LETTER TO THE EDITOR

ASSESSMENT OF p53 OVEREXPRESSION BY NON-IMMUNOHISTOCHEMICAL METHODS

The editorials by Wynford-Thomas¹ and Hall and Lane² address some very important questions concerning the use of immunohistochemistry for the assessment of p53 protein in tumours. The accompanying papers by Baas et al.³ and Lambkin et al.⁴ further confirm that in order to obtain meaningful results with immunhistochemistry, experimental conditions such as variations in fixation, antibodies, and section pretreatment must be carefully standardized. Moreover, the interpretation of results could be subjective as only a percentage of cells stain positive for p53 in many cases. These and other shortcomings of immunohistochemical techniques have been adequately addressed in the latest editorial.²

We agree with Baas et al. that the analysis of p53 mutations at the molecular level is cumbersome, time-consuming, and generally not suitable for routine use at present. Molecular analysis, although often considered the gold standard in terms of its ability to reveal genetic, and presumably protein level, alterations, has its limitations as well, one point being that not all mutations exist in the commonly examined exons 5–9 and some occur in intronic regions. Recently, two large studies have established that p53 protein accumulation is an independent risk factor in breast carcinoma 5.6 and some other studies have presented evidence that p53 gene mutations may be prognostic of unfavourable outcome in other cancers as well.

Not mentioned in the recent editorial, however, were methods other than immunohistochemistry for the detection of p53 protein overexpression. A number of groups, including ours, have developed⁷⁻⁹ or used^{10–18} enzyme-linked immunosorbent assays (ELISAs) for p53 protein which are based on non-competitive immunoassay principles employing monoclonal and polyclonal anti-p53 antibodies. ELISAs have a number of distinct advantages over immunohistochemistry. In these procedures, in order to obtain signal, p53 must

bind to two different antibodies instead of the single p53-specific antibody used in immuno-histochemical methods, with increased antigen specificity being the expected pay-off. Moreover, endogenous peroxidase activity or biotin-binding proteins, which are sometimes present in tissues, are not a source of interference in ELISA assays due to the effective immunopurification of the antigen of interest from all other tissue components. Tissue fixation variability and pretreatment effects could also be eliminated.

From an interpretative point of view, quantitative ELISAs are more objective because the results can be evaluated using numerical cut-off values, simplifying the statistical analysis of both sample data and quality control. ELISA sensitivity is excellent and femptomole to attomole levels of analyte could be routinely measured. The extraction of cellular proteins from tumour tissue is technically simple, arguably requiring less expertise and specialized equipment than tissue sectioning. ELISAs can also be mechanized throughputs of 100-200 samples per day could be accomplished with minimal effort. Our group9 and Vojtesek et al. 10 have recently shown that p53 can be assayed in the same tumour extracts prepared for steroid hormone receptor quantification. If p53 is introduced as a routine prognostic indicator for breast cancer patients, its measurement in the same cytosolic extracts used for receptor analyses will save both labour and tumour tissue. Admittedly, one of the current limitations of p53 ELISAs is the lack of an accepted standard preparation for calibration purposes.

In our opinion, if p53 becomes the routine breast cancer prognostic indicator that a number of studies suggest, the ELISA-type assays, similar to those currently used for steroid hormone receptors, should be seriously considered for some of the advantages cited above. We propose that, in addition to immunohistochemistry, ELISA-based

methods of p53 protein detection should be used in future clinical trials examining p53 as a predictor of patient outcome in breast carcinoma.

Eleftherios P. Diamandis and Michael A. Levesque Department of Clinical Biochemistry, The Toronto Hospital, Toronto Western Division, 399 Bathurst Street, Toronto, Ontario, M5T 2S8, Canada

Addressee for correspondence: E. P. Diamandis, MD, PhD, The Toronto Hospital, Toronto, Western Division, Department of Clinical Biochemistry, 399 Bathurst Street, Toronto, Ontario, M5T 2S8, Canada.

REFERENCES

- 1. Wynford-Thomas D. p53 in tumor pathology: can we trust immunohistochemistry? *J Pathol* 1992; **166:** 329–330.
- 2. Hall PA, Lane DP. p53 in tumor pathology: can we trust immunohistochemistry?—revisited! *J Pathol* 1994; **172:** 1–4.
- Baas IO, Mulder JWR, Offerhaus JA, Vogelstein B, Hamilton SR. An evaluation of six antibodies for immunohistochemistry of mutant p53 gene product in archival colorectal neoplasms. *J Pathol* 1994; 172: 5-12.
- Lambkin HA, Mothersill CM, Kelehan P. Variations in immunohistochemical detection of p53 protein over-expression in cervical tumors with different antibodies and methods of detection. *J Pathol* 1994: 172: 13–18.
- 5. Thor AD, Moore DH, Edgerton SM, *et al.* Accumulation of the p53 tumor suppressor gene protein: an independent marker of prognosis in breast cancers. *J Natl Cancer Inst* 1992; **84:** 845–855.
- 6. Allred DR, Clark GM, Elledge R, et al. Association of p53 protein overexpression with tumor cell proliferation rate and clinical out-

