Essential roles for the splicing regulator nSR100/SRRM4 during nervous system development

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Alternative splicing (AS) generates vast transcriptomic complexity in the vertebrate nervous system. However, the extent to which trans-acting splicing regulators and their target AS regulatory networks contribute to nervous system development is not well understood. To address these questions, we generated mice lacking the vertebrate- and neural-specific Ser/Arg repeat-related protein of 100 kDa (nSR100/SRRM4). Loss of nSR100 impairs development of the central and peripheral nervous systems in part by disrupting neurite outgrowth, cortical layering in the forebrain, and axon guidance in the corpus callosum. Accompanying these developmental defects are widespread changes in AS that primarily result in shifts to nonneural patterns for different classes of splicing events. The main component of the altered AS program comprises 3- to 27-nucleotide (nt) neural microexons, an emerging class of highly conserved AS events associated with the regulation of protein interaction networks in developing neurons and neurological disorders. Remarkably, inclusion of a 6-nt, nSR100-activated microexon in Unc13b transcripts is sufficient to rescue a neuritogenesis defect in nSR100 mutant primary neurons. These results thus reveal critical in vivo neurodevelopmental functions of nSR100 and further link these functions to a conserved program of neuronal microexon splicing.

Keywords: alternative splicing; SR proteins; nervous system development; microexons

Supplemental material is available for this article.

Received November 18, 2014; revised version accepted February 27, 2015.

Proper development and function of the mammalian nervous system depend on the tight coordination of multiple layers of gene regulation. During development, neurons progress through maturation stages to acquire their subtype-specific functions (Gao et al. 2013). For example, neurons born in the subventricular zone (SVZ) of the embryonic cortex use endocrine and exocrine cues while migrating dorsally to establish and populate specific cortical layers. Similarly, neuronal projections in the brain and periphery rely on successive adjustments of intrinsic to extrinsic factors to arrive at their targets. Considerable remodeling of the cytoskeleton, vesicular transport, and other subcellular processes allows neurons to achieve their designated morphologies and functions. While similar repertoires of genes are associated with these processes, it is becoming apparent that extensive variation at the level of post-transcriptional regulation generates the remarkable transcriptomic and proteomic diversity required for establishing biological complexity during vertebrate nervous system development (Li et al. 2007; Norris and Calarco 2012; Lipscombe et al. 2013a; Zheng and Black 2013).

Alternative splicing (AS) is the process by which different pairs of splice sites are selected to produce multiple transcripts from a single gene. It is controlled by the concerted action of multiple cis-acting motifs and cognate trans-acting factors that promote or repress the assembly of productive splicing complexes (spliceosomes) at splice sites (Chen and Manley 2009; Braunschweig et al. 2013). Widely expressed and tissue-restricted RNA-binding proteins combine to regulate AS decisions via positive- and negative-acting cis motifs located in exons or flanking introns, referred to as splicing enhancers and silencers, respectively. AS represents a major source of biological diversity that likely afforded the evolution of complexity associated with the development and function of the vertebrate nervous system (Barbosa-Morais et al. 2012;
Indeed, AS patterns are more complex in the brain than other tissues, and many of these events happen in genes implicated in complex neuronal processes, such as the control of synaptic plasticity associated with cognition (Lipscombe 2005; Ule and Darnell 2006). While most tissue differential splicing patterns are species-specific in vertebrates, there is a higher frequency of conserved alternative cassette exon inclusion events in vertebrate brains than in other tissue types [Barbosa-Morais et al. 2012; Merkin et al. 2012]. This suggests the existence of a core set of conserved functions for AS across vertebrate species in addition to roles for AS underlying species-specific neurodevelopmental and behavioral characteristics. However, little is known about the in vivo functions of the protein factors that are responsible for establishing AS complexity in the nervous system or the functions of the individual AS events that are controlled by these factors.

