Critical Reviews in Clinical Laboratory **Sciences**

http://informahealthcare.com/lab ISSN: 1040-8363 (print), 1549-781X (electronic)

informa healthcare

Crit Rev Clin Lab Sci, 2014; 51(3): 138-148 © 2014 Informa Healthcare USA, Inc. DOI: 10.3109/10408363.2014.886180

REVIEW ARTICLE

The bifacial role of helminths in cancer: Involvement of immune and non-immune mechanisms

Katerina Oikonomopoulou¹, Davor Brinc², Andreas Hadjisavvas³, Georgios Christofi⁴, Kyriacos Kyriacou³, and Eleftherios P. Diamandis^{1,2}

¹Department of Pathology and Laboratory Medicine, Mount Sinai Hospital, Toronto, Ontario, Canada, ²Department of Clinical Biochemistry, University Health Network, Toronto, Ontario, Canada, ³Department of Electron Microscopy/Molecular Pathology, The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus, and ⁴Department of Veterinary Services, Ministry of Agriculture, Natural Resources and Environment, Nicosia, Cyprus

Abstract

Infectious agents have been associated with cancer due to activation of pro-carcinogenic inflammatory processes within their host. Several reports, however, indicate that specific pathogens may be able to elicit anti-tumor immune responses that can lead to protection from tumorigenesis or cancer regression. Amongst these "beneficial" pathogens are some helminthic parasites that have already been connected with prevention of autoimmune diseases and allergies, immune conditions increasingly associated with cancer. Even though helminths have co-existed with humans and their ancestors for millions of years, investigations of their impact on human (patho)physiology are relatively new and the functions of components that can explain the helminth bi-directional influence on carcinogenesis are not well understood. This review aims to discuss evidence for the helminth-induced immune, genetic, epigenetic, proteomic, hormonal and metabolic changes that may ultimately mediate the potential pro- or anti-carcinogenic role of helminths. This overview may serve future investigations in clarifying the tumorigenic role of the most common helminthic parasites. It may also inspire the development of anti-cancer regimens and vaccines, in parallel to ongoing efforts of using helminth-based components for the prevention and/or treatment of autoimmune diseases and allergies.

Abbreviations: AID: activation-induced cytidine deaminase; APC: adenomatous polyposis coli; BCG: Bacillus Calmette-Guérin; CXCL8: CXC ligand 8, also known as interleukin 8 (IL-8); FDA: Food and Drug Administration; GST: glutathione-S-transferase; HDAC: histone deacetylases; IARC: International Agency for Research on Cancer; IBD: inflammatory bowel disease; IFNγ: interferon gamma; IgG: immunoglobulin G; IL-10: interleukin 10; Myc: a regulator gene that encodes for a transcription factor; **NF-κB**: nuclear factor kappa-light-chain-enhancer of activated B cells is a DNA transcription factor; p53: tumor protein 53 is a tumour suppressor; RARβ2: retinoic acid receptor β2 gene; KRas: Kirsten rat sarcoma oncogene; RET: oncogene encoding a receptor tyrosine kinase for members of the glial cell line-derived neurotrophic factor family; RNS: reactive nitrogen species; ROS: reactive oxygen species; SCFA: short-chain fatty acids; STAT3: signal transducer and activator of transcription 3; SHE: Syrian hamster embryo; T: Thomsen-Friedenreich mucin-type carbohydrate antigen; TGFβ: transforming growth factor beta; Th cells: Thelper cells; TLR: Toll-like receptors; Tn: precursor of Thomsen-Friedenreich antigen; TNFα: tumor necrosis factor alpha; Tregs: regulatory T cells; TSO: Trichuris suis ova; VIP: vasoactive intestinal polypeptide.

Referee: Dr., Morley D. Hollenberg, Department of Physiology and Pharmacology, University of Calgary, Calgary, Alberta, Canada.

Address for correspondence: Dr Eleftherios P. Diamandis, Department of Pathology and Laboratory Medicine, Mount Sinai Hospital, 60 Murray Street, Room L6-201, Toronto, ON M5T 3L9, Canada. Tel: 416 586-8443. Fax: 416 619-5521. E-mail: ediamandis@mtsinai.on.ca Dr Katerina Oikonomopoulou, Department of Pathology and Laboratory Medicine, Mount Sinai Hospital, 60 Murray Street, Room L6-201, Toronto, ON M5T 3L9, Canada. Tel: 416 586-4800; ext. 8813. Fax: 416 619-5521. E-mail: koikonomopoulou@mtsinai.on.ca

Kevwords

Cancer, helminths, immunity, infection, inflammation

History

Received 15 August 2013 Revised 11 December 2013 Accepted 25 December 2013 Published online 3 March 2014

Introduction

Infections and pathogen-induced inflammation have generally been considered to favor carcinogenesis^{1,2}, taking the blame for an estimated 15.6% of all cancer cases worldwide³. Helminths are among the parasites investigated for their adverse carcinogenic effects on their hosts^{4,5}. A strong association between Schistosoma haematobium and urinary bladder cancer has been historically reported as early as 1911 and confirmed by later reports⁶. The possible relationship



between helminthic infections and cancer also came under the spotlight with the famous observations by Johanes Fibiger on the induction of gastric cancer in rats by the helminth Spiroptera carcinoma, that awarded him a Nobel Prize in Physiology or Medicine in 1926⁷. Even though Fibiger's data had been considered highly debatable and contradictory, according to later reports, they clearly inspired thinking about the association of helminth infections with malignancy.

In contrast to promoting cancer, a few reports have also provided evidence that certain types of pathogens can decrease cancer risk or facilitate tumor regression². In this regard, the hygiene hypothesis, which postulates that the rise of allergic and autoimmune pathologies in Western societies is the result of reduced exposure to certain pathogens at a young age^{8–13} has also been re-evaluated to explain the increased number of some types of cancer in economically-developed countries¹. Specifically, helminths have been proposed as playing a central role in the formation of the hygiene hypothesis¹⁴.

Nowadays, the International Agency for Research on Cancer (IARC) recognizes the role of helminths in promoting carcinogenesis¹⁵. However, reports about their potential anticarcinogenic role are still sparse. This review promotes the idea that the influence of helminthic infections on their host may occur in a complex and context-dependent fashion that may either promote or inhibit carcinogenesis. Our goal is to overview some of the helminth-induced mechanisms that either directly, or through other pathways, influence cancer genesis or progression. While the balance of the ideas discussed in this review tips in favor of the role of the immune system in the helminth-induced carcinogenic changes, other potential inter-connected or independent mechanisms are also presented and discussed.

Parallel-to-humans helminth evolution and anti-helminth immunity

Several interactions between various microorganisms and the human host have occurred as the human population moved from the hunter-gatherer lifestyle to the domestication of animals and the development of agriculture¹⁴. Taking advantage of these new settings, some microbes may have evolved to circumvent the host immune response and, therefore, allow progression of their life cycle with minimal harm to the host. In return, some microbes can exert beneficial functions on host physiology (e.g. metabolism)¹⁶. An example of beneficial microorganisms are the commensals such as lactobacilli and many actinomycetes, including saprophytic mycobacteria, which are relatively harmless microbes that can populate the human body as microflora and affect its (patho)physiology¹⁴.

Helminths have been specifically singled out as organisms that can potentially influence both the host immune system and its metabolism. These worms were inherited by humans from early hominids that existed more than one million years ago and have exerted selection pressure on human populations after the era of animal domestication, which began about 10 000 years ago^{14,17,18}. It has been specifically shown that genetic variability of cytokines associated with autoimmunity is correlated with the diversity of an area's parasites, a major portion of which is comprised of helminths¹⁹.

More specifically, the immune response to helminths can involve several innate and adaptive molecular and cellular components^{20,21}. Host tissue responds to the parasite at the portal of entry (e.g. gastrointestinal mucosa); innate immune cells (e.g. macrophages, dendritic cells, mast cells), as well as non-immune parenchymal cells respond to parasites by activation of antigen processing/presentation, cell migration and cytokine secretion. Response of local tissue, as well as tissue with specialized immune function (e.g. Payer's patches in gastrointestinal mucosa, draining lymph nodes), leads to engagement of T and B cells^{20,21}. The fully developed response to parasites is often characterized as a Th1 or a Th2type response, presenting as classical or alternative inflammation^{20–23}. The overall immune response aims to achieve parasite control and expulsion while, at the same time, regulating host pathology and supporting the pathogen life cycle. This is often achieved by a balanced Th1/Th2 response or by the activation of immune regulatory pathways involving Tregs (e.g. $CD4^+CD25^+Foxp3^+$), as well as immune regulatory cytokines (IL-10, $TGF\beta$)^{20–22}. The Th1/Th2 balanced response that is accompanied by an immune regulatory component is often described as a "modified" Th1 or Th2 response. The result is an attenuated host inflammatory response, as well as a less effective adaptive immune response.

