apparatus or in the secretory granules) or extracellularly after secretion, and it can be autolytic or dependent on the activity of another enzyme. Autoactivation is a common phenomenon among kallikreins. KLK2, but not KLK3, is capable of autoactivation.<sup>31</sup> KLK4 is also autoactivated during the refolding process, and there is evidence that KLK6 is also capable of autoactivation.<sup>32</sup>

Proteolytic activation is irreversible. Hence, other means of switching off the activity of these enzymes are needed. Once activated, serine proteases are controlled by ubiquitous endogenous inhibitors.<sup>33</sup> Some molecular complexes of kallikreins with protease inhibitors have clinical applicability because they can improve the diagnostic sensitivity or specificity of cancer biomarkers such as PSA.<sup>34</sup>

The coexpression of many kallikreins in the same tissues and the parallel differential regulation of groups of kallikreins in pathologic conditions raise the possibility of the existence of a common mechanism that controls expression of groups of kallikrein genes in a cluster, as a "locus control region." Added to this are the relatively short distances between adjacent kallikreins (which could be as short as the 1.5 kb between *KLK1* and *KLK15*) and the absence of classic promoter sequences, as shown by prediction analysis, in all kallikreins except *KLK1–3*.

Kallikreins can be also targeted by microRNAs (miRNAs). The first bioinformatic prediction of the potential interaction between miRNAs and kallikreins with experimental verification has been recently published.<sup>35</sup> miRNAs represent an important tool of posttranscriptional regulation of kallikrein activity that can explain aberrancies between the mRNA and protein expression levels.<sup>36</sup>

#### Kallikrein Expression in Cancer

#### Kallikreins as Cancer Biomarkers

Accumulating evidence indicates that many kallikreins are differentially expressed in various malignancies. KLK6 (zyme/protease M) was originally isolated by differential display from an ovarian cancer library,<sup>37</sup> and KLK10 was cloned by subtractive hybridization from a breast cancer library<sup>38</sup> and later proved to act as a tumor suppressor gene.<sup>39</sup>

A number of kallikreins were shown to be putative prognostic and/or predictive cancer markers. In breast cancer, the expression of *KLK5* and *KLK14* is indicative of poor patient prognosis,<sup>40</sup> whereas higher levels of *KLK9*, *KLK13*, and *KLK15* mRNA and the KLK3 protein forecast a favorable disease outcome.<sup>41</sup> The apparent relationship between kallikreins and testicular cancer has been published,<sup>42</sup> and the differential expression of *KLK10*, *KLK14*, and *KLK13* splice variants in testicular cancer tissues have also been reported.<sup>43</sup>

A microarray study has identified at least one kallikrein (*KLK11*) is overexpressed in lung carcinoma.<sup>44</sup> Recently, *in silico* analysis provided evidence that some kallikreins are differentially regulated in pancreatic cancer.<sup>45</sup> This was confirmed by microarray analysis.<sup>46</sup> Recent evidence also indicates overexpression of three kallikreins (*KLK7*, *KLK8*, and *KLK10*) in colon cancer.<sup>45</sup> Another report showed downregulation of the KLK10 gene in acute lymphoblastic leukemia.<sup>47</sup>

The potential clinical utility of kallikreins as cancer biomarkers has been proved by many reports. Prostate-specific antigen (KLK3) and, more recently, human glandular kallikrein (KLK2) are useful biomarkers for prostate cancer.<sup>48</sup> KLK11 is also shown to be a potential marker for ovarian and prostate cancer.<sup>49</sup> Recent reports demonstrate that kallikrein mRNA and proteins can be useful serum biomarkers for diagnosis, monitoring, and prognosis of different cancers.<sup>15</sup> In addition to their diagnostic/prognostic utilities, kallikreins have potential for being used for therapeutic applications. A synthetic KLK1 inhibitor is found to suppress cancer cell invasiveness in human breast cancer cell lines.<sup>50</sup>

An interesting observation is that many kallikreins were found to be dysregulated in malignancies of different tissues, for example, KLK5 in ovarian and breast cancer<sup>51</sup> and the downregulation of *KLK14* in multiple malignancies.<sup>52</sup> This lack of "tissue specificity" points to the possibility that kallikreins might be involved in a "common" pathway or biological process that is involved in cancer initiation and/or progression.

### Differential Expression of Kallikreins in Ovarian Cancer

The dysregulation of kallikreins in ovarian cancer is well documented. KLK6 was isolated by differential display from an ovarian cancer library.<sup>37</sup> Kallikreins were identified among the top differentially expressed genes in ovarian cancer in a global analysis.<sup>53</sup> Recently, an *in silico* analysis of kallikrein gene expression in ovarian cancer was performed by using the databases of the Human Genome Anatomy Project. This study showed that at least seven kallikreins are upregulated in ovarian cancer compared with that in normal ovarian tissues. This was also confirmed at the protein level.<sup>54</sup> A review showing the prognostic value of many members of the human kallikrein family in ovarian cancer has been also published.<sup>18</sup>

#### Subcellular Localization of Kallikreins in Ovarian Cancer

Immunohistochemistry (IHC) enables kallikrein protein distribution in different cell types, independently from its quantity in the tissue. In addition, it provides a semiquantitative analysis of expression levels. Because kallikreins are secreted proteins, it was not unexpected that immunostaining of kallikreins was mainly cytoplasmic and in some tissues displayed a characteristic pattern that was membranous, droplet-like, supranuclear, subnuclear, or luminal. KLK4 appears to be a notable exception. Recently, Xi et al. suggested that one variant of KLK4 is a predominately nuclear protein that is overexpressed in prostate cancer.<sup>25</sup> Many kallikreins were analyzed by IHC and showed upregulation in ovarian cancer compared with that in normal ovarian tissues. KLK4 is localized to the cytoplasm of ovarian cancer, but not normal cells, with focal membranous staining.<sup>55</sup> KLK10 and KLK14 are found to have an intracytoplasmic pattern of staining in the epithelial cells (and occasional stromal cells) of serous ovarian cancers. 56,57 Underwood et al., using peptide antibody against the KLK8 protein, showed cytoplasmic granular staining (that might represent a secretion pathway) in tumor cells of different histologic types. In endometrioid carcinoma, the staining was most prominent in the glandular lumens.<sup>58</sup> KLK9 shows moderate cytoplasmic staining, with no nuclear or stromal staining pattern in ovarian cancer cells.<sup>59</sup>

#### Kallikrein Splice Variants in Ovarian Cancer

The mechanism by which a single gene gives rise to more than one mRNA transcript is referred to as differential splicing. This system is often tightly regulated in a cell type–specific or developmental stage–specific manner and increases genome complexity by generating different proteins from the same mRNA. The presence of more than one mRNA form for the same gene is common among kallikreins. These variant mRNAs may result from alternative splicing, a retained intronic segment, or use of an alternative transcription

initiation site. To date, there are at least 82 documented splice variants of the 15 kallikrein genes.<sup>60</sup> A better understanding of alternative splicing can lead to the use of gene variants as drug targets, therapeutic agents, or diagnostic markers.<sup>60</sup> Slawin et al. reported a prognostic significance of a splice variant–specific RT-PCR assay for *KLK2* in detecting prostate cancer metastasis.<sup>61</sup> Nakamura et al. reported differential expression of the brain and prostate types of *KLK11* between benign, hyperplastic, and malignant prostate cancer cell lines.<sup>62</sup> Some of the alternatively spliced forms were also found to be tissue specific.