Neural-enriched splicing regulators, including the Nova, Rboh, and Ptbp proteins, have been characterized using mouse models. Nova proteins, which were originally identified as the autoantigens in patients with paraneoplastic opsoclonus myoclonus ataxia [Buckanovich et al. 1993; Yang et al. 1998], control the inhibitory synapse, and their knockout results in cortical migration [Yano et al. 2010] and neuromuscular junction (NMJ) defects [Ruggiu et al. 2009]. Rboh1 and Rboh2 mutant mice are susceptible to seizures and display disrupted cerebellar development [Gehman et al. 2012]. Depending on the strain background, Ptb2 knockout mice die at birth or else exhibit cortical degeneration and lethality during the first few postnatal weeks [Licatalosi et al. 2012; Li et al. 2014]. Additional studies using Nova knockout mice have revealed functions for specific Nova-regulated splice variants [including alternative exons in the Dab1 gene] that facilitate the proper migration of newly born cortical neurons [Yano et al. 2010] and exons in the Agrin gene that are important for the formation of NMJs [Ruggiu et al. 2009]. However, aside from these examples, few other neuronal genes have been characterized at isoform resolution in vivo [Norris and Calarco 2012; Lipscombe et al. 2013b; Zheng and Black 2013].

We previously identified and characterized the vertebrate- and neural-specific Ser/Arg repeat-related protein of 100 kDa [nSR100/SRRM4] [Calarco et al. 2009; Raj et al. 2011, 2014]. Knockdown and overexpression experiments performed in cell culture revealed that nSR100 promotes the inclusion of 30%–50% of the conserved human and mouse cassette alternative exons that display brain-specific inclusion patterns in transcriptome profiling data [Raj et al. 2014]. Knockdown of nSR100 in Neuro2a cells and developing zebrafish was shown to impair neurite outgrowth and branching of trigeminal ganglia, respectively [Calarco et al. 2009], and in utero knockdown of nSR100 in mice prevented differentiation of neuronal progenitors in the cortex [Raj et al. 2011]. Recently, the Bronx waltzer (bv) mouse mutation was mapped to the nSR100 gene [Nakano et al. 2012]. bv homozygotes display hearing and balance defects attributed to degeneration of inner ear hair cells. The apparent limited pheno-
cotypic consequences of the bv mutation are likely because this mutation eliminates only the terminal exon and part of the 3′ untranslated region (UTR) of nSR100 transcripts, leaving the majority of the nSR100 protein intact.

nSR100-regulated exons were found to be concentrated in genes that function in various aspects of neuronal development and function [Calarco et al. 2009; Raj et al. 2011, 2014; Nakano et al. 2012]. These and other neural-regulated exons that are >27 nucleotides (nt) in length are highly concentrated in surface-accessible disordered regions of proteins and function in the regulation of protein–protein interactions [Buljan et al. 2012; Ellis et al. 2012]. Furthermore, in a very recent study, we showed that nSR100 strongly promotes the inclusion of very short, 3′- to 27-nt, neuronal “microexons” [Irimia et al. 2014]. The corresponding microexon residues are concentrated within—or immediately adjacent to—protein–protein or protein–lipid interaction domains. Most of these exons display striking increases in inclusion during neuronal maturation, coincident with increased expression of nSR100. Notably, they also show significant decreases in inclusion—coincident with reduced expression of nSR100—in the cortices of individuals with autism spectrum disorder (ASD) [Irimia et al. 2014]. A key function of nSR100 thus appears to be the widespread regulation of protein interactions required for the maturation and proper function of neurons. However, the scope of the in vivo functions of nSR100 during nervous system development has not been previously addressed.