The actual type of response can vary according to individual species of infectious agents and can also change during the course of an infection. For example, a Th1 response is followed by a Th2 response in Schistosoma mansoni and Echinococcus granulosus infections; the Th2 response predominates and is essential for worm expulsion in Heligmosomoides polygyrus; and a mixed Th1/Th2 response develops in *Trichuris muris* murine infections^{21,24}. Disease progression and treatment can also be associated with these changes: for example, in patients undergoing chemotherapy for E. granulosus, a Th1 cytokine profile predominates, while in case of a relapse, the profile shifts to a Th2 dominance^{25,26}. As discussed below, the host immune response to helminths may be of relevance to cancer, as it may either locally or systemically modify long-term pro-inflammatory or antiinflammatory conditions, or it may induce or suppress concurrent host anti-tumor immune responses.

Parasite-induced host cancer-related changes

As proposed by the hygiene hypothesis, allergy and autoimmune diseases have been connected to the decreased exposure to microbes and their products in Western societies⁸⁻¹³. These allergic and autoimmune pathologies have, in turn, been connected to carcinogenesis^{27–32}. For example, autoimmune disease patients have been related to having a higher cancer risk^{27,28}, while allergic cancer patients are more prone to a favorable disease progression and response to therapy³³. In support of a more direct linkage between microbe exposure and cancer, and in extension of the original "immunity-focused" hygiene hypothesis, recent evidence points to an increased incidence of certain types of cancer in Westernized, economically-developed countries, in higher socio-economic status groups, and in families with decreased daycare attendance or low number of siblings^{1,34–37}. It is also



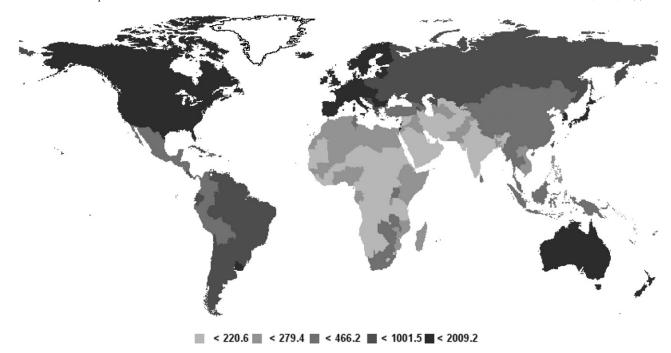


Figure 1. Worldwide cancer prevalence. Estimated 5-year prevalence proportions per 100 000 of all cancers, excluding non-melanoma skin cancer (both sexes, adult population). Source: GLOBOCAN 2008 (IARC) – 7.7.2013^{154,155}.

noted that some major helminthic infections are reported in areas of the world with decreased prevalence of certain types of cancer such as breast, ovarian or prostate (Figure 1). Although, in the case of a multifactorial disease such as cancer, direct conclusions should not be based merely on such limited observations, it is still an intriguing but overlooked epidemiological phenomenon that merits future investigation.

In principle, the altered immune patterns established during an infection can influence the host "tolerance" to cancer genesis, growth and metastasis, changes that can overall translate to an increase or decrease in the number of certain cancer types in pathogen-deprived communities. In this context, a discussion follows on: (a) potential intermediate immune mechanisms that may be involved in the anti- or pro-carcinogenic processes modulated by helminthic infections and (b) other more "direct" noninflammatory pathways through which helminths may modulate carcinogenesis (mechanisms are schematically depicted in Figure 2).

Immune and inflammatory mechanisms

Inflammation

Inflammation can be described as a tissue response to injury, stress or homeostatic disruption, which aims to establish homeostasis but, when left uncontrolled, can lead to a pathology (e.g. autoimmunity, sepsis, fibrosis, metaplasia, cancer^{1,38}). Typical helminth-derived tissue lesions suggest inflammatory and, in some cases, already metaplastic changes. For example, fibrosis, muscular hypertrophy, ulcers and urothelium hyperplasia have been observed in S. haematobium infections⁴. At the same time, recent studies on carcinogenesis have emphasized several apparent links between inflammation and tumor formation or progression³⁹.

For instance, an increased risk of colorectal and systemic cancers in inflammatory bowel disease patients and of hepatocellular carcinoma in hepatitis patients, a possible association of obesity-related low-grade inflammation with tumorigenesis, as well as a potential cancer risk reduction during long-term treatment with non-steroidal anti-inflammatory drugs, have been found in epidemiological studies 40-42

Promotion of anti-tumor immunity

Tumor cell-destructive T cells and antibodies specific for cancer-associated protein and carbohydrate antigens have been described 43-49. In terms of the type of immune response most effective against tumors, cytotoxic T cells and Th1 T cells have often been associated with anti-tumor effects⁵⁰. There are several possible mechanisms by which helminth infection may promote anti-tumor immunity. Parasites may, for example, have indirect actions by stimulating maturation of antigen-presenting cells, thereby increasing the likelihood of an anti-tumor host response. In a non-helminthic but wellknown example, Bacillus Calmette-Guerin (BCG) is successfully used for the treatment of urinary bladder cancer, primarily via induction of host Th1-type responses^{51,52}.

It has also been proposed that cross-reactivity between tumor and helminthic antigens may have an anti-tumorigenic role⁵³. In fact, this issue of helminth-tumor antigen crossreactivity has been documented as a reason for the increased false positive rates of diagnosing infection caused by E. granulosus⁵⁴, a helminth which has been related to carcinogenesis^{55,56}. Notably, antibodies in response to a helminthic infection like E. granulosus are known to persist for years after pathogen removal^{25,26,57}. One of the most immunogenic helminthic components, the T/Tn antigen (also known as the Thomsen-Friedenreich antigen) is a



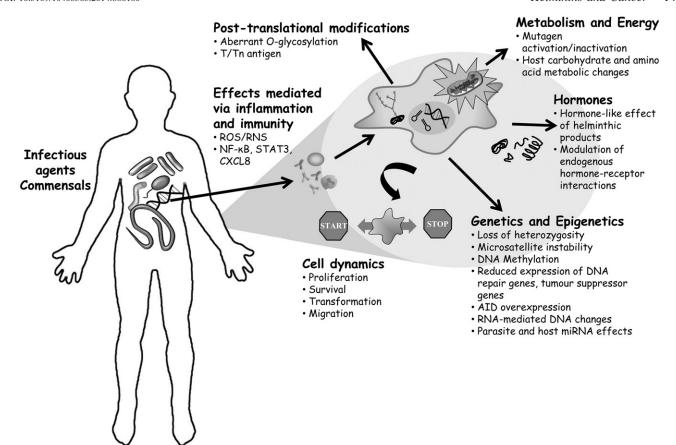


Figure 2. Potential helminth-related processes linked to cancer. A helminthic infection may generate pro- or anti-carcinogenic inflammatory responses and drive immunity in favor of or against a concurrent or subsequent development of a tumor mass. Other alterations at the genetic, epigenetic, proteomic, hormonal and metabolic level may further mediate the beneficial or unfavorable helminthic effects on cancer. AID: Activation-induced cytidine deaminase.

glycoprotein shared by many types of tumors 54,58-60 and expressed more frequently in patients with less extensive malignancy⁵⁴. In support of the potential anti-carcinogenic role of the cross-reactive Tn antigen, experimental vaccination regimens based on T/Tn have been reported successful against breast cancer recurrence⁶¹. In parallel to the idea of cross-reactivity, the "concomitant immunity hypothesis" was suggested to explain the resistance found to the growth of a tumor in animal models previously bearing the same malignant mass⁶²⁻⁶⁴. Concomitant anti-tumor immunity was considered the result of either immunogenic (e.g. possibly due to the presence of shared antigenic epitopes like the aforementioned T/Tn antigen)^{54,58}, or non-immunogenic factors (e.g. the presence of putative anti-mitotic components)⁶². The beneficial effect may persist even after removal of the original mass (sinecomitant immunity)^{62,64}. However, it should be noted that the presence of an anti-tumor immune response does not by itself prevent or cure tumors, as cancer cells have developed mechanisms to evade the host immunity, such as antigen removal or establishment of an immunosuppressive environment.

