

Multiple human tissue kallikreins are regulated by lympho-epithelial Kazal-type inhibitor and digest desmoglein 1

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Background

- Stratum corneum (SC) desquamation requires proteolysis of desmosomes by trypsin-like and chymotrypsin-like serine proteases (SP)¹
- Serine protease inhibitors (SPI) regulate SC desquamation as evidenced by the SP/SPI imbalance observed in Netherton syndrome (NS)^{2,3,4}
 - NS is caused by nonsense mutations in *serine protease inhibitor Kazal-type 5* (*SPINK5*; encodes lympho-epithelial Kazal-type inhibitor (LEKTI))
 - The decrease in LEKTI expression/activity leads to unopposed SP activity resulting in over-desquamation
- Among SC proteases implicated in desquamation are human tissue kallikreins (hKs), 15 secreted SP with trypsin-like and/or chymotrypsin-like specificity⁵
- Most kallikreins (i.e. kallikreins 1, 4-8, 10, 13, 14) are expressed in the stratum granulosum (SG) at the mRNA and/or protein level^{6,7} within lamellar granules (e.g. hK5, hK7, hK8)^{8,9} and secreted to the intercellular spaces of the SC
- To date, hK5 and hK7 have been shown to digest desmosomal proteins¹⁰:
 - corneodesmosin (CDSN)
 - desmocollin 1 (DSC1)
 - desmoglein 1 (DSG1)
- hK5 and hK7 are inhibited by SPI that co-localize to the SG and SC^{11,12,13}:
 - Lympho-epithelial Kazal-type inhibitor (LEKTI)
 - Secretory leukocyte protease inhibitor (SLPI)
 - Elafin/protease inhibitor 3
 - α_2 -macroglobulin like-1 (A2M1)

Materials & Methods

- Materials:** 7-Amino-4-methylcoumarin (AMC) peptide substrates Boc-Val-Pro-Arg-AMC, H-Pro-Phe-Arg-AMC, Boc-Gln-Ala-Arg-AMC were purchased from Bachem Bioscience and MetSas-His-Ala-Pro-Val-AMC was obtained from Calbiochem. Recombinant mature hK1 and hK6 were expressed and purified from a baculovirus-infected cell line system as previously described¹⁴. Recombinant pro-hK13 and mature hK13 and hK14 were produced in the *Pastorius* expression system as described in detail elsewhere^{15,16}. Recombinant desmoglein 1 and desmoglein 2 were purchased from Sigma-Aldrich. Recombinant hK13 and hK14 containing intact domains 1-6 (LEKTI1-6), 6-8 and partial domain 9 (rLEKTI6-9), 9-12 (rLEKTI9-12) and 12-15 (rLEKTI12-15) were produced in a baculovirus-infected cell line system as previously reported¹⁷. Recombinant SLPI and a recombinant DSG1Fc chimera were obtained from BD Biosciences Inc. Anti-LEKTI monoclonal antibody 1C11G6 was produced as previously described¹⁸.
- Kinetic inhibition assays:** Individual hKs (120nM of hK1, hK2, hK5, hK13 and hK14 or 300nM of hK6) were pre-incubated with varying concentrations of LEKTI fragments (0-400nM) for 10 min and diluted (1:10) in 10 μ l of optimal buffer for different time points. The mixtures were subsequently added to 90 μ l of optimal buffer containing several fixed AMC peptide concentrations (4-300nM) within microtitre plate wells. Initial rates of hK-mediated peptide hydrolysis were monitored by measuring free AMC fluorescence on the Wallac 1420 Victor fluorometer with excitation and emission filters of 360nm and 480nm, respectively. Assays were expressed relative to control incubations from which inhibitors were excluded. Michaelis-Menten parameters (K_m , V_{max}), the equilibrium inhibition constant (K_i) and inhibitory mechanisms were determined by linear and non-linear regression analysis using the Enzfitter Kinetics Module 1.1.
- In vitro digestion experiments:** Individual hKs were incubated with SLPI, rLEKTI fragments and DSG1Fc at 37°C. Reactions were monitored over time by reducing SDS-PAGE followed by Silver-staining the SLPI and DSG1Fc or transferred to nitrocellulose membranes and probed with anti-LEKTI mAb 1C11G6 for rLEKTI fragments.
- N-terminal sequencing:** N-terminal sequence analysis was performed to identify hK cleavage sites within SLPI and DSG1Fc. MS (500ng) were incubated with LEKTI, SLPI or DSG1 (0.5 μ g), separated by SDS-PAGE, transferred to a polyvinylidene difluoride membrane and stained with Coomassie Brilliant Blue R-250. Silver-stained DSG1Fc fragments from a polyvinylidene difluoride membrane were subjected to Edman degradation consisting of 5 cycles of Edman chemistry on a ProteinBotmer Gas-phase Microsequencer. Following by phenylthiohydantoin (PTH) analysis on an HPLC column.