To investigate the functions of nSR100 in vivo, we generated mice carrying a conditional exon deletion in the nSR100 [Srrm4] gene that results in widespread loss of the full-length protein. We observed that nSR100 is essential for early postnatal survival of a large majority of mutant animals, with the few surviving animals displaying balance defects similar to those seen in bv/bv mice but also exhibiting persistent tremors. Additionally, loss of nSR100 in mice results in impaired neurite outgrowth in the diaphragm, early neuronal commitment of neural progenitors leading to defective cortical layering, and a failure of callosal axons to cross the midline in the forebrain. Using RNA sequencing [RNA-seq] profiling, we defined all classes of AS [including alternative microexons and longer alternative cassette exons, 5′ and 3′ splice sites, and retained introns] that are controlled by nSR100 in vivo, a great majority of which were not previously reported. A large fraction of alternative cassette exons and microexons positively regulated by nSR100 are neurally enriched, which is not the case for other classes of nSR100-dependent splicing events. Moreover, a higher proportion of neural microexons is affected by disruption of nSR100 than are other neural-regulated AS events. These include highly conserved exons with the potential to insert only one or two amino acids in proteins of key functional relevance to neuronal maturation. Remarkably, an nSR100-regulated 6-nt microexon in the Unc13b gene promotes neurite growth in mouse primary neurons. Cortical neurons from nSR100Δ7–8/Δ7–8 mice display a neuritogenesis defect, and expression of Unc13b transcripts including
the microexon, but not transcripts lacking the microexon, is sufficient to rescue the mutant phenotype. Collectively, our results define critical new in vivo functions of nSR100 during mouse development and further link these functions to the disruption of a conserved program of nSR100-dependent neuronal microexons.

Results

Perinatal mortality in nSR100 mutant mice

Our previous studies using in vivo knockdown of nSR100 in zebrafish and mouse embryos suggested that nSR100 may play a role in several aspects of nervous system development (Calarco et al. 2009; Raj et al. 2011). To address the extent of nSR100 functions in the developing nervous system, we generated mice carrying a conditional knock-out nSR100lox allele from embryonic stem cells obtained from EUCOMM (European Conditional Mouse Mutagenesis Program). The nSR100lox allele includes a gene trap upstream of a LacZ reporter and LoxP sites framing nSR100 exons 7 and 8 [Fig. 1A]. Southern blotting confirmed the integrity of the integration site in nSR100lox mice [Fig. 1B]. We crossed nSR100lox/lox mice with mice carrying the widely expressed CMV-Cre recombinase transgene to obtain nSR100α7−8 mice, in which exons 7 and 8 have been deleted throughout the animal and in the germline. This deletion introduces a +2 frameshift in downstream exons and causes complete loss of full-length nSR100 transcripts and protein in homozygous nSR100lox−8 mice [Fig. 1C,D]. Immunoblotting revealed that a 25-kDa protein fragment could be detected in homozygous and heterozygous mutant mice using an antibody to the N terminus of nSR100. RT–PCR confirmed that transcripts encompassing nSR100 exons 1–6 and the gene trap insertion are preserved in the mutant mouse [Supplemental Fig. S1A]. The small N-terminal fragment produced from the nSR100α7−8 allele lacks the RS-rich domain of nSR100 [Supplemental Fig. S1B], which, based on previous studies of nSR100 and other SR proteins, is predicted to function in the formation of protein–protein and/or protein–RNA interactions required for splicing complex formation [Wu and Maniatis 1993; Shen and Green 2004; Raj et al. 2014]. In contrast to the full-length protein, overexpression of the truncated protein in Neuro2a cells depleted of endogenous nSR100 fails to restore nSR100-dependent splicing [Supplemental Fig. S1C, lanes 1–3]. Moreover, when coexpressed with full-length nSR100 in Neuro2a cells, the truncated mutant does not interfere with splicing of nSR100 target exons [Supplemental Fig. S1C, lanes 4–6]. These observations provide evidence that the 25-kDa protein fragment lacks critical functional activities associated with full-length nSR100.