Immune suppression

Evidence suggests that a "functional" immune system can control spontaneously arising tumors^{65–67}. In addition, immune deficiencies are known to increase the predisposition

to carcinogenesis^{68–70}. Along the same lines, an immunosuppressive environment can be observed in tumor tissues and may be responsible for the limited efficacy of the overall host immune response or of the immune-based anti-cancer therapeutics⁷¹. Suppression of the immune response is known to be caused by helminths; the effect could be mediated through inflammatory components or directly by specific helminthderived products 17,20. For example, helminth cysteine proteinase inhibitors can modify antigen processing and subsequently prevent T and B cell responses, antigen B from E. granulosus, diverse lipid and carbohydrate structures can affect T cell differentiation, while cytokine homologues can mimic the effect of host cytokines and reprogram the immune response^{20,72,73}. Hypo-responsiveness to unrelated antigens in chronic infections has also been described, probably as a result of activation of immune regulatory pathways²⁰. The impact that these immunosuppressive changes may have on carcinogenesis remains an open question, in need of further investigation.

Interactions with concurrent infections

It is known that a persistent helminthic infection can protect the host from a subsequent infection⁷⁴. In principle, during a concurrent helminthic infection, cross-reactive antigens and the overall immune changes may affect the phenotype and survival of other potentially carcinogenic infectious agents in



the human host. Two of the main effects that helminths and the resulting host immune or structural changes may have in terms of carcinogenesis are: (a) to restrict the growth of microbial populations that are potentially carcinogenic and (b) to permit the growth of carcinogen-producing/activating species.

Many helminthic infections can, for example, result in host tissue morphological changes (fibrosis, obstruction, etc.) or offer additional venues for bacterial establishment, all of which may favor the development of local infections or restrict the clearance of endogenous microbial byproducts that can be potentially toxic to the host tissues⁴. In addition, helminths can alleviate the microbially-induced proinflammatory cytokine secretion by secreting IL-10 and $TGF\beta^{75}$. They are also known to promote induction of alternatively activated macrophages and subsequent exacerbation of an enteric bacterial infection that results in colitis in vivo⁷⁶. Moreover, helminths can make animals more resistant to pathogens sensitive to Th2-like responses and more susceptible to pathogens in which protection depends on a Th1-like response⁷⁴. For example, the Th2-inducing helminth H. polygyrus can alter the efficiency of the immune response in removing the Th1-inducing bacterial pathogen Citrobacter rodentium in murine models⁷⁶. Citrobacter rodentium is known to trigger a transient hyperplastic state that increases susceptibility of the animals to carcinogen-induced tumor formation 7. Thus, regulation of the host response to certain infections as the result of the helminth-induced immunity can, in principle, have a profound effect on the direction of responses in favor or against carcinogenic stimuli⁷⁸.

Changes in microbiome/commensal populations

The human microbiota are known to generate metabolites such as through the digestion of plant polysaccharides, the production of short-chain fatty acids, and vitamin K production, which are beneficial for normal physiology^{79–82}. Recent studies have expanded the knowledge about the microbiome effect on the host: for example, microbiomegenerated butyrate has been shown to contribute significantly to ATP generation, prevent autophagy, serve as a histone deacetylases' (HDAC) inhibitor, and induce G-protein coupled receptor signaling in mouse colonocytes^{83,84}. Any disruption of the microbiota balance can potentially have an impact on the susceptibility of the host to many diseases such as allergies, obesity, diabetes, cancer and other infections 16,85,86. Specifically, in the case of tumorigenesis, the human microbiome can have a pivotal role in ultimately regulating the pro- or anti-carcinogenic immune response that will decide the fate and establishment of a concurrent cancer cell⁸⁷. Microbes may cause/favor tumors by promoting immune cell transformation (e.g. Helicobacter pylori and MALT lymphoma), by causing direct dysplasia/metaplasia of parenchymal tissue, or by triggering inflammation⁸⁵. Altered abundance of individual bacterial species, a condition termed dysbiosis, is known to drive inflammation and inflammationassociated colorectal cancers⁸⁸. Inflammation can, in turn, alter the gastrointestinal environment and cause dysbiosis, i.e. by promoting the presence of tumor-associated microbes (as observed in the mouse model of Escherichia coli NC101

and the azoxymethane-treated IL-10-deficient mouse model of colitis-associated cancer⁸⁹). Changes caused in the gut microbiota of mice by obesity have also been shown to increase levels of deoxycholic acid (a bile acid metabolite), which can, upon entering the systemic circulation, reach hepatic stellate cells and induce a senescence-associated secretory phenotype that is characterized by release of inflammatory and tumor-promoting factors in the liver⁹⁰. Moreover, it has recently been shown that bacterial genome integration events into the human somatic cell genome are enriched in certain tumor samples (e.g. acute myeloid leukemia, stomach adenocarcinoma), raising the possibility of a bacterially-mediated genomic influence on host carcinogenic processes⁹¹.

Evidence about the specific effect of helminths on the host microbiota in the case of human infections is still sparse. In principle, helminths, typically a part of the host macrobiome that can also have commensal features, may influence the microbial processes occurring within a host, in a manner similar to what was described about the concurrent infection effect. Preliminary evidence, derived from work on the role of the pig helminth *Trichuris suis*, has reported alterations in the colon microbiota composition and associated metabolism in the precense of the parasitic worm⁹². Major helminthmediated alterations would be expected at the level of mucosal and systemic immunology that could impact the microflora. Regulating the survival of Th1- or Th2-dependent microorganisms, as mentioned in the previous section, is one example of how helminths may be able to mediate such an effect⁷⁴. Helminths are also suggested to induce the release of inflammatory components (i.e. IL-22) capable of maintaining the mucosal barrier function, therefore, possibly regulating the survival and/or expansion of the mucosal microbiota⁹³. In addition, helminths may disrupt host metabolism by modulating the availability of nutrients required by other commensals⁹⁴, possibly forcing analogous adaptations to the infectious or host microbiome species (i.e. bacteria thriving in a polysaccharide as opposed to a fatty-diet environment⁸⁶).

A potential helminth-microbiome axis in favor of increasing numbers of beneficial-to-the-host microbial species may also result in tumor prevention (e.g. H. pylori has an inverse association with esophageal adenocarcinoma)85. Specific microbiota species can also play a key role in production of beneficial food-derived metabolites known to be associated with reduced cancer risk, such as in the case of broccoli⁹⁵. Microbial species can also regulate the metabolic effects of other "pathogenic" microorganisms that could otherwise be able to promote carcinogenesis, e.g. by causing an increase in the conversion of pro-carcinogens to carcinogenic molecules via secretion of bacterial enzymes like β-glucuronidase⁹⁶. Products of bacterial metabolism may also regulate the host immune system against carcinogenesis. For example, a recent study shows that microbially-induced shortchain fatty acids (SCFA) can stimulate Tregs, an event which may prevent chronic inflammatory conditions in the gut⁹⁷. The relation of helminthic infections with these potentially anti-carcinogenic microbial species should be delineated in the future.

The aforementioned potential helminth-mediated impact on human microbiome/commensals and its subsequent pro- or



anti-carcinogenic complications may take place on sites closely associated with the microbes (e.g. GI tract, oral cavity, liver). A systemic or paracrine effect on other sites, in the case of carcinogenesis, may involve different mechanisms such as control of overall immunity or hormonal levels. These potential distal pro- or anti-carcinogenic effects remain to be proven.

Dependency on microbial load

The impact of microbial infections or commensal species on the carcinogenic alterations in the human host may also depend on the microbial load. The term "hormesis" assumes that an agent can have both a beneficial and an unfavorable effect depending on the dose⁹⁸, and it has been suggested that the microbially-induced benefits described by the hygiene hypothesis is one example of a hormetic effect on human hosts⁹⁹. In support of this idea, it has been reported that a higher level of infestation by S. haematobium can lead to multiplication of the patients' risk for bladder cancer⁴. Further, the spectrum of hyperplasia, metaplasia, dysplasia and squamous cell cancer has been associated with established (late-stage), rather than early, schistosomal infection 100.