Several kallikrein splice variants were identified in ovarian cancer. Dong et al.<sup>55</sup> identified three alternative splice forms of KLK4 expressed in ovarian cancer tumor tissues and cell lines, but not in normal ovaries: one with intronic insertion from intron 3; the second has intronic insertion from intron 2 and exon 4 deletion, and the third has deleted exon 4. A novel *KLK5* mRNA transcript with a short 5' untranslated region and a *KLK7* splice variant with a long 3' untranslated region are highly expressed in ovarian cancer cell lines but are expressed in very low levels in normal ovarian epithelial cells.<sup>63</sup> Another splice variant was identified, *KLK5-SV2*, which is overexpressed in ovarian cancer tissues and cell lines.<sup>64</sup> A recent report showed that *KLK6-splice variant 1* is expressed at much higher levels in ovarian cancer compared with the "classic" variant.<sup>65</sup>

#### Mechanisms of Kallikrein Involvement in Ovarian Cancer

The mechanism by which kallikreins might be involved in the pathogenesis and/ or progression of ovarian cancer is not yet fully understood. Preliminary reports indicate a possible role of kallikreins in controlling vital processes, like apoptosis, angiogenesis, and tumor metastasis by cleavage of specific substrates, including growth factors, hormone receptors, or connective tissue. The involvement in growth and apoptotic activities was reported for KLK3 (PSA), which can digest insulin-like growth factor–binding protein (IGFBP-3)<sup>66</sup> and parathyroid hormone–related protein (PTHrP).

As proteolytic enzymes, kallikreins can be involved in tumor progression because of their role in extracellular matrix degradation. Many studies have shown that a variety of proteolytic enzymes are overproduced either by the cancer cells themselves or by the surrounding stromal cells, with an associated unfavorable clinical prognosis. Experimental evidence indicates that KLK2 and KLK4 can activate the proform of another serine protease, the urokinase-type plasminogen activator (uPA).<sup>67</sup> Urokinase activates plasmin from its inactive form (plasminogen), which is ubiquitously located in the extracellular space leading to degradation of the extracellular matrix proteins. Plasmin can also activate precursor forms of collagenases, thus promoting the degeneration of collagen in the basement membrane surrounding the capillaries and lymph nodes. Another kallikrein, KLK7, can degrade the alpha chain of human fibrinogen.<sup>68</sup>

Modulation of angiogenic activity is another possible mechanism for kallikrein involvement in cancer. The kinin family of vasoactive peptides, liberated by KLK1 action, is believed to regulate the angiogenic process.<sup>69</sup> It was recently reported that immunolabeling of KLK1 is intense in the angiogenic endothelial cells derived from mature corpora lutea.<sup>69</sup> Also, KLK3 is reported to have anti-angiogenic activities.<sup>70</sup>

A recent study has shown that expression of kallikreins increases the malignant behavior of ovarian cancer cells.<sup>71</sup> Transfecting cancer cells with kallikreins led to significantly increased invasive behavior, and when these cells were inoculated into the peritoneum of nude mice, they resulted in a remarkable increase in tumor burden.

Ovarian cancer is a "hormone-related" malignancy. Sex hormones are known to affect its initiation and/or progression.<sup>72</sup> Oral contraceptive pills decrease the risk of ovarian cancer,<sup>73</sup> and the growth of ovarian carcinoma cell lines is sensitive to estrogen.<sup>74</sup> Progesterone promotes cell differentiation and apoptosis, and it has been shown to inhibit DNA synthesis and cell division.<sup>75</sup> Also, studies have shown a prognostic value of the progesterone receptor in ovarian cancer.<sup>76</sup> Moreover, appreciable evidence implicates androgens in the pathogenesis of ovarian cancer<sup>77</sup> and supports the existence of a physiologic interaction between androgens and the ovarian surface epithelium, as well as the possible role of this interaction in ovarian neoplasia.<sup>78</sup> Androgens have also been shown to stimulate growth of rodent ovarian epithelial cells in vivo, leading to benign ovarian neoplasms.<sup>79</sup> Ovarian cancer patients have higher levels of circulating androgens than do women without cancer.<sup>80</sup> Additionally, the majority of ovarian cancers express androgen receptor (AR),<sup>81,82</sup> and ovarian cancer cell growth is inhibited in vitro by antiandrogens.<sup>83</sup> Recent observations show a correlation between AR and susceptibility to ovarian cancer.<sup>82</sup> Given the fact that most kallikreins are regulated by sex hormones,<sup>17</sup> kallikreins could represent downstream targets by which steroids are involved in the malignant process. This, however, could not be verified in a recent study.<sup>65</sup> The elevation of serum concentration of kallikreins in cancer could also be due to the increased vasculature (angiogenesis) of cancerous tissues and the destruction of the glandular architecture of the tissues involved, with subsequent leakage of these proteins into the general circulation.

#### Clinical Utility of Kallikreins in Ovarian Cancer

#### Kallikreins as Diagnostic Markers

The clinical utility of kallikreins in ovarian cancer spans both the diagnostic and prognostic applications. For diagnostic purposes, many kallikreins have been shown to be elevated, at both the mRNA and protein levels, in ovarian cancer compared with patients with normal ovaries. Kallikreins can be measured in serum, tissue, or ascites fluid.<sup>84</sup> Reports on the diagnostic value of kallikreins in ovarian cancer are summarized in Table 8.2. Among all kallikreins, KLK6 and KLK10 show the best promise as serum biomarkers for ovarian cancer, specially the serous type.