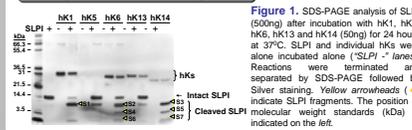
Results

I. Secretory leukocyte protease inhibitor (SLPI) and Elafin do not inhibit hKs

Protease (nM)	Molar ratio (Protease:SLPI)	SLPI (nM)	Residual Activity (%)	Inhibition (%)
Elastase (18)	1:5	90	0.2	99.8
hK1 (12)	1:100	1200	100.0	-0.0
hK5 (12)	1:100	1200	100.3	-0.6
hK6 (12)	1:100	1200	94.0	6.0
hK13 (12)	1:100	1200	120.0	-20.0
hK14 (12)	1:100	1200	91.9	8.1

Protease (nM)	Molar ratio (Protease:Elafin)	Elafin (nM)	Residual Activity (%)	Inhibition (%)
Elastase (12)	1:2	24	0.2	99.8
hK1 (12)	1:100	103.2	100.3	-3.2
hK5 (12)	1:100	1200	102.4	-2.4
hK6 (12)	1:100	1200	100.3	-0.3
hK13 (12)	1:100	1200	101.4	-1.4
hK14 (12)	1:100	1200	99.6	0.4

II. Digestion of SLPI by hKs



hK	hK-generated SLPI fragment	N-terminal sequence	hK cleavage site	Location of cleavage site within SLPI
hK1, hK6, hK14	S12/S3/4/5	Y ¹ KKPP	R ² Q ³ Y ⁴ P ⁵	1 st WAP ² domain
hK6, hK14	S6/7	M ¹ A ² L ³ N ⁴ P ⁵ P ⁶	R ⁷ Q ⁸ M ⁹ T ¹⁰	2 nd WAP ² domain

III. Digestion of rLEKTI by hKs

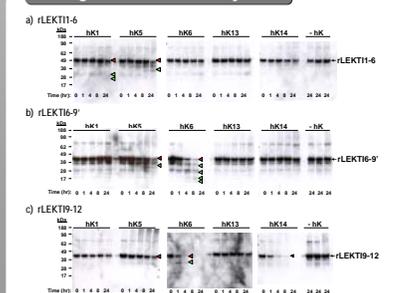


Figure 2. Immunodetection of a) rLEKTI1-6, b) rLEKTI6-9 and c) rLEKTI9-12 after incubation with individual hKs (10ng) were incubated with hKs (1ng) in hK reaction buffer for various time points (0, 4, 8, 24 hours) at 37°C. Reactions were terminated and separated by SDS-PAGE followed by Western blotting using anti-LEKTI mAb 1C11G6. rLEKTI fragments were incubated alone for 24 hours in each hK reaction buffer ('hK- lanes'). Red arrowheads (♦) indicate intact rLEKTI fragments; green arrowheads (♦) denote degraded rLEKTI fragments. The position of molecular weight standards (kDa) is indicated on the left.

IV. rLEKTI inhibition of hKs

hK	Mechanism	rLEKTI-6 (nM)	V_{max} (nmol L ⁻¹ min ⁻¹)	K_m (nM)	K_i (nM)	R ²
hK1	No inhibition	0	2884 ± 35	0.46 ± 1.91E-02	-	-
hK5	Mixed	12	1140 ± 18	0.93 ± 4.00E-02	2.35 ± 0.22	0.99
		24	1029 ± 24	1.06 ± 3.16E-02	-	-
		60	350 ± 15	3.72 ± 0.26	-	-
hK6	Non-competitive	12	912 ± 9	0.23 ± 2.12E-02	21.59 ± 1.24	0.97
		24	653 ± 11	0.29 ± 1.94E-02	-	-
		60	580 ± 9	0.33 ± 2.03E-02	-	-
hK13	Mixed	12	266 ± 6	0.43 ± 2.97E-02	24.13 ± 3.82	0.98
		24	777 ± 12	0.40 ± 2.95E-02	-	-
		60	712 ± 10	0.57 ± 2.50E-02	-	-