Genomic, proteomic, hormonal and cell alterations

Genetic instability and epigenetics

Genetic and epigenetic changes can be observed in helminthinfected tissues⁴. For example, DNA methylation has been described in S. haematobium-infected bladder tissues and tumors, and in S. mansoni-infected murine liver⁴. Increased incidence of p53 overexpression was found in colorectal cancer samples from S. mansoni-infected patients 101. Oxidative and nitrative DNA damage was associated with Opisthorchis viverrini in the case of cholangiocarcinoma, with subsequent double-stranded breaks and genome instability, as well as tumor suppressor gene promoter hypermethylation and histone ubiquitination and acetylation 102. Methylative damage was detected in patients with bladder cancer and schistosomiasis (O⁶-methyldeoxyguanosine) in tumor and normal mucosa, as well as in infection mouse models 103,104. It should be noted that methylation DNA damage has been suggested to result from anti-schistosomal drug treatment in vivo and be affected by the extent of formation of alkylating intermediates¹⁰⁵. N-nitroso compounds, derived from environmental sources or due to bacterial infection and formation of infiltrating leukocytes, which have been linked to both schistosomiasis and cancer, are known to convert into active alkylating intermediates ¹⁰⁶. Furthermore, promoter hypermethylation of RARβ₂ and APC genes was detected in schistosomiasis-related bladder cancer patients ¹⁰⁷. In *Taenia solium* infections, that can be associated with brain and hematological malignancies, increased frequency of DNA damage in peripheral blood lymphocytes has been observed 108,109.

The described genetic/epigenetic changes may occur as a result of concurrent helminth-induced inflammation. As an example, ROS/RNS presence and inflammatory cytokine signaling can cause DNA mutations, epigenetic changes and promote cell survival/proliferation, events conducive to tumor cell development 42,110–112. However, helminths may cause

host genome damage via other inflammation-unrelated mechanisms. For example, RNA-mediated damage of DNA in T. solium infection has been described⁴ and T. solium cysticerci are known to release an RNA factor that could transform Syrian hamster embryo (SHE) fibroblasts in vitro¹¹³.

microRNAs

The functional study of miRNA expression by the different helminth species and their effect on the expression of pro- or anti-carcinogenic host genes and miRNAs is in its infancy. The potential roles of host/parasite miRNAs include pathogen life-cycle progression and host cellular control of pathogens^{114–118}. A number of studies, albeit in parasitic protozoa, also show that parasite-mediated changes in host miRNAs may additionally promote inflammation¹¹⁹.

Altered glycosylation

Post-translational modifications, such as glycosylation, play a role in cell-cell communication and cell adhesion, as well as in modulating the functions of a variety of proteins; altered glycosylation patterns have been described in different pathologies, including cancer^{120,121}. The link between specific helminth-induced post-translational modifications, such as the S. mansoni-induced aberrantly O-glycosylated apolipoprotein C-III peptides in human urine 122, and their effect on host pathology are not well studied in helminth infections.

Metabolism, enzyme dysregulation and energy

Changes in metabolizing enzymes such as N-nitrosodimethylamine-N-demethylase (NDMA), cytochrome b5 and glutathione-S-transferase (GST) can be observed in helminth infections. Depending on the role that metabolizing enzymes have in activation or inhibition of a mutagen, an increase or decrease in these enzymes may result in the activation of proneoplastic compounds or reduced inactivation of active mutagens⁴. In S. haematobium infection, enhanced activation of β-glucuronidase can increase the release of carcinogens in a hamster model⁴, while Schistosoma japonicum-liverinfected mice show decreased metabolism of mutagens¹²³. Evidence from animal studies on the increased carcinogenic role of O. viverrini with the combined administration of the carcinogen N-nitrosodimethylamine may also indicate a role for this helminth in regulating the metabolic capacity of the host against this carcinogen⁴. Notably, similar experiments in Fasciola hepatica infection models pointed to a bidirectional effect of the microorganism on carcinogenesis, namely promoting carcinogenesis during the acute phase of infection (i.e. by inducing immune or genetic changes) and restricting carcinogenesis during the chronic phase, by inhibiting the activation of the carcinogen in the liver (e.g. N-nitrosodimethylamine-initiated malignant transformation)⁴.

Furthermore, changes in endogenous host metabolic functions have been observed in infected patients or animal models. For example, S. mansoni infection was associated with changes in urine metabolites in vivo, suggesting increases in glycolysis and alterations in the metabolism of amino acids like tryptophan and alanine¹²⁴. Abnormal tryptophan metabolites can be detected also



S. haematobium infection, while O. viverrini infection increases nitrosamine and nitrate production, as well as iron accumulation⁴.

Hormonal imbalance

The susceptibility of a host to a parasitic infection is often regulated by endocrine factors, either due to modulation of innate immunity or by hormones having several direct effects on the parasites 125. Parasites themselves can also cause alterations in functions of the host endocrine system during an infection, i.e. by secreting hormones and neuropeptides or by triggering changes in the host's hormonal levels, influencing host immunity and parasite survival^{125,126}. Moreover, hormone-host receptor interactions can be affected during a helminthic infection, as in the case of Spirometra mansonoides infection where a growth hormone-like function has been attributed to a compound secreted by the parasite 127. The evolutionary aim of the parasite-host hormonal interactions is for the microbe to achieve favorable establishment, growth and reproduction at the lowest possible cost for the host 126.

A sustained hormonal modulation as a result of a helminth infection may have various secondary effects on host (patho)physiology at several levels, where hormones can be involved (i.e. development, differentiation, cell growth, etc.)¹²⁵. Hormonal changes induced during the infection may potentially lead to uncontrolled cell growth and sustained pro-carcinogenic signals. Not only are hormones able to alter the immune state of the host ^{125,126} and affect its responsiveness to a new trigger, but they may also induce pathways typically involved in hormone-dependent carcinogenesis (i.e. endocrine-related tumors like prostate, breast, ovarian, etc.). Another hormonal effect that helminths can potentially have is indirectly via their action on microbiota: it has been suggested that certain microbes may modulate host estrogen metabolism leading to enhanced absorption and higher level of estrogen exposure, which may in turn promote tumors in women⁸⁵. Another example supporting the hormonal hypothesis is the case of O. viverrini, a helminth that causes cholangiocarcinoma, or cancer of the bile ducts. It is now known that the parasite expresses a granulin-like growth factor that can potentially induce cell growth and development of carcinoma¹²⁸. Trematodes are also known to secrete peptides that mimic vasoactive intestinal polypeptide (VIP) leading to villus pathology and hyperplasia 125. The relationship between parasite-triggered hormonal alterations and cancer is an intriguing field open to future investigations.

Cell proliferation, survival and invasion

Helminth-derived factors can directly promote host cell proliferation. For example, products from O. viverrini are able to stimulate murine fibroblast proliferation ¹²⁹. Effects on cell proliferation, survival and invasion potential can, in principle, be mediated by the helminth-induced inflammatory responses as well. Recent studies show that oncogene overexpression and activation in epithelial cells (e.g. Kras, Myc, RET) together with secretion of pro-inflammatory cytokines (e.g. IL-6), chemokines (e.g. CXCL8) and growth factors by epithelial cells, stromal cells, and infiltrating

immune cells can lead to cell hyperplasia and transformation^{42,130–134}. The NF-κB signaling pathway is shown to be of importance for mediating cell invasion of a metastatic ovarian cancer cell line in a mouse xenograft model, as well as facilitating breast cancer metastasis and survival of castrateresistant prostate cancer^{110,111}. NF-κB activation in stromal, immune and tumor cells may be synergistic for tumor promotion. For example, NF-κB activation in myeloid cells leads to IL-6 secretion and subsequent induction of STAT3 signaling in pre-malignant intestinal epithelial cells that, together with cell autonomous NF-kB signaling, lead to proliferation and survival of tumor cells in a mouse model of experimentally-induced colorectal cancer¹¹¹. In some models, however, autonomous NF-kB signaling can promote survival of host cells, which results in preventing compensatory proliferation that would otherwise lead to pro-tumorigenic functions. Finally, some of the receptors and pathways classically involved in inflammation may have inflammation-independent roles in tumorigenesis. For example, TLR2 signaling can promote cell proliferation and tumorigenesis independently of its role in inflammation¹³⁵.