Type of Analysis	Kallikrein	Clinical Significance	References
Protein in serum	KLK6	<ul> <li>Serum levels elevated in 66% to 68% of cancer patients</li> <li>The diagnostic sensitivities at 90% and</li> </ul>	90, 92
		95% specificity are 52% and 47%, respectively	
	KLK10	• Serum levels elevated in cancer compared with normal and benign disease (54% sensitivity at 90% specificity)	89
		• Elevated in 35% of CA-125–negative cancers (at 90% specificity)	
Protein in serum and ovarian	KLK10	<ul> <li>Significantly associated with serous type</li> <li>A member of a multianalyte test for ovarian cancer diagnosis</li> </ul>	88
tissue	KLK14	• Elevated serum levels in 65% of ovarian cancer patients versus normal	57
		• Higher levels in 40% of ovarian cancer	
Protein in ovarian	KLK5	<ul> <li>Elevated in 55% of ovarian cancers compared with normal</li> </ul>	97
mRNA from	KLK4	<ul> <li>Elevated in 100% of serous carcinoma of late stage</li> </ul>	55
ovariari tissue	KLK6	<ul> <li>Significantly elevated in low-malignant- potential tumors and ovarian cancer</li> </ul>	98
	KLK5, KLK7	<ul> <li>Significantly elevated in ovarian cancer, especially serous type</li> </ul>	63
	KLK7	• Elevated in 67% of low-malignant-potential tumors and 78% of carcinomas	99
	KLK8	• Overexpressed in 67% of ovarian cancers and 40% of low-malignant-potential tumors compared with normal	58
	KLK10	<ul> <li>Significantly elevated in 91% of serous cancers, 73% of nonserous cancers, and 73% of primary peritoneal carcinoma compared with normal</li> </ul>	100
	KLK14	<ul> <li>Downregulated in ovarian cancer</li> <li>Stepwise decrease in normal &gt; benign &gt;</li> </ul>	52, 101
IHC of ovarian	KLK6 and KLK10	<ul><li>cancer</li><li>Expressed in 100% of CA-125–negative cancers</li></ul>	87
Protein from ascites fluid	KLK5–8, 10, 11, 13, 14	<ul> <li>Ovarian cancer ascites contained higher levels compared with benign effusions and ascites from other cancer types</li> </ul>	86
Microarray	KLK5–8	• Among the top upregulated genes in ovarian cancer compared with normal tissue and other diseases	102

 Table 8.2
 Kallikreins as diagnostic markers for ovarian cancer

		Table 8.2 (continued)	
	KLK6–8,	• Significantly upregulated in ovarian	103
	10, 11	cancer	
	KLK6–8,	<ul> <li>Overexpressed in ovarian cancer</li> </ul>	104
	10, 11		
Bioinformatics	KLK5–8,	• Parallel overexpression in ovarian cancer	54
analysis	10,11, 14	compared with normal	
-	KLK6	• Elevated 25-fold in ovarian cancer	53, 63

 Table 8.2 (continued)

Recent reports indicate the potential diagnostic utility of kallikreins in ascites fluid. Among kallikreins, KLK6–10 showed the highest statistical power in distinguishing ovarian cancer ascites from that of benign causes and other cancer groups. It was shown that kallikreins could identify false-negative cases of cytology.<sup>85,86</sup> Combinations of kallikreins achieved areas under the receiver operating characteristics (ROC) curve of 0.994 and 0.961 in separating ovarian cancer from benign effusion and from other cancer groups, respectively.<sup>86</sup>

The diagnostic utility of kallikreins can also extend to their use as immunohistochemical markers. A recent study has shown that in ovarian cancers that lacked CA125 expression by IHC, all specimens (100%) expressed KLK10 and KLK6.<sup>87</sup> Kallikrein mRNAs are also detected in circulating tumor cells in the blood and ascites fluid of ovarian cancer patients, but this application lacked sensitivity and specificity for detecting disseminated disease.<sup>85</sup>

Although the sensitivity and specificity of individual kallikrein proteins are not superior to that of standard markers, like CA-125, the use of kallikreins as a part of a multianalyte test significantly improves the diagnostic sensitivity and specificity.<sup>88</sup> In patients with early-stage cancer (stage I/II), the combination of CA-125 and KLK10 results in 21% increase in sensitivity compared with that of CA-125 alone.<sup>89</sup> The combination of KLK6 and CA-125 can also lead to improved sensitivity of detection of early-stage disease. When combined with CA-125, at 90% specificity, sensitivity increases to 72% (for all patients) and to 42% in stage I or II disease.<sup>90</sup> A recent review included kallikreins among the most promising new markers that are now being investigated to complement CA-125 for ovarian cancer diagnosis/ prognosis.<sup>91</sup>

Another interesting application is distinguishing benign and borderline ovarian tumors. Preliminary reports indicate that KLK10 and KLK6 can have better ability than CA-125 in distinguishing ovarian cancer from benign ovarian tumors.<sup>89,90</sup>

#### **Prognostic Applications**

Clinical utility of kallikreins in ovarian cancer extends beyond diagnosis. Table 8.3 summarizes published data about the prognostic utility of different kallikreins in ovarian cancer. These data show that a group of kallikreins,

cancer	
ovarian	
.u	
genes/proteins	
of kallikrein	
rognostic utility c	
P	
Table 8.3	

Kallikrein Gene/Protein	Sample Type	Analysis Method	Prognostic Value	References
KLK4	mRNA	RT-PCR	<ul> <li>Unfavorable prognosis:</li> <li>Overexpressed in late-stage, higher- grade tumors and no response to chemotherapy</li> <li>associated with shorter DFS and OS</li> <li>independent indicator of poor prognosis in patients with low-grade tumors</li> </ul>	105
	Protein from effusion cells and solid tumor	IHC and immunoblotting	<ul> <li>Favourable prognosis:</li> <li>lower in grade IV compared with grade III</li> <li>associated with longer OS</li> </ul>	106
KLKS	mRNA	RT-PCR	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in late-stage and higher- grade tumors</li> <li>associated with shorter DFS and OS</li> <li>independent indicator of poor prognosis in patients with low-grade tumors</li> </ul>	107
	mRNA and protein	SQ-RT-PCR, Southern, Northern, and Western blots and immunohistochemistry	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in ovarian tumor tissues and cell lines mainly of late stage and serous histotype</li> </ul>	63
	Ovarian cancer cytosols	Immunoassay	<ul> <li>Unifavorable prognosis:</li> <li>overexpressed in patients with late- stage and higher-grade tumors</li> <li>associated with shorter DFS and OS</li> </ul>	108