hK	Mechanism	rLEKTI6-9 (nM)	V_{max} (nmol L ⁻¹ min ⁻¹)	K_m (nM)	K_i (nM)	R ²
hK1	No inhibition	0	2879 ± 34	0.44 ± 2.06E-02	-	-
hK5	Mixed	12	1640 ± 20	0.67 ± 2.68E-02	4.68 ± 0.66	0.98
		24	1079 ± 24	1.41 ± 2.71E-02	-	-
		60	451 ± 21	2.96 ± 2.37E-02	-	-
hK6	Non-competitive	12	1186 ± 35	0.44 ± 2.13E-02	47.58 ± 1.40	0.99
		24	795 ± 13	0.32 ± 4.7E-02	-	-
		60	501 ± 4	0.32 ± 3.47E-02	-	-
hK13	Non-competitive	12	1069 ± 24	0.51 ± 3.83E-02	22.12 ± 9.51	0.98
		24	905 ± 20	0.42 ± 3.44E-02	-	-
		60	757 ± 12	0.51 ± 2.68E-02	-	-
hK14	Non-competitive	12	4196 ± 23	0.25 ± 0.07E-02	10.26 ± 1.25	0.98
		24	1578 ± 24	0.18 ± 1.13E-02	-	-
		60	171 ± 3	0.32 ± 1.78E-02	-	-

Table 6. Inhibitory profile of rLEKTI9-12

hK	Mechanism	rLEKTI9-12 (nM)	V_{max} (nmol L ⁻¹ min ⁻¹)	K_m (nM)	K_i (nM)	R ²
hK1	No inhibition	0	2795 ± 33	0.44 ± 1.87E-02	-	-
hK5	Mixed	12	1144 ± 26	0.75 ± 4.8E-02	2.75 ± 0.24	0.99
		24	823 ± 24	1.64 ± 1.10E-02	-	-
		60	707 ± 62	5.09 ± 4.44E-02	-	-
hK6	Non-competitive	12	937 ± 16	0.14 ± 1.68E-03	195.32 ± 11.66	0.99
		24	698 ± 11	0.24 ± 1.76E-02	-	-
		60	593 ± 8	0.25 ± 1.77E-02	-	-
hK13	Non-competitive	12	1054 ± 19	0.40 ± 2.04E-02	408.63 ± 16.47	0.99
		24	868 ± 11	0.40 ± 2.04E-02	-	-
		60	832 ± 17	0.52 ± 3.41E-02	-	-
hK14	Mixed	12	3480 ± 34	0.18 ± 0.07E-03	10.26 ± 1.25	0.98
		24	2608 ± 61	0.23 ± 1.57E-02	-	-
		60	1901 ± 20	0.38 ± 1.08E-02	-	-

Table 7. Inhibitory profile of rLEKTI12-15

hK	Mechanism	rLEKTI12-15 (nM)	V_{max} (nmol L ⁻¹ min ⁻¹)	K_m (nM)	K_i (nM)	R ²
hK1	No inhibition	0	6279 ± 113	0.27 ± 0.01E-05	-	-
hK5	Mixed	12	5991 ± 77	0.32 ± 0.01E-02	21.80 ± 2.40	0.99
		24	5598 ± 92	0.47 ± 0.02E-02	-	-
		60	4796 ± 66	0.65 ± 0.02E-02	-	-
hK6	No inhibition	0	-	-	-	-
hK13	No inhibition	0	-	-	-	-
hK14	No inhibition	0	-	-	-	-

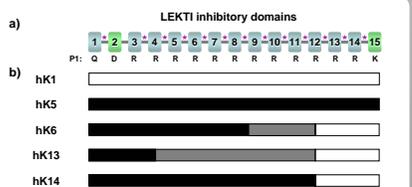


Figure 3. Summary of rLEKTI inhibition of hKs. a) Schematic of LEKTI inhibitory domains (Genbank accession no. NP_006837). Blue boxes denote Kazal-type inhibitory domains; green boxes represent non-Kazal-type inhibitory domains. The symbol indicates the location of putative pro-peptide conversion motifs (P1-KR4). The identity of the amino acid at the P1 position is indicated below each domain. b) Boxes indicate strength of rLEKTI inhibition of individual hKs based on K_i values. White boxes indicate no inhibition; grey boxes indicate moderate inhibition ($K_i > 200$ nM); black boxes represent strong inhibition ($K_i < 50$ nM).