Anti-inflammatory helminthic therapy as a paradigm for cancer therapeutics

Given the diverse actions of helminths and their involvement in a variety of diseases ranging from immune pathologies to cancer, the therapeutic manipulation of the "beneficial" helminthic species in clinical practice seems an intriguing idea. Several components of parasitic helminths are already under discussion for the treatment of immune and allergic pathologies such as Crohn's disease, ulcerative colitis, allergies, multiple sclerosis, inflammatory bowel disease, and more recently autism^{72,136-139}. Of those, attention is drawn to Trichuris suis, a pig pathogenic helminth that is not a natural parasite for humans but can still induce immune changes in the human host 139. Several trials have sought to evaluate the efficacy of T. suis parasitic eggs (known as T. suis ova or TSO) in the treatment of allergic and autoimmune diseases.

In one of the trials, a randomized controlled protocol was performed in 54 patients with severe ulcerative colitis (by means of index score >4)¹⁴⁰. One group of 30 patients received 2500 T. suis ova orally at 2-week intervals for 12 weeks, while the remaining 24 patients were included in the placebo group. Improvement of the disease activity index score occurred in 43% of patients under the helminth treatment as opposed to 17% of control subjects (p = 0.04). The study also reported no significant side effects of the parasitic treatment.

The safety of TSO for use in clinical practice was specifically addressed in other similar trials; overall these studies, referring to the management of multiple sclerosis, allergic rhinitis and food allergy, showed that patients experienced mild-to-moderate gastrointestinal side effects and mild eosinophilia 141-144. Notably, these effects were not observed in the inflammatory bowel disease (IBD) studies (e.g. ulcerative colitis; see above)^{140,145,146}, a difference that has been attributed to sample preparation variations, concurrent patient treatments and the characteristics of IBD (i.e. gastrointestinal symptoms)¹³⁹.



Helminths and Cancer DOI: 10.3109/10408363.2014.886180

Many potential mechanisms that may facilitate the beneficial action of T. suis have been discussed, including alterations in the commensal populations and regulation of specific immune components (e.g. Tregs or dendritic cells), critical for the helminth-triggered Th2 responses and immune tolerance¹³⁹. Even though the exact processes are largely undiscovered, these trials can serve as an efficient and safe paradigm for developing equivalent cancer therapeutics, under the idea that several of the helminth-induced immunological responses may have the ability to regulate cancer genesis or progression.

As supportive evidence for the potentially favorable therapeutic efficiency of helminths in cancer treatment, studies have pointed to a role for the shared helminth-tumor T/Tn antigen; these antigens are structurally similar to the blood type A glycoprotein and can be expressed in normal tissues, albeit at lower levels compared to malignant cells 147,148. Despite the non-specific expression of T/Tn antigen, vaccinations based on the T/Tn antigen have been reported to be effective in preventing breast cancer recurrence, an outcome that could be explained as the result of potential anti-tumor immune responses, elicited due to the increase of helper T lymphocytes and the decrease of T suppressor/cytotoxic cells ratio⁶¹. There has also been an interest in the use of Tn antigen in the management of prostate cancer patients with, however, limited applicability thus far possibly due to the inconsistent expression of this antigen on prostate tumors¹⁴⁹.

The idea of using an infectious agent for the management of cancer patients is not new and resembles the idea behind the BCG vaccine and the William Coley's toxin regimens. The BCG vaccine contains an attenuated form of Mycobacterium bovis that is now a formulated FDA-approved vaccine for the first-line intravesical treatment of bladder cancer^{51,52}. It has been hypothesized that the anti-cancer effects of the vaccine enhance the host Th1 cytokine production (e.g. IFN-γ, $TNF\alpha$)^{150,151}. Similar preliminary efforts by William Coley took place in the 1900s using an attenuated bacterial mixture (Streptococcus pyogenes and Serratia marcescens) for the management of cancer patients, an effect that was attributed to the potential action of $TNF\alpha^{152,153}$.

Given the tight immune interconnections of helminthic infections with their hosts, as well as the links between cancer and inflammatory conditions that can be treated using helminthic components and the anti-carcinogenic actions of the BCG and Coley's regimens, the role of helminths in the modulation of carcinogenesis (both in favor and against) should witness intense future translational research.

Conclusions and future perspectives

Helminths have been a part of the human evolutionary environment for millions of years. Both the current parasites and the human host may thus have inherited features allowing more "symbiotic" rather than parasitic relationships. Even though microorganisms are known to generally have both protective and unfavorable effects in the human host, the specific role of helminths in cancer is only now starting to unravel. Despite the plethora of evidence, discussed in this manuscript, pointing to the carcinogenic impact of parasitic

agents like helminths, we aim to draw attention to the often neglected beneficial effects that relatively benign organisms like some helminths can have, as dictated by the supporters of the original hygiene hypothesis. We believe that some helminth-triggered effects and, more specifically, immunemediated changes, can be beneficial for cancer prevention/ regression in the host and should be investigated in more detail in the future.

Clinical trials using helminths in the treatment of allergies and autoimmune pathologies are currently ongoing and cancer therapeutics can also benefit from these novel mechanistic and clinical studies since the findings can potentially provide insights into the role of helminths in modulating the host immune response towards cancer prevention. For example, explorations of the adjuvant and cross-reactive effects of these organisms, as well as their immune functions, have the potential to lead to therapeutic regimens, analogous to the T/Tn vaccination studies discussed in earlier sections, in order to prevent, regress or slow down cancer progression or to improve quality of life for cancer patients. As helminths may be part of the human macrobiome or influence commensal bacterial populations in the human host, it is also important to highlight the possibility of developing personalized treatments for cancer patients, based on their individual spectrum of commensal species in order to maximize the therapeutic benefit in cancer clinical practice.

Acknowledgements

Due to space limitations, we apologize for not citing more reviews and original papers related to this topic.

Declaration of interest

The authors report that no conflicts of interest exist.

References

- 1. Rook GA, Dalgleish A. Infection, immunoregulation, and cancer. Immunol Rev 2011;240:141-59.
- Oikonomopoulou K, Brinc D, Kyriacou K, Diamandis EP. Infection and cancer: revaluation of the hygiene hypothesis. Clin Cancer Res 2013;19:2834-41.
- 3. Pisani P, Parkin DM, Munoz N, Ferlay J. Cancer and infection: estimates of the attributable fraction in 1990. Cancer Epidemiol Biomarkers Prev 1997;6:387-400.
- 4. Mayer DA, Fried B. The role of helminth infections in carcinogenesis. Adv Parasitol 2007;65:239-96.
- Fried B, Reddy A, Mayer D. Helminths in human carcinogenesis. Cancer Lett 2011;305:239-49.
- Gouda I, Mokhtar N, Bilal D, et al. Bilharziasis and bladder cancer: a time trend analysis of 9843 patients. J Egypt Natl Canc Inst 2007;19:158-62.
- Stolley PD, Lasky T. Johannes Fibiger and his Nobel Prize for the hypothesis that a worm causes stomach cancer. Ann Intern Med 1992:116:765-9
- 8. Bach JF. The effect of infections on susceptibility to autoimmune and allergic diseases. N Engl J Med 2002;347:911-20.
- Strachan DP. Hay fever, hygiene, and household size. BMJ 1989; 299:1259-60.
- Strachan DP. Family size, infection and atopy: the first decade of the "hygiene hypothesis". Thorax 2000;55:S2–10.
- 11. Rook GA. 99th Dahlem conference on infection, inflammation and chronic inflammatory disorders: darwinian medicine and the 'hygiene' or 'old friends' hypothesis. Clin Exp Immunol 2010;160:



- Wills-Karp M, Santeliz J, Karp CL. The germless theory of allergic disease: revisiting the hygiene hypothesis. Nat Rev Immunol 2001:1:69-75.
- 13. Ngoi SM, Sylvester FA, Vella AT. The role of microbial byproducts in protection against immunological disorders and the hygiene hypothesis. Discov Med 2011;12:405-12.
- Rook GA. Review series on helminths, immune modulation and the hygiene hypothesis: the broader implications of the hygiene hypothesis. Immunology 2009;126:3-11.
- 15. Schistosomes, liver flukes and Helicobacter pylori. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Lyon, 7-14 June 1994. IARC Monogr Eval Carcinog Risks Hum 1994;61:1-241.
- 16. Devaraj S, Hemarajata P, Versalovic J. The human gut microbiome and body metabolism: implications for obesity and diabetes. Clin Chem 2013;59:617-28.
- 17. Dunne DW, Cooke A. A worm's eye view of the immune system: consequences for evolution of human autoimmune disease. Nat Rev Immunol 2005;5:420-6.
- 18. Jackson JA, Friberg IM, Little S, Bradley JE. Review series on helminths, immune modulation and the hygiene hypothesis: immunity against helminths and immunological phenomena in modern human populations: coevolutionary legacies? Immunology 2009;126:18-27.
- Fumagalli M, Pozzoli U, Cagliani R, et al. Parasites represent a major selective force for interleukin genes and shape the genetic predisposition to autoimmune conditions. J Exp Med 2009;206: 1395-408.
- 20. Maizels RM, Yazdanbakhsh M. Immune regulation by helminth parasites: cellular and molecular mechanisms. Nat Rev Immunol
- 21. Anthony RM, Rutitzky LI, Urban Jr JF, et al. Protective immune mechanisms in helminth infection. Nat Rev Immunol 2007;7: 975 - 87.
- 22. Sica A, Larghi P, Mancino A, et al. Macrophage polarization in tumour progression. Semin Cancer Biol 2008;18:349-55.
- Girgis NM, Gundra UM, Loke P. Immune regulation during helminth infections. PLoS Pathog [Online] 2013;9:e1003250.
- Mourglia-Ettlin G, Marques JM, Chabalgoity JA, Dematteis S. Early peritoneal immune response during Echinococcus granulosus establishment displays a biphasic behavior. PLoS Negl Trop Dis [Online] 2011;5:e1293.
- Zhang W, Li J, McManus DP. Concepts in immunology and diagnosis of hydatid disease. Clin Microbiol Rev 2003;16:18–36.
- Zhang W, Ross AG, McManus DP. Mechanisms of immunity in hydatid disease: implications for vaccine development. J Immunol 2008:181:6679-85.
- 27. Hemminki K, Liu X, Ji J, et al. Autoimmune disease and subsequent digestive tract cancer by histology. Ann Oncol 2011; 23:927-33.
- 28. Hemminki K, Liu X, Ji J, et al. Subsequent COPD and lung cancer in patients with autoimmune disease. Eur Respir J 2011; 37:463-5
- Merrill RM, Isakson RT, Beck RE. The association between allergies and cancer: what is currently known? Ann Allergy Asthma Immunol 2007;99:102-16.
- 30. Prizment AE, Folsom AR, Cerhan JR, et al. History of allergy and reduced incidence of colorectal cancer, Iowa Women's Health Study. Cancer Epidemiol Biomarkers Prev 2007;16: 2357-62.
- 31. Maisonneuve P, Lowenfels AB, Bueno-de-Mesquita HB, et al. Past medical history and pancreatic cancer risk: results from a multicenter case-control study. Ann Epidemiol 2010;20:92-8.
- 32. Engkilde K, Thyssen JP, Menne T, Johansen JD. Association between cancer and contact allergy: a linkage study. BMJ Open [Online] 2011;1:e000084.
- 33. Pompei R, Lampis G, Ingianni A, et al. Allergy and tumour outcome after primary cancer therapy. Int Arch Allergy Immunol 2004;133:174-8.
- 34. Rastogi T, Devesa S, Mangtani P, et al. Cancer incidence rates among South Asians in four geographic regions: India, Singapore, UK and US. Int J Epidemiol 2008;37:147-60.
- Gutensohn N, Cole P. Childhood social environment and Hodgkin's disease. N Engl J Med 1981;304:135-40.

- 36. Ma X, Buffler PA, Selvin S, et al. Daycare attendance and risk of childhood acute lymphoblastic leukaemia. Br J Cancer 2002;86: 1419-24
- 37. Gilham C, Peto J, Simpson J, et al. Day care in infancy and risk of childhood acute lymphoblastic leukaemia: findings from UK casecontrol study. BMJ 2005;330:1294.
- Medzhitov R. Origin and physiological roles of inflammation. Nature 2008;454:428-35.
- Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. Nature 2008;454:436-44.
- Berg M, Soreide K. Prevention: will an aspirin a day keep the colorectal cancer away? Nat Rev Clin Oncol 2011;8:130-1.
- Kaiser J. Will an aspirin a day keep cancer away? Science 2012; 337:1471-3
- Trinchieri G. Cancer and inflammation: an old intuition with rapidly evolving new concepts. Annu Rev Immunol 2012;30: 677-706.
- Stevanovic S. Identification of tumour-associated T-cell epitopes for vaccine development. Nat Rev Cancer 2002;2: 514-20.
- 44. Hudis CA. Trastuzumab mechanism of action and use in clinical practice. N Engl J Med 2007;357:39-51.
- Heimburg-Molinaro J, Lum M, Vijay G, et al. Cancer vaccines and carbohydrate epitopes. Vaccine 2011;29:8802-26.
- Mellman I, Coukos G, Dranoff G. Cancer immunotherapy comes of age. Nature 2011;480:480-9.
- Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. Nat Rev Cancer 2012;12:265-77.
- Scott AM, Wolchok JD, Old LJ. Antibody therapy of cancer. Nat Rev Cancer 2012;12:278-87.
- Takahashi N, Ohkuri T, Homma S, et al. First clinical trial of cancer vaccine therapy with artificially synthesized helper/killerhybrid epitope long peptide of MAGE-A4 cancer antigen. Cancer Sci 2012;103:150-3.
- Zamarron BF, Chen W. Dual roles of immune cells and their factors in cancer development and progression. Int J Biol Sci 2011;7:651-8.
- 51. Brandau S, Suttmann H. Thirty years of BCG immunotherapy for non-muscle invasive bladder cancer: a success story with room for improvement. Biomed Pharmacother 2007;61:299-305.
- Alexandroff AB, Nicholson S, Patel PM, Jackson AM. Recent advances in bacillus Calmette-Guerin immunotherapy in bladder cancer. Immunotherapy 2010;2:551-60.
- 53. Osinaga E. Expression of cancer-associated simple mucin-type O-glycosylated antigens in parasites. IUBMB Life 2007;59: 269 - 73
- 54. Pfister M, Gottstein B, Cerny T, Cerny A. Immunodiagnosis of echinococcosis in cancer patients. Clin Microbiol Inf 1999;5: 693 - 7
- Akgul H, Tez M, Unal AE, et al. Echinococcus against cancer: why not? Cancer 2003;98:1999-2000.
- Darani HY, Soozanger N, Khorami S, et al. Hydatid cyst protoscolices induce cell death in WEHI-164 fibrosarcoma cells and inhibit the proliferation of baby hamster kidney fibroblasts in vitro. J Parasitol Res 2012;2012:304183.
- 57. Zhang W, McManus DP. Recent advances in the immunology and diagnosis of echinococcosis. FEMS Immunol Med Microbiol 2006;47:24-41.
- Springer GF. Immunoreactive T and Tn epitopes in cancer diagnosis, prognosis, and immunotherapy. J Mol Med (Berl) 1997; 75:594-602
- Alvarez ED, Medeiros A, Miguez M, et al. O-glycosylation in Echinococcus granulosus: identification and characterization of the carcinoma-associated Tn antigen. Exp Parasitol 2001;98: 100 - 9.
- van Knapen F. Echinococcus granulosus infection and malignancy. BMJ 1980;281:195-6.
- Springer GF, Desai PR, Spencer BD, et al. T/Tn antigen vaccine is effective and safe in preventing recurrence of advanced breast carcinoma. Cancer Detect Prev 1995;19:374-80.
- Gorelik E, Segal S, Feldman M. On the mechanism of tumor "concomitant immunity". Int J Cancer 1981;27:847–56.
- Cox FE. Concomitant infections, parasites and immune responses. Parasitology 2001;122:S23-38.