		Table 8.3 (continued)		
Kallikrein Gene/Protein	Sample Type	Analysis Method	Prognostic Value	References
			• independent indicator of poor prognosis in patients with high-grade tumors and optimal debulking success	
KLK6	Ovarian cancer cytosols	Immunoassay	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in late-stage and serous tumors</li> <li>associated with shorter DFS and OS</li> <li>independent indicator of poor prognosis in low-grade tumors and optimal debulking success</li> </ul>	109
	Serum from normal women, women with benign disease, and women with ovarian cancer	Immunoassay	<ul> <li>Unfavorable prognosis:</li> <li>higher serum levels in late-stage, higher-grade, serous tumors, suboptimal debulking, and a poor response to chemotherapy</li> <li>indicator of decreased DFS and OS</li> </ul>	06
KLK7	Ovarian cancer cytosols	Immunoassay	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in advanced stage, higher grade, suboptimal debulking, and serous or undifferentiated histotypes</li> <li>associated with significantly shorter DFS but not OS</li> <li>not an independent prognosticator for ovarian cancer</li> </ul>	110
	mRNA from normal, benign, and cancerous ovarian tissues and late-stage	SQ-RT-PCR, Southern, Northern, and Western blots and immunohistochemistry	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in ovarian tumor tissues and cell lines mainly of late-stage and serous histotype</li> </ul>	63

continu
8:3 8
Table

	References		111			112		113				114		59		
tinued)	Prognostic Value		Unfavorable prognosis:	<ul> <li>overexpressed in higher-grade tumors</li> <li>associated with shorter DFS</li> <li>independent indicator of noor</li> </ul>	prognosis in patients with low-grade tumors and optimal debulking success	Favorable prognosis:	<ul> <li>overexpressed in lower-grade tumors</li> <li>associated with longer DFS and OS</li> <li>independent indicator of longer DFS</li> </ul>	<i>Favorable</i> prognosis: • overextressed in lower grade no	residual tumor, and optimal debulking	<ul> <li>associated with significantly longer</li> <li>DFS and OS</li> </ul>	<ul> <li>an independent marker of favorable prognosis</li> </ul>	Favorable prognosis:	<ul> <li>overexpressed in younger age, lower- grade, and early-stage tumors</li> </ul>	Favorable prognosis:	• overexpressed in early stage and	<ul> <li>optimal debulking success</li> <li>associated with longer DFS and OS</li> </ul>
Table 8.3 (cont	Analysis Method		Q-RT-PCR			RT-PCR		Immunoassay				RT-PCR		Q-RT-PCR		
	Sample Type	serous ovarian cancer cell lines	mRNA from	cancerous ovarian tissue		mRNA from ovarian	cancer tissues	Protein from ovarian cancer cvtosols				mRNA from ovarian	cancer tissues	mRNA from ovarian	cancer tissues	
	Kallikrein Gene/Protein	~				KLK8								KLK9		

178

Kallikrein Gene/Protein	Sample Type	Analysis Method	Prognostic Value	References
			<ul> <li>independent indicator of prolonged DFS in patients with low-grade tumors and optimal debulking success</li> </ul>	
KLK10	Normal, benign, and cancerous ovarian cytosols	Immunoassay	<ul> <li>Unfavorable prognosis:</li> <li>overexpressed in cancer patients with late-stage, serous tumors and suboptimal debulking success</li> <li>associated with shorter DFS and OS</li> <li>independent indicator of DFS and OS</li> <li>in patients with late-stage tumors</li> </ul>	56
	Serum from normal women, women with benign disease, and women with ovarian cancer	Immunoassay	<ul> <li>Unfavorable prognosis:</li> <li>higher serum levels in late-stage, advanced-grade, large residual tumor, suboptimal debulking, and poor response to chemotherapy</li> <li>indicator of decreased DFS and OS</li> <li>independent indicator of OS</li> </ul>	88
KLKII	Proteins of ovarian tumor extracts	Immunoassay	<ul> <li><i>Favorable</i> prognosis:</li> <li>overexpressed in early-stage and low-grade tumors</li> <li>associated with longer DFS and OS</li> <li>independent indicator of DFS</li> </ul>	115
	Ovarian cancer cytosols	Immunoassay	<ul> <li><i>Favorable</i> prognosis:</li> <li>overexpressed in patients with early- stage disease, and good response to chemotherapy</li> <li>associated with longer DFS and OS</li> </ul>	116

Table 8.3 (continued)

Kallikrein Gene/Protein	Sample Type	Analysis Method	Prognostic Value	References
			<ul> <li>independent indicator of OS</li> <li>independent indicator of DFS and OS in patients with low-grade tumors</li> </ul>	
KLK13	Ovarian cancer cytosols	Immunoassay	<ul><li>Favorable prognosis:</li><li>overexpressed in early stage, no residual tumor after surgery, and</li></ul>	117
			<ul><li>optimal debulking success</li><li>independent indicator of longer DFS and OS</li></ul>	
KLK14	mRNA from normal, benign, and cancerous ovarian tissues	Q-RT-PCR	<ul> <li><i>Favorable</i> prognosis:</li> <li>overexpressed in early stage, optimal debulking success, and good response to chemotherapy</li> <li>independent indicator of longer DFS and OS</li> </ul>	101
KLK15	mRNA from benign and cancerous ovarian tissues	Q-RT-PCR	Unfavorable prognosis: <ul> <li>independent indicator of decreased</li> <li>DFS and OS</li> </ul>	118
IHC, immunohis PCR, semiquant:	tochemistry; RT-PCR, re itative RT-PCR; Q-RT-PC	verse transcriptase-polymerase chain reacti 2R, quantitative RT-PCR.	ion; DFS, disease-free survival; OS, overall sur	vival; SQ-RT-

Table 8.3 (continued)

namely *KLK4–7*, *KLK8*, *KLK10*, and *KLK15*, are markers of poor prognosis in ovarian cancer. That is, higher kallikrein mRNA and/or protein levels correlate with more aggressive forms of the disease and a decreased disease-free survival (DFS) and overall survival (OS) of patients. The remaining subset of kallikreins, namely *KLK8*, *KLK9*, *KLK11*, and *KLK13–14*, seem to be markers of favorable prognosis, with higher levels of their mRNA or proteins associated with earlier-stage disease and increased DFS and OS.

Data from a recent report suggested that KLK6 might have value for patient monitoring in ovarian cancer.<sup>92</sup> A recent pilot study shows that some kallikrein proteins could be included with other clinical variables to develop a multi-parametric "score" that can predict the surgical outcome and thus help in preoperative risk stratification and identifying candidates for alternative or adjuvant therapeutic strategies.<sup>93</sup> Another study showed evidence that a group of kallikreins and multiparametric combinations with other biomarkers and clinical variables can significantly assist with ovarian cancer classification, prognosis, and response to platinum-based chemotherapy.<sup>94</sup>

Kallikrein expression in ovarian cancer may also be clinically useful in determining the prognosis in subgroups of patients. Subclassification of large heterogeneous groups into smaller subgroups is becoming an important tool for individualizing treatment options in ovarian cancer patients and thus avoiding unnecessary treatments with high costs and unwanted side effects.