V. Digestion of DSG1 by hKs

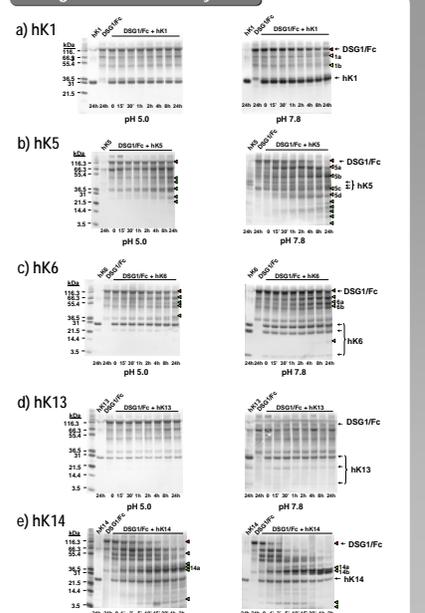


Figure 4. SDS-PAGE analysis of DSG1Fc after incubation with a) hK1, b) hK5, c) hK6, d) hK13 and e) hK14 at pH 5.0 and 7.8. DSG1Fc (500ng) was incubated with hKs (50ng) in hK reaction buffer for various time points at 37°C. Reactions were terminated and separated by SDS-PAGE followed by Silver-staining. DSG1Fc and individual hKs were incubated alone for 24 hours in each hK reaction buffer (lanes 1 and 2). Red arrowheads (♦) indicate intact DSG1Fc; green arrowheads (♦) denote DSG1Fc fragments; yellow arrowheads (♦) indicate DSG1Fc fragments set for N-terminal sequencing. The position of molecular weight standards (kDa) is indicated on the left. Note: DSG1Fc is a chimeric protein formed of the 4 extracellular cadherin followed by a peptide linker and 2 immunoglobulin constant region domains.

Table 8. hK cleavage sites within DSG1

hK	hK-generated DSG1Fc fragment	N-terminal sequence	hK cleavage site ¹	Location of cleavage site within DSG1
hK1	1a	I ¹ RR ² ROEP	K ¹⁹⁷ L ¹⁹⁸	2 nd CAD ² domain
hK5	5a/b	D ⁴ N ⁵ KVTKK	R ⁴⁴² N ⁴⁴⁶	4 th CAD domain
	5c	D ⁴ H ⁵ GADG	K ⁴²² D ⁴²³	2 nd CAD domain
	5b	Y ⁴²² V ⁴²³ MGNNG	R ⁴²² L ⁴²³	4 th CAD domain
hK6	6a	Y ¹⁴⁶ L ¹⁴⁷ DIND	R ¹⁴⁶ V ¹⁴⁷	1 st CAD domain
	6b	A ³⁶⁵ K ³⁶⁹ Y	K ³⁶⁸ A ³⁶⁹	3 rd CAD domain
hK14	14a/b	S ²⁸ P ²⁹ SEPGN	R ⁴²² D ⁴²³	4 th CAD domain
			Y ²⁸ S ²⁹	Between 4 th CAD domain & TM ³ fragment

1. Numbering starts from desmoglein 1 propeptide sequence (Genbank accession no. NP_001933)
2. CAD: cadherin domain. DSG1 contains 4 extracellular CAD domains
3. TM: transmembrane region

Summary

- Multiple hKs represent specific targets of LEKTI, but not of SLPI or elafin
 - rLEKTI1-6, 6-9 and 9-12 strongly inhibit hK5 and hK14 and to a slightly lesser extent hK6 and hK13, but do not affect hK1 activity
 - rLEKTI12-15 only inhibits hK5 and not other hKs studied
- rLEKTI6-9 is degraded by hK6 and rLEKTI9-12 by hK6 and hK14, which may lead to inactivation
- hK6 and hK14 may inactivate SLPI via cleavage at L⁷²
- Desmoglein 1 is a potential *in vivo* substrate of hK5, hK6 and hK14, but not of hK1 or hK13

Conclusion

In addition to hK5 and hK7, other hKs including hK6, hK13 and hK14 are implicated in SC desquamation due to their specific inhibition by LEKTI and/or digestion of desmoglein 1

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