- 64. Fisher B, Saffer EA, Fisher ER. Comparison of concomitant and sinecomitant tumor immunity. Proc Soc Exp Biol Med 1970;135:
- Dunn GP, Bruce AT, Ikeda H, et al. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol 2002;3: 991-8.
- Nakachi K, Hayashi T, Imai K, Kusunoki Y. Perspectives on cancer immuno-epidemiology. Cancer Sci 2004;95:921-9.
- Koebel CM, Vermi W, Swann JB, et al. Adaptive immunity maintains occult cancer in an equilibrium state. Nature 2007;450:
- 68. Boshoff C, Weiss R. AIDS-related malignancies. Nat Rev Cancer 2002;2:373-82.
- Haliotis T, Ball JK, Dexter D, Roder JC. Spontaneous and induced primary oncogenesis in natural killer (NK)-cell-deficient beige mutant mice. Int J Cancer 1985;35:505-13.
- Shankaran V, Ikeda H, Bruce AT, et al. IFNgamma and lymphocytes prevent primary tumour development and shape tumour immunogenicity. Nature 2001;410:1107-11.
- 71. Draghiciu O, Nijman HW, Daemen T. From tumor immunosuppression to eradication: targeting homing and activity of immune effector cells to tumors. Clin Dev Immunol 2011;2011:439053.
- Adisakwattana P, Saunders SP, Nel HJ, Fallon PG. Helminthderived immunomodulatory molecules. Adv Exp Med Biol 2009; 666:95-107.
- Virginio VG, Monteiro KM, Drumond F, et al. Excretory/ secretory products from in vitro-cultured Echinococcus granulosus protoscoleces. Mol Biochem Parasitol 2012;183:15-22.
- MacDonald AS, Araujo MI, Pearce EJ. Immunology of parasitic helminth infections. Infect Immun 2002;70:427-33.
- van Riet E, Hartgers FC, Yazdanbakhsh M. Chronic helminth infections induce immunomodulation: consequences and mechanisms. Immunobiology 2007;212:475-90.
- Weng M, Huntley D, Huang IF, et al. Alternatively activated macrophages in intestinal helminth infection: effects on concurrent bacterial colitis. J Immunol 2007;179:4721-31.
- 77. Luperchio SA, Schauer DB. Molecular pathogenesis of Citrobacter rodentium and transmissible murine colonic hyperplasia. Microbes Infect 2001;3:333-40.
- Moreau E, Chauvin A. Immunity against helminths: interactions with the host and the intercurrent infections. J Biomed Biotechnol 2010;2010:428593.
- Macfarlane GT, Macfarlane S. Fermentation in the human large intestine: its physiologic consequences and the potential contribution of prebiotics. J Clin Gastroenterol 2011;45:S120-7.
- Resta SC. Effects of probiotics and commensals on intestinal epithelial physiology: implications for nutrient handling. J Physiol 2009;587:4169-74.
- 81. Vipperla K, O'Keefe SJ. The microbiota and its metabolites in colonic mucosal health and cancer risk. Nutr Clin Pract 2012;27:
- 82. Holmes E, Li JV, Athanasiou T, et al. Understanding the role of gut microbiome-host metabolic signal disruption in health and disease. Trends Microbiol 2011;19:349-59.
- 83. Donohoe DR, Garge N, Zhang X, et al. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. Cell Metab 2011;13:517-26.
- Dumas ME. The microbial-mammalian metabolic axis: beyond simple metabolism. Cell Metab 2011;13:489-90.
- Plottel CS, Blaser MJ. Microbiome and malignancy. Cell Host Microbe 2011;10:324-35.
- Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. Cell 2012; 148:1258-70
- Khan AA, Shrivastava A, Khurshid M. Normal to cancer microbiome transformation and its implication in cancer diagnosis. Biochim Biophys Acta 2012;1826:331-7.
- Hajishengallis G, Darveau RP, Curtis MA. The keystone-pathogen hypothesis. Nat Rev Microbiol 2012;10:717-25.
- Arthur JC, Perez-Chanona E, Muhlbauer M, et al. Intestinal inflammation targets cancer-inducing activity of the microbiota. Science 2012;338:120-3.
- Yoshimoto S, Loo TM, Atarashi K, et al. Obesity-induced gut microbial metabolite promotes liver cancer through senescence secretome. Nature 2013;499:97-101.

- 91. Riley DR, Sieber KB, Robinson KM, et al. Bacteria-human somatic cell lateral gene transfer is enriched in cancer samples. PLoS Comput Biol [Online] 2013;9:e1003107.
- 92. Wu S, Li RW, Li W, et al. Worm burden-dependent disruption of the porcine colon microbiota by Trichuris suis infection. PLoS One [Online] 2012;7:e35470.
- 93. Leung JM, Loke P. A role for IL-22 in the relationship between intestinal helminths, gut microbiota and mucosal immunity. Int J Parasitol 2013;43:253-7.
- 94. Li RW, Wu S, Li W, et al. Alterations in the porcine colon microbiota induced by the gastrointestinal nematode Trichuris suis. Infect Immun 2012;80:2150-7
- 95. Lai RH, Miller MJ, Jeffery E. Glucoraphanin hydrolysis by microbiota in the rat cecum results in sulforaphane absorption. Food Funct 2010;1:161-6.
- Kulkarni N, Reddy BS. Inhibitory effect of Bifidobacterium longum cultures on the azoxymethane-induced aberrant crypt foci formation and fecal bacterial beta-glucuronidase. Proc Soc Exp Biol Med 1994:207:278-83.
- 97. Smith PM, Howitt MR, Panikov N, et al. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science 2013;341:569-73.
- Mattson MP. Hormesis defined. Ageing Res Rev 2008;7:1-7.
- 99. Bukowski JA, Lewis RJ. Is the hygiene hypothesis an example of hormesis? Nonlinearity Biol Toxicol Med 2003;1:155-66.
- 100. El-Bolkainy MN, Mokhtar NM, Ghoneim MA, Hussein MH. The impact of schistosomiasis on the pathology of bladder carcinoma. Cancer 1981;48:2643-8.
- 101. Madbouly KM, Senagore AJ, Mukerjee A, et al. Colorectal cancer in a population with endemic Schistosoma mansoni: is this an atrisk population? Int J Colorectal Dis 2007;22:175-81.
- 102. Murata M, Thanan R, Ma N, Kawanishi S. Role of nitrative and oxidative DNA damage in inflammation-related carcinogenesis. J Biomed Biotechnol [Online] 2012;2012:623019.
- 103. Badawi AF, Mostafa MH, Aboul-Azm T, et al. Promutagenic methylation damage in bladder DNA from patients with bladder cancer associated with schistosomiasis and from normal individuals. Carcinogenesis 1992;13:877-81.
- Badawi AF, Cooper DP, Mostafa MH, et al. Promutagenic methylation damage in liver DNA of mice infected with Schistosoma mansoni. Carcinogenesis 1993;14:653-7.
- 105. Badawi AF, Awney HA, Mostafa MH. Formation of promutagenic methylation damage in tissue-DNA of mice treated with antischistosomal agents. Cancer Lett 1993;75:167-73.
- 106. Badawi AF, Mostafa MH, O'Connor PJ. Involvement of alkylating agents in schistosome-associated bladder cancer: the possible basic mechanisms of induction. Cancer Lett 1992;63:171-88.
- 107. Eissa S, Swellam M, El-Khouly IM, et al. Aberrant methylation of RARbeta2 and APC genes in voided urine as molecular markers for early detection of bilharzial and nonbilharzial bladder cancer. Cancer Epidemiol Biomarkers Prev 2011;20:1657–64.
- 108. Herrera LA, Ramirez T, Rodriguez U, et al. Possible association between Taenia solium cysticercosis and cancer: increased frequency of DNA damage in peripheral lymphocytes from neurocysticercosis patients. Trans R Soc Trop Med Hyg 2000;94: 61-5.
- 109. Herrera LA, Rodriguez U, Gebhart E, Ostrosky-Wegman P. Increased translocation frequency of chromosomes 7, 11 and 14 in lymphocytes from patients with neurocysticercosis. Mutagenesis 2001;16:495–7.
- 110. Hsu S, Kim M, Hernandez L, et al. IKK-epsilon coordinates invasion and metastasis of ovarian cancer. Cancer Res 2012;72: 5494-504
- 111. DiDonato JA, Mercurio F, Karin M. NF-kappaB and the link between inflammation and cancer. Immunol Rev 2012;246: 379-400.
- Bauer J, Namineni S, Reisinger F, et al. Lymphotoxin, NF-kB, and 112. cancer: the dark side of cytokines. Dig Dis 2012;30:453-68.
- 113. Herrera LA, Santiago P, Rojas G, et al. Immune response impairment, genotoxicity and morphological transformation induced by Taenia solium metacestode. Mutat Res 1994;305: 223 - 8.
- 114. Hao L, Cai P, Jiang N, et al. Identification and characterization of microRNAs and endogenous siRNAs in Schistosoma japonicum. BMC Genomics [Online] 2010;11:55.