#### Therapeutic Applications

It is possible that some kallikreins may become valuable therapeutic targets when the biological pathways that are involved are delineated. For example, the enzymatic activity of these serine proteases may initiate or terminate biological events (e.g., tumor invasion, angiogenesis, activation or inhibition of hormones, growth factors, other enzymes, receptors, or cytokines). Once known, these events could be manipulated, for therapeutic purposes, by specific enzyme inhibitors or activators. Another potential therapeutic approach is the cell-specific activation of therapeutic agents.<sup>95</sup> Preliminary reports show potential success by using the KLK3 (PSA) promoter to express molecules in a tissue-specific fashion.<sup>96</sup> A third possible therapeutic approach involves immunotherapy and/or development of cancer vaccines. With our increasing knowledge of the hormonal regulation of kallikreins, hormonal activation (or repression) of kallikrein activity could be investigated in the future.

#### Conclusion

Accumulating evidence, at both the mRNA and protein levels, indicates that many kallikreins are differentially expressed in ovarian cancer. The mechanism by which kallikreins are involved in the pathogenesis and/or progression of

cancer is not fully understood and is possibly through controlling vital processes, like apoptosis, angiogenesis, and tumor metastasis, by cleavage of specific substrates. Kallikreins can be measured in serum, tissue, and ascites fluid of ovarian cancer patients. Reports show that kallikreins can be useful serum biomarkers for diagnostic, monitoring, and prognostic purposes. They can be also useful immunohistochemical markers. In addition, kallikreins have potential for being used for therapeutic applications. KLK6 and KLK10 show the best promise as serum biomarkers for ovarian cancer. Although kallikreins might not have superior sensitivity and specificity to that of existing markers, the use of kallikreins as a part of a multianalyte test significantly improves the diagnostic and prognostic accuracy. Further large-scale studies are needed to evaluate the applicability of this approach.

#### References

- 1. Clements J. The molecular biology of the kallikreins and their roles in inflammation. In: Farmer S, ed. *The Kinin System*. New York: Academic Press; 1997:71–97.
- Clements J, Hooper J, Dong Y, Harvey T. The expanded human kallikrein (KLK) gene family: genomic organisation, tissue-specific expression and potential functions. *Biol Chem.* 2001;382:5–14.
- Paliouras M, Borgono C, Diamandis EP. Human tissue kallikreins: the cancer biomarker family. *Cancer Lett*. 2007;249:61–79.
- 4. Yousef GM, Obiezu CV, Luo LY, et al. Human tissue kallikreins: from gene structure to function and clinical applications. *Adv Clin Chem.* 2005;39:11–79.
- Diamandis EP, Yousef GM, Olsson AY. An update on human and mouse glandular kallikreins. *Clin Biochem*. 2004;37:258–260.
- Olsson AY, Lilja H, Lundwall A. Taxon-specific evolution of glandular kallikrein genes and identification of a progenitor of prostate-specific antigen. *Genomics*. 2004;84: 147–156.
- Yousef GM, Chang A, Scorilas A, Diamandis EP. Genomic organization of the human kallikrein gene family on chromosome 19q13.3-q13.4. *Biochem Biophys Res Commun.* 2000;276:125–133.
- 8. Yousef GM, Diamandis EP. Human kallikreins: common structural features, sequence analysis and evolution. *Curr Genom*. 2003;4:147–165.
- Yousef GM, Ordon MH, Foussias G, Diamandis EP. Genomic organization of the siglec gene locus on chromosome 19q13.4 and cloning of two new siglec pseudogenes. *Gene*. 2002;286:259–270.
- Yousef GM, Diamandis M, Jung K, Diamandis EP. Molecular cloning of a novel human acid phosphatase gene (ACPT) that is highly expressed in the testis. *Genomics*. 2001; 74:385–395.
- 11. Diamandis EP, Yousef GM, Clements J, et al. New nomenclature for the human tissue kallikrein gene family. *Clin Chem.* 2000;46:1855–1858.
- 12. Lundwall A, Band V, Blaber M, et al. A comprehensive nomenclature for serine proteases with homology to tissue kallikreins. *Biol Chem.* 2006;387:637–641.
- Diamandis EP, Yousef GM. Human tissue kallikreins: a family of new cancer biomarkers. *Clin Chem.* 2002;48:1198–1205.
- 14. Shaw JL, Diamandis EP. Distribution of 15 human kallikreins in tissues and biological fluids. *Clin Chem.* 2007;53:1423–1432.

- 8 The Human Kallikrein Gene Family
- 15. Yousef GM, Obiezu CV, Luo LY, et al. Human tissue kallikreins: from gene structure to function and clinical applications. *Adv Clin Chem.* 2005;39:11–79.
- 16. Borgono CA, Michael IP, Diamandis EP. Human tissue kallikreins: physiologic roles and applications in cancer. *Mol Cancer Res.* 2004;2:257–280.
- 17. Paliouras M, Diamandis EP. Coordinated steroid hormone-dependent and independent expression of multiple kallikreins in breast cancer cell lines. *Breast Cancer Res Treat*. 2007;102:7–18.
- Yousef GM, Diamandis EP. Kallikreins, steroid hormones and ovarian cancer: is there a link? *Minerva Endocrinol*. 2002;27:157–166.
- Bhoola KD, Figueroa CD, Worthy K. Bioregulation of kinins: kallikreins, kininogens, and kininases. *Pharmacol Rev.* 1992;44:1–80.
- Meneton P, Bloch-Faure M, Hagege AA, et al. Cardiovascular abnormalities with normal blood pressure in tissue kallikrein-deficient mice. *Proc Natl Acad Sci U S A*. 2001;98:2634–2639.
- Lilja H. A kallikrein-like serine protease in prostatic fluid cleaves the predominant seminal vesicle protein. J Clin Invest. 1985;76:1899–1903.
- Heidtmann HH, Nettelbeck DM, Mingels A, Jager R, Welker HG, Kontermann RE. Generation of angiostatin-like fragments from plasminogen by prostate-specific antigen. *Br J Cancer*. 1999;81:1269–1273.
- Deperthes D, Frenette G, Brillard-Bourdet M, et al. Potential involvement of kallikrein hK2 in the hydrolysis of the human seminal vesicle proteins after ejaculation. J Androl. 1996;17:659–665.
- 24. Hu JC, Zhang C, Sun X, et al. Characterization of the mouse and human PRSS17 genes, their relationship to other serine proteases, and the expression of PRSS17 in developing mouse incisors. *Gene.* 2000;251:1–8.
- 25. Xi Z, Klokk TI, Korkmaz K, et al. Kallikrein 4 is a predominantly nuclear protein and is overexpressed in prostate cancer. *Cancer Res.* 2004;64:2365–2370.
- Komatsu N, Takata M, Otsuki N, et al. Expression and localization of tissue kallikrein mRNAs in human epidermis and appendages. *J Invest Dermatol.* 2003;121:542–549.
- 27. Yousef GM, Kishi T, Diamandis EP. Role of kallikrein enzymes in the central nervous system. *Clin Chim Acta*. 2003;329:1–8.
- Oikonomopoulou K, Hansen KK, Saifeddine M, et al. Proteinase-mediated cell signalling: targeting proteinase-activated receptors (PARs) by kallikreins and more. *Biol Chem.* 2006;387:677–685.
- 29. Riegman PH, Vlietstra RJ, Klaassen P, et al. The prostate-specific antigen gene and the human glandular kallikrein-1 gene are tandemly located on chromosome 19. *FEBS Lett*. 1989;247:123–126.
- Cleutjens KB, van Eekelen CC, van der Korput HA, Brinkmann AO, Trapman J. Two androgen response regions cooperate in steroid hormone regulated activity of the prostate-specific antigen promoter. *J Biol Chem.* 1996;271:6379–6388.
- Denmeade SR, Lovgren J, Khan SR, Lilja H, Isaacs JT. Activation of latent protease function of pro-hK2, but not pro-PSA, involves autoprocessing. *Prostate*. 2001;48: 122–126.
- 32. Little SP, Dixon EP, Norris F, et al. Zyme, a novel and potentially amyloidogenic enzyme cDNA isolated from Alzheimer's disease brain. *J Biol Chem*. 1997;272:25135–25142.
- 33. Laskowski M, Qasim MA. What can the structures of enzyme-inhibitor complexes tell us about the structures of enzyme substrate complexes? *Biochim Biophys Acta*. 2000;1477: 324–337.
- Becker C, Lilja H. Individual prostate-specific antigen (PSA) forms as prostate tumor markers. *Clin Chim Acta*. 1997;257:117–132.
- Chow TF, Crow M, El-Said H, Diamandis EP, Yousef GM. Kallikreins as microRNA targets: an in-silico and experimental-based analysis. *Biol Chem.* 2008 Jun; 389(6):731–738.
- 36. Yousef GM. microRNAs: a new frontier in kallikrein research. *Biol Chem.* 2008 Jun; 389(6):689–694.