- come in node-negative breast cancer. *J Natl Cancer Inst* 1993; **85:** 200–206
- 7. Midgley CA, Fisher CJ, Bartek J, Vojtesek B, Lane DP, Barnes BM. Analysis of p53 expression in human tumors: an antibody raised against human p53 expression in *Escherichia coli. J Cell Sci* 1992; **101**: 183–189.
- 8. Vojtesek B, Bartek J, Midgley CA, Lane DP. An immunochemical analysis of human nuclear phosphoprotein p53: new monoclonal antibodies and epitope mapping using recombinant p53. *J Immunol Methods* 1992; **151:** 237–244.
- Hassapoglidou S, Diamandis EP, Sutherland DJA. Quantification of p53 protein in tumor cell lines, breast tissue extracts and serum with time-resolved immunofluorometry. *Oncogene* 1993; 8: 1501– 1509
- 10. Vojtesek B, Fisher CJ, Barnes DM, Lane DP. Comparison between p53 staining in tissue sections and p53 protein levels measured by an ELISA technique. *Br J Cancer* 1993; **67**: 1254–1258.
- 11. Bartek J, Bartkova J, Vojtesek B, *et al.* Aberrant expression of the p53 oncoprotein is a common feature of a wide spectrum of human malignancies. *Oncogene* 1991; **6:** 1699–1703.
- 12. Tominaga O, Hamelin R, Trouvat V, et al. Frequently elevated content of immunochemically defined wild-type p53 protein in colorectal adenomas. Oncogene 1993; 8: 2653–2658.
- 13. Bartkova J, Bartek J, Vojtesek B, *et al.* Immunochemical analysis of the p53 oncoprotein in matched primary and metastatic human tumors. *Eur J Cancer* 1993; **29A**: 881–886.
- Yewdell JW, Gannon JV, Lane DP. Monoclonal antibody analysis of p53 expression in normal and transformed cells. *J Virol* 1986; 59: 444–452.
- Remvikos Y, Tominaga O, Hammel P, et al. Increased p53 protein content of colorectal tumors correlates with poor survival. Br J Cancer 1992; 66: 758-764.
- Brown R, Clugston C, Burns P, et al. Increased accumulation of p53 protein in cisplatin-resistant ovarian cell lines. Int J Cancer 1993; 55: 678–684
- 17. Lu X, Lane DP. Differential induction of transcriptionally active p53 following UV or ionizing radiation: defects in chromosome instability syndromes? *Cell* 1993; **75**: 765–778.
- 18. Winter SF, Minna JD, Johnson BE, Takahashi T, Gazdar AF, Carbone DP. Development of antibodies against p53 in lung cancer patients appears to be independent on the type of p53 mutation. *Cancer Res* 1992; **52:** 4168–4174.

AUTHOR'S REPLY

Diamandis and Levesque draw attention to the potential value of ELISA assay as a clinical diagnostic tool for the assessment of aberrant p53 expression in breast cancer. The authors correctly point out the desirability of having a quantitative measurement of protein content which would be more objective than immunocytochemistry (ICC). Their arguments could, however, have been more balanced.

For example, one of the advantages that they claim for ELISA is its avoidance of the major uncertainties which result from tissue fixation. While this is of course true, it is the use of frozen tissue which is important, not the technique of analysis—immunocytochemistry is equally free of this problem on frozen tissue! More importantly,

they fail to mention a major drawback of ELISA compared to ICC, which is its inability to take account of variations in the proportion of malignant cells present in the sample, which, according to the Vojtesek study, can vary at least three-fold. This is, of course, simply a new example of the longstanding dispute between immunocytochemical and biochemical assessment, well illustrated by the problem of oestrogen receptor analysis in breast cancer.

There is no doubt that accurate quantitation of a cellular protein in clinical samples will always be problematic. Ultimately, most clinicians will want a technique which will identity cases having a level of p53 expression associated with poor prognosis. The accuracy required of any routine method will depend, therefore, on the degree of overlap or separation between this sub-set and the 'normal' range. Classical immunocytochemistry (without antigen retrieval) may turn out to be an effective way of making this discrimination. Alternatively, the best compromise may be to use ELISA as the primary measurement, but to control for cellularity as far as possible by visual assessment of parallel histological sections. However, until we know more from research studies about the range and significance of p53 expression levels in breast pathology, it is probably premature to make any

firm recommendations for methods of routine clinical assessment.

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D. Wynford-Thomas CRC Thyroid Tumour Biology Research Group, Department of Pathology, University of Wales College of Medicine, Heath Park, Cardiff, CF4 4XN, U.K.

REFERENCE

1. Vojtesek B, Fisher CJ, Barnes DM, Lane DP. Comparison between p53 staining in tissue sections and p53 protein levels measured by an ELISA technique. *Br J Cancer* 1993; **67:** 1254–1258.