- Liu Q, Tuo W, Gao H, Zhu XQ. MicroRNAs of parasites: current status and future perspectives. Parasitol Res 2010;107:501-7.
- Simoes MC, Lee J, Djikeng A, et al. Identification of Schistosoma mansoni microRNAs. BMC Genomics [Online] 2011;12:47.
- 117. Cheng G, Jin Y. MicroRNAs: potentially important regulators for schistosome development and therapeutic targets against schistosomiasis. Parasitology 2012;139:669-79.
- 118. Manzano-Roman R, Siles-Lucas M. MicroRNAs in parasitic diseases: potential for diagnosis and targeting. Mol Biochem Parasitol 2012;186:81-6.
- Hu G, Zhou R, Liu J, et al. MicroRNA-98 and let-7 regulate expression of suppressor of cytokine signaling 4 in biliary epithelial cells in response to Cryptosporidium parvum infection. J Infect Dis 2010;202:125-35.
- 120. Blomme B, Van SC, Callewaert N, Van VH. Alteration of protein glycosylation in liver diseases. J Hepatol 2009;50:592-603.
- 121. Chandler K, Goldman R. Glycoprotein disease markers and single protein-omics. Mol Cell Proteomics 2013;12:836-45.
- Balog CI, Mayboroda OA, Wuhrer M, et al. Mass spectrometric identification of aberrantly glycosylated human apolipoprotein C-III peptides in urine from Schistosoma mansoni-infected individuals. Mol Cell Proteomics 2010;9:667-81.
- Aji T, Matsuoka H, Ishii A, et al. Retention of a mutagen, 3-amino-1-methyl-5H-pyrido[4,3-b]indole (Trp-P-2), in the liver of mice infected with Schistosoma japonicum. Mutat Res 1994; 305:265-72.
- 124. Wang Y, Holmes E, Nicholson JK, et al. Metabonomic investigations in mice infected with Schistosoma mansoni: an approach for biomarker identification. Proc Natl Acad Sci USA 2004;101: 12676-81.
- Beckage NE. Endocrine and neuroendocrine host-parasite relationships. Receptor 1993;3:233-45.
- Hernandez-Bello R, Escobedo G, Guzman C, et al. Immunoendocrine host-parasite interactions during helminth infections: from the basic knowledge to its possible therapeutic applications. Parasite Immunol 2010;32:633-43.
- 127. Phares K. An unusual host-parasite relationship: the growth hormone-like factor from plerocercoids of spirometrid tapeworms. Int J Parasitol 1996;26:575-88.
- 128. Smout MJ, Laha T, Mulvenna J, et al. A granulin-like growth factor secreted by the carcinogenic liver fluke, Opisthorchis viverrini, promotes proliferation of host cells. PLoS Pathog 2009; 5:e1000611.
- 129. Thuwajit C, Thuwajit P, Kaewkes S, et al. Increased cell proliferation of mouse fibroblast NIH-3T3 in vitro induced by excretory/secretory product(s) from Opisthorchis Parasitology 2004;129:455-64.
- 130. Cataisson C, Salcedo R, Hakim S, et al. IL-1R-MyD88 signaling in keratinocyte transformation and carcinogenesis. J Exp Med 2012;209:1689-702.
- Iliopoulos D, Hirsch HA, Struhl K. An epigenetic switch involving NF-kappaB, Lin28, Let-7 MicroRNA, and IL6 links inflammation to cell transformation. Cell 2009;139:693-706.
- 132. Guerra C, Collado M, Navas C, et al. Pancreatitis-induced inflammation contributes to pancreatic cancer by inhibiting oncogene-induced senescence. Cancer Cell 2011;19:728-39.
- Zeng S, Yang Y, Tan Y, et al. ERBB2-induced inflammation in lung carcinogenesis. Mol Biol Rep 2012;39:7911-17.
- Ray KC, Moss ME, Franklin JL, et al. Heparin-binding epidermal growth factor-like growth factor eliminates constraints on activated Kras to promote rapid onset of pancreatic neoplasia. Oncogene 2013. [Epub ahead of print]. doi: 10.1038/onc.2013.3.
- Tye H, Kennedy CL, Najdovska M, et al. STAT3-driven upregulation of TLR2 promotes gastric tumorigenesis independent of tumor inflammation. Cancer Cell 2012;22:466-78.

- 136. Erb KJ. Can helminths or helminth-derived products be used in humans to prevent or treat allergic diseases? Trends Immunol 2009:30:75-82.
- 137. El-Malky M, Nabih N, Heder M, et al. Helminth infections: therapeutic potential in autoimmune disorders. Parasite Immunol 2011;33:589-93.
- Reddy MVR. Immunomodulators of helminthes: promising therapeutics for autoimmune disorders and allergic diseases. Indian J Clin Biochem 2010;25:109-10.
- 139. Jouvin MH, Kinet JP. Trichuris suis ova: testing a helminth-based therapy as an extension of the hygiene hypothesis. J Allergy Clin Immunol 2012;130:3-10.
- 140. Summers RW, Elliott DE, Urban Jr JF, et al. Trichuris suis therapy for active ulcerative colitis: a randomized controlled trial. Gastroenterology 2005;128:825-32.
- 141. Bager P, Kapel C, Roepstorff A, et al. Symptoms after ingestion of pig whipworm Trichuris suis eggs in a randomized placebo-controlled double-blind clinical trial. PLoS One 2011;6: e22346.
- 142. Bager P, Arnved J, Ronborg S, et al. Trichuris suis ova therapy for allergic rhinitis: a randomized, double-blind, placebo-controlled clinical trial. J Allergy Clin Immunol 2010;125:123-30.
- 143. Fleming JO, Isaak A, Lee JE, et al. Probiotic helminth administration in relapsing-remitting multiple sclerosis: a phase 1 study. Mult Scler 2011;17:743-54.
- 144. Lack G. Clinical practice. Food allergy. N Engl J Med 2008;359: 1252 - 60
- 145. Summers RW, Elliott DE, Urban Jr JF, et al. Trichuris suis therapy in Crohn's disease. Gut 2005;54:87-90.
- Summers RW, Elliott DE, Qadir K, et al. Trichuris suis seems to be safe and possibly effective in the treatment of inflammatory bowel disease. Am J Gastroenterol 2003;98:2034-41.
- 147. Cao Y, Stosiek P, Springer GF, Karsten U. Thomsen-Friedenreichrelated carbohydrate antigens in normal adult human tissues: a systematic and comparative study. Histochem Cell Biol 1996;106: 197-207.
- 148. Schmitt FC, Figueiredo P, Lacerda M. Simple mucin-type carbohydrate antigens (T, sialosyl-T, Tn and sialosyl-Tn) in breast carcinogenesis. Virchows Arch 1995;427:251-258.
- 149. Li Q, Anver MR, Butcher DO, Gildersleeve JC. Resolving conflicting data on expression of the Tn antigen and implications for clinical trials with cancer vaccines. Mol Cancer Ther 2009;8: 971 - 9.
- 150. Liu WM, Fowler DW, Gravett AM, et al. Supernatants from lymphocytes stimulated with Bacillus Calmette-Guerin can modify the antigenicity of tumours and stimulate allogeneic T-cell responses. Br J Cancer 2011;105:687-93.
- 151. Ahirwar DK, Manchanda PK, Mittal RD, Bid HK. BCG response prediction with cytokine gene variants and bladder cancer: where we are? J Cancer Res Clin Oncol 2011;137: 1729-38.
- 152. Coley WB. The treatment of malignant tumors by repeated inoculations of erysipelas. With a report of ten original cases. 1893. Clin Orthop Relat Res 1991:3–11.
- 153. Wiemann B, Starnes CO. Coley's toxins, tumor necrosis factor and cancer research: a historical perspective. Pharmacol Ther 1994; 64:529-64.
- 154. Ferlay J, Shin HR, Bray F, et al. GLOBOCAN 2008, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 10. Lyon, France: International Agency for Research on Cancer 2010.
- Bray F, Ren JS, Masuyer E, Ferlay J. Global estimates of cancer prevalence for 27 sites in the adult population in 2008. Int J Cancer 2013;132:1133-45.