- Anisowicz A, Sotiropoulou G, Stenman G, Mok SC, Sager R. A novel protease homolog differentially expressed in breast and ovarian cancer. *Mol Med.* 1996;2:624–636.
- Liu XL, Wazer DE, Watanabe K, Band V. Identification of a novel serine protease-like gene, the expression of which is down-regulated during breast cancer progression. *Cancer Res.* 1996;56:3371–3379.
- Goyal J, Smith KM, Cowan JM, Wazer DE, Lee SW, Band V. The role for NES1 serine protease as a novel tumor suppressor. *Cancer Res.* 1998;58:4782–4786.
- Yousef GM, Scorilas A, Kyriakopoulou LG, et al. Human kallikrein gene 5 (KLK5) expression by quantitative PCR: an independent indicator of poor prognosis in breast cancer. *Clin Chem.* 2002;48:1241–1250.
- 41. Diamandis EP, Yousef GM. Human tissue kallikrein gene family: a rich source of novel disease biomarkers. *Expert Rev Mol Diagn*. 2001;1:182–190.
- 42. Luo LY, Yousef G, Diamandis EP. Human tissue kallikreins and testicular cancer. *Apmis.* 2003;111:225–232.
- 43. Chang A, Yousef GM, Jung K, Rajpert-De Meyts E, Diamandis EP. Identification and molecular characterization of five novel kallikrein gene 13 (KLK13; KLK-L4) splice variants: differential expression in the human testis and testicular cancer. *Anticancer Res.* 2001;21:3147–3152.
- Bhattacharjee A, Richards WG, Staunton J, et al. Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc Natl Acad Sci U S A*. 2001;98:13790–13795.
- 45. Yousef GM, Borgono CA, Popalis C, et al. In-silico analysis of kallikrein gene expression in pancreatic and colon cancers. *Anticancer Res.* 2004;24:43–51.
- 46. Iacobuzio-Donahue CA, Ashfaq R, Maitra A, et al. Highly expressed genes in pancreatic ductal adenocarcinomas: a comprehensive characterization and comparison of the transcription profiles obtained from three major technologies. *Cancer Res.* 2003;63: 8614–8622.
- 47. Roman-Gomez J, Jimenez-Velasco A, Agirre X, et al. The normal epithelial cell-specific 1 (NES1) gene, a candidate tumor suppressor gene on chromosome 19q13.3-4, is downregulated by hypermethylation in acute lymphoblastic leukemia. *Leukemia*. 2004;18: 362–365.
- 48. Stephan C, Jung K, Lein M, Diamandis EP. PSA and other tissue kallikreins for prostate cancer detection. *Eur J Cancer*. 2007;43:1918–1926.
- 49. Diamandis EP, Okui A, Mitsui S, et al. Human kallikrein 11: a new biomarker of prostate and ovarian carcinoma. *Cancer Res.* 2002;62:295–300.
- Wolf WC, Evans DM, Chao L, Chao J. A synthetic tissue kallikrein inhibitor suppresses cancer cell invasiveness. *Am J Pathol.* 2001;159:1797–1805.
- 51. Yousef GM, Stephan C, Scorilas A, et al. Differential expression of the human kallikrein gene 14 (KLK14) in normal and cancerous prostatic tissues. *Prostate*. 2003;56:287–292.
- Yousef GM, Magklara A, Chang A, Jung K, Katsaros D, Diamandis EP. Cloning of a new member of the human kallikrein gene family, KLK14, which is down-regulated in different malignancies. *Cancer Res.* 2001;61:3425–3431.
- Scheurle D, DeYoung MP, Binninger DM, Page H, Jahanzeb M, Narayanan R. Cancer gene discovery using digital differential display. *Cancer Res.* 2000;60:4037–4043.
- 54. Yousef GM, Polymeris ME, Yacoub GM, et al. Parallel overexpression of seven kallikrein genes in ovarian cancer. *Cancer Res.* 2003;63:2223–2227.
- 55. Dong Y, Kaushal A, Bui L, et al. Human kallikrein 4 (KLK4) is highly expressed in serous ovarian carcinomas. *Clin Cancer Res.* 2001;7:2363–2371.
- Luo LY, Katsaros D, Scorilas A, et al. Prognostic value of human kallikrein 10 expression in epithelial ovarian carcinoma. *Clin Cancer Res.* 2001;7:2372–2379.
- 57. Borgono CA, Grass L, Soosaipillai A, et al. Human kallikrein 14: a new potential biomarker for ovarian and breast cancer. *Cancer Res.* 2003;63:9032–9041.
- Underwood LJ, Tanimoto H, Wang Y, Shigemasa K, Parmley TH, O'Brien TJ. Cloning of tumor-associated differentially expressed gene-14, a novel serine protease overexpressed by ovarian carcinoma. *Cancer Res.* 1999;59:4435–4439.

- Yousef GM, Kyriakopoulou LG, Scorilas A, et al. Quantitative expression of the human kallikrein gene 9 (KLK9) in ovarian cancer: a new independent and favorable prognostic marker. *Cancer Res.* 2001;61:7811–7818.
- 60. Kurlender L, Borgono C, Michael IP, et al. A survey of alternative transcripts of human tissue kallikrein genes. *Biochim Biophys Acta*. 2005;1755:1–14.
- 61. Slawin KM, Shariat SF, Nguyen C, et al. Detection of metastatic prostate cancer using a splice variant-specific reverse transcriptase-polymerase chain reaction assay for human glandular kallikrein. *Cancer Res.* 2000;60:7142–7148.
- 62. Nakamura T, Mitsui S, Okui A, et al. Alternative splicing isoforms of hippostasin (PRSS20/KLK11) in prostate cancer cell lines. *Prostate*. 2001;49:72–78.
- Dong Y, Kaushal A, Brattsand M, Nicklin J, Clements JA. Differential splicing of KLK5 and KLK7 in epithelial ovarian cancer produces novel variants with potential as cancer biomarkers. *Clin Cancer Res.* 2003;9:1710–1720.
- 64. Yousef GM, White NM, Kurlender L, et al. The kallikrein gene 5 splice variant 2 is a new biomarker for breast and ovarian cancer. *Tumour Biol.* 2004;25:221–227.
- 65. Shan SJ, Scorilas A, Katsaros D, Diamandis EP. Transcriptional upregulation of human tissue kallikrein 6 in ovarian cancer: clinical and mechanistic aspects. *Br J Cancer*. 2007;96:362–372.
- Cohen P, Graves HC, Peehl DM, Kamarei M, Giudice LC, Rosenfeld RG. Prostatespecific antigen (PSA) is an insulin-like growth factor binding protein-3 protease found in seminal plasma. *J Clin Endocrinol Metab.* 1992;75:1046–1053.
- 67. Takayama TK, McMullen BA, Nelson PS, Matsumura M, Fujikawa K. Characterization of hK4 (Prostase), a prostate-specific serine protease: activation of the precursor of prostate specific antigen (pro-PSA) and single-chain urokinase-type plasminogen activator and degradation of prostatic acid phosphatase. *Biochemistry*. 2001;40: 15341–15348.
- 68. Smyth MJ. Starum corneum chymotryptic enzyme. In: Barrett AJ, Rawlings NP, Woessner JF, eds. *Handbook of Proteolytic Enzymes*. London: Academic Press; 1998:87–89.
- Plendl J, Snyman C, Naidoo S, Sawant S, Mahabeer R, Bhoola KD. Expression of tissue kallikrein and kinin receptors in angiogenic microvascular endothelial cells. *Biol Chem.* 2000;381:1103–1115.
- Fortier AH, Nelson BJ, Grella DK, Holaday JW. Antiangiogenic activity of prostatespecific antigen. J Natl Cancer Inst. 1999;91:1635–1640.
- Prezas P, Arlt MJ, Viktorov P, et al. Overexpression of the human tissue kallikrein genes KLK4, 5, 6, and 7 increases the malignant phenotype of ovarian cancer cells. *Biol Chem.* 2006;387:807–811.
- 72. Godwin AK, Testa JR, Hamilton TC. The biology of ovarian cancer development. *Cancer*. 1993;71:530–536.
- Holschneider CH, Berek JS. Ovarian cancer: epidemiology, biology, and prognostic factors. Semin Surg Oncol. 2000;19:3–10.
- Langdon SP, Hawkes MM, Lawrie SS, et al. Oestrogen receptor expression and the effects of oestrogen and tamoxifen on the growth of human ovarian carcinoma cell lines. *Br J Cancer*. 1990;62:213–216.
- Murdoch WJ. Perturbation of sheep ovarian surface epithelial cells by ovulation: evidence for roles of progesterone and poly(ADP-ribose) polymerase in the restoration of DNA integrity. *J Endocrinol.* 1998;156:503–508.
- Munstedt K, Steen J, Knauf AG, Buch T, von Georgi R, Franke FE. Steroid hormone receptors and long term survival in invasive ovarian cancer. *Cancer*. 2000;89: 1783–1791.
- 77. Risch HA. Hormonal etiology of epithelial ovarian cancer, with a hypothesis concerning the role of androgens and progesterone. *J Natl Cancer Inst.* 1998;90:1774–1786.
- 78. Levine DA, Boyd J. The androgen receptor and genetic susceptibility to ovarian cancer: results from a case series. *Cancer Res.* 2001;61:908–911.

- Silva EG, Tornos C, Fritsche HA Jr, et al. The induction of benign epithelial neoplasms of the ovaries of guinea pigs by testosterone stimulation: a potential animal model. *Mod Pathol.* 1997;10:879–883.
- Helzlsouer KJ, Alberg AJ, Gordon GB, et al. Serum gonadotropins and steroid hormones and the development of ovarian cancer. *JAMA*. 1995;274:1926–1930.
- Chadha S, Rao BR, Slotman BJ, van Vroonhoven CC, van der Kwast TH. An immunohistochemical evaluation of androgen and progesterone receptors in ovarian tumors. *Hum Pathol.* 1993;24:90–95.
- 82. Kuhnel R, de Graaff J, Rao BR, Stolk JG. Androgen receptor predominance in human ovarian carcinoma. *J Steroid Biochem.* 1987;26:393–397.
- 83. Slotman BJ, Rao BR. Response to inhibition of androgen action of human ovarian cancer cells in vitro. *Cancer Lett.* 1989;45:213–220.
- Luo LY, Soosaipillai A, Grass L, Diamandis EP. Characterization of human kallikreins 6 and 10 in ascites fluid from ovarian cancer patients. *Tumour Biol.* 2006;27:227–234.
- Oikonomopoulou K, Scorilas A, Michael IP, et al. Kallikreins as markers of disseminated tumour cells in ovarian cancer – a pilot study. *Tumour Biol.* 2006;27:104–114.
- Shih I, Salani R, Fiegl M, et al. Ovarian cancer specific kallikrein profile in effusions. *Gynecol Oncol.* 2007;105:501–507.
- 87. Rosen DG, Wang L, Atkinson JN, et al. Potential markers that complement expression of CA125 in epithelial ovarian cancer. *Gynecol Oncol.* 2005;99:267–277.
- Meinhold-Heerlein I, Bauerschlag D, Zhou Y, et al. An integrated clinical-genomics approach identifies a candidate multi-analyte blood test for serous ovarian carcinoma. *Clin Cancer Res.* 2007;13:458–466.
- Luo LY, Katsaros D, Scorilas A, et al. The serum concentration of human kallikrein 10 represents a novel biomarker for ovarian cancer diagnosis and prognosis. *Cancer Res.* 2003;63:807–811.
- Diamandis EP, Scorilas A, Fracchioli S, et al. Human kallikrein 6 (hK6): a new potential serum biomarker for diagnosis and prognosis of ovarian carcinoma. *J Clin Oncol.* 2003; 21:1035–1043.
- 91. Bast RC Jr, Badgwell D, Lu Z, et al. New tumor markers: CA125 and beyond. *Int J Gynecol Cancer*. 2005;15(Suppl 3):274–281.
- Diamandis EP, Yousef GM, Soosaipillai AR, Bunting P. Human kallikrein 6 (zyme/ protease M/neurosin): a new serum biomarker of ovarian carcinoma. *Clin Biochem*. 2000; 33:579–583.
- Dorn J, Schmitt M, Kates R, et al. Primary tumor levels of human tissue kallikreins affect surgical success and survival in ovarian cancer patients. *Clin Cancer Res.* 2007;13: 1742–1748.
- Zheng Y, Katsaros D, Shan SJ, et al. A multiparametric panel for ovarian cancer diagnosis, prognosis, and response to chemotherapy. *Clin Cancer Res.* 2007;13:6984–6992.
- Denmeade SR, Nagy A, Gao J, Lilja H, Schally AV, Isaacs JT. Enzymatic activation of a doxorubicin-peptide prodrug by prostate-specific antigen. *Cancer Res.* 1998;58: 2537–2540.
- 96. Lee SJ, Kim HS, Yu R, et al. Novel prostate-specific promoter derived from PSA and PSMA enhancers. *Mol Ther.* 2002;6:415–421.
- 97. Yousef GM, Polymeris ME, Grass L, et al. Human kallikrein 5: a potential novel serum biomarker for breast and ovarian cancer. *Cancer Res.* 2003;63:3958–3965.
- Tanimoto H, Underwood LJ, Shigemasa K, Parmley TH, O'Brien TJ. Increased expression of protease M in ovarian tumors. *Tumour Biol*. 2001;22:11–18.
- Tanimoto H, Underwood LJ, Shigemasa K, et al. The stratum corneum chymotryptic enzyme that mediates shedding and desquamation of skin cells is highly overexpressed in ovarian tumor cells. *Cancer*. 1999;86:2074–2082.
- 100. Shvartsman HS, Lu KH, Lee J, et al. Overexpression of kallikrein 10 in epithelial ovarian carcinomas. *Gynecol Oncol.* 2003;90:44–50.

- Yousef GM, Fracchioli S, Scorilas A, et al. Steroid hormone regulation and prognostic value of the human kallikrein gene 14 in ovarian cancer. *Am J Clin Pathol*. 2003;119: 346–355.
- 102. Hibbs K, Skubitz KM, Pambuccian SE, et al. Differential gene expression in ovarian carcinoma: identification of potential biomarkers. *Am J Pathol*. 2004;165:397–414.
- 103. Adib TR, Henderson S, Perrett C, et al. Predicting biomarkers for ovarian cancer using gene-expression microarrays. *Br J Cancer*. 2004;90:686–692.
- Bignotti E, Tassi RA, Calza S, et al. Differential gene expression profiles between tumor biopsies and short-term primary cultures of ovarian serous carcinomas: identification of novel molecular biomarkers for early diagnosis and therapy. *Gynecol Oncol.* 2006;103: 405–416.
- Obiezu CV, Scorilas A, Katsaros D, et al. Higher human kallikrein gene 4 (KLK4) expression indicates poor prognosis of ovarian cancer patients. *Clin Cancer Res.* 2001; 7:2380–2386.
- 106. Davidson B, Xi Z, Klokk TI, et al. Kallikrein 4 expression is up-regulated in epithelial ovarian carcinoma cells in effusions. *Am J Clin Pathol*. 2005;123:360–368.
- 107. Kim H, Scorilas A, Katsaros D, et al. Human kallikrein gene 5 (KLK5) expression is an indicator of poor prognosis in ovarian cancer. *Br J Cancer*. 2001;84:643–650.
- Diamandis EP, Borgono CA, Scorilas A, et al. Immunofluorometric quantification of human kallikrein 5 expression in ovarian cancer cytosols and its association with unfavorable patient prognosis. *Tumour Biol.* 2003;24:299–309.
- 109. Hoffman BR, Katsaros D, Scorilas A, et al. Immunofluorometric quantitation and histochemical localisation of kallikrein 6 protein in ovarian cancer tissue: a new independent unfavourable prognostic biomarker. *Br J Cancer*. 2002;87:763–771.
- 110. Shan SJ, Scorilas A, Katsaros D, Rigault de lL, I, Massobrio M, Diamandis EP. Unfavorable prognostic value of human kallikrein 7 quantified by ELISA in ovarian cancer cytosols. *Clin Chem.* 2006;52:1879–1886.
- 111. Kyriakopoulou LG, Yousef GM, Scorilas A, et al. Prognostic value of quantitatively assessed KLK7 expression in ovarian cancer. *Clin Biochem*. 2003;36:135–143.
- 112. Magklara A, Scorilas A, Katsaros D, et al. The human KLK8 (neuropsin/ovasin) gene: identification of two novel splice variants and its prognostic value in ovarian cancer. *Clin Cancer Res.* 2001;7:806–811.
- 113. Borgono CA, Kishi T, Scorilas A, et al. Human kallikrein 8 protein is a favorable prognostic marker in ovarian cancer. *Clin Cancer Res.* 2006;12:1487–1493.
- 114. Shigemasa K, Tian X, Gu L, et al. Human kallikrein 8 (hK8/TADG-14) expression is associated with an early clinical stage and favorable prognosis in ovarian cancer. *Oncol Rep.* 2004;11:1153–1159.
- 115. Diamandis EP, Borgono CA, Scorilas A, Harbeck N, Dorn J, Schmitt M. Human kallikrein 11: an indicator of favorable prognosis in ovarian cancer patients. *Clin Biochem.* 2004;37:823–829.
- 116. Borgono CA, Fracchioli S, Yousef GM, et al. Favorable prognostic value of tissue human kallikrein 11 (hK11) in patients with ovarian carcinoma. *Int J Cancer*. 2003; 106:605–610.
- 117. Scorilas A, Borgono CA, Harbeck N, et al. Human kallikrein 13 protein in ovarian cancer cytosols: a new favorable prognostic marker. *J Clin Oncol.* 2004;22:678–685.
- 118. Yousef GM, Scorilas A, Katsaros D, et al. Prognostic value of the human kallikrein gene 15 expression in ovarian cancer. *J Clin Oncol.* 2003;21:3119–3126.