We observed that >85% of nSR100α7−8/α7−8 mice died in the first few hours after birth [Supplemental Table S1]. Although these mice present no gross morphological phenotype at late embryonic stages or at birth [Fig. 1E], they show signs of respiratory defects, including irregular breathing and heavy gasping, and become cyanotic soon after birth. This phenotype contrasts sharply with those conditional allele in intron 3. Predicted band size is 15.4 kb in wild-type, 16.4 kb in conditional, and 19.4 kb in knockout alleles, respectively. (C) RT–PCR on embryonic day 16.5 (E16.5) whole-brain total RNA using primers amplifying exon 2 to exon 9. No transcript could be detected in homozygous mutants. (D) Immunoblotting on E17.5, whole-brain lysates using an antibody to nSR100. Full-length nSR100 protein is completely lost in homozygous mutants [arrow], but a 25-kDa fragment is expressed from the α7−8 allele. (E) E17.5 mutant embryos display normal morphology.
of the previously described nSR100 mutant bv mouse, in which only the last 103 amino acids from the C terminus of nSR100 are lost. Homozygous bv mice are viable and display a phenotype limited to the degeneration of the inner hair cells of the inner ear [Deol and Gluecksohn-Waelsch 1979; Nakano et al. 2012]. Notably, the few homozygous nSR100Δ7–8/Δ7–8 survivors that we obtained from crossing heterozygous parents display a head tilting and circling behavior reminiscent of the balancing defect observed for the bv mutant strain. However, in contrast to the bv mutant, all surviving nSR100Δ7–8/Δ7–8 individuals additionally display pronounced tremors, a phenotype that is often associated with neurobiological defects [Supplemental Videos S1–S3]. Homozygous mutants escaping perinatal mortality did not display significant differences in life span compared with wild-type littermates [data not shown]. Embryos harvested at embryonic day 17.5 (E17.5) and E18.5 were found at Mendelian ratios [Supplemental Table S1], indicating that loss of nSR100 does not cause early embryonic lethality. The extensive perinatal mortality and neurobiological phenotypes observed in surviving nSR100Δ7–8/Δ7–8 mice highlight the importance of nSR100 during embryonic development as well as its role in the proper functioning of the adult nervous system.

Loss of nSR100 impairs diverse neuronal processes

The respiratory problems accompanying perinatal mortality in nSR100Δ7–8/Δ7–8 mice suggested that the innervation of the diaphragm might be impaired by loss of the nSR100 protein. We first asked whether nSR100 is expressed in the peripheral nervous system where motor neurons innervating the diaphragm are located. We surveyed nSR100 expression at different time points during development using both the LacZ cassette in the nSR100lox mouse as a reporter for nSR100 gene expression and in situ RNA hybridization in wild-type mice. X-Gal staining and in situ hybridization showed that nSR100 is expressed in both the brain and the neural tube during early neurogenesis, with nSR100 mRNA expressed as early as E8.5 and LacZ reporter expression starting at E9.5 [Supplemental Figs. S2A,B, S3A]. Immunostaining of sections from nSR100+/lox mice using anti-β-galactosidase antibody and anti-NeuN antibody to mark post-mitotic neurons revealed that only neurons express nSR100 in the brain [Supplemental Fig. S3B]. Moreover, in situ hybridization at E17.5 showed that nSR100 expression is maintained in the brain during development, with high expression in the cerebral cortex and hippocampus late in embryogenesis [Supplemental Fig. S2C]. These results are consistent with recent analyses of RNA-seq data from diverse human and mouse cell and tissue types and a neuronal differentiation time series, which indicated that nSR100 expression is neuron-specific, occurs in the brain and dorsal root ganglia, and increases in the brain from E11 to E18 before decreasing in the adult [Irimia et al. 2014; Raj et al. 2014]. Taken together, these data confirm that nSR100 is neuronal-specific and is expressed in both the central and peripheral nervous system in developing mice.

We next visualized the innervation of the diaphragm just before birth at E18.5 using an antibody to neurofilament on whole-mount preparations. This staining revealed that primary branches deriving from the phrenic nerve appear thinner in nSR100Δ7–8/Δ7–8 mice (Fig. 2A). In addition, we observed that the total length covered by secondary motor axons is greatly reduced and that the number of secondary axons is decreased by almost twofold in homozygous mutants, a phenotype not seen in heterozygotes [Fig. 2B,C]. These defects are already present at E16.5 [Supplemental Fig. S4A–C], suggesting that the lack of secondary branches does not stem from degeneration or pruning but rather from deficient sprouting in the mutant mice. The overall distance covered by primary axons was not affected at either E18.5 or E16.5 [Fig. 2D; Supplemental Fig. S4D]. Each individual secondary branch forming in the mutants projects as far as its wild-type

**Figure 2.** Loss of nSR100 impairs neurite outgrowth in motor neurons. (A) Whole-mount staining of E18.5 diaphragms with anti-neurofilament antibody (green) to highlight innervation. Orange dots mark secondary branches in the insets. Bars: left panels, 1000 μm; insets, 500 μm. [B,C] The total distance covered by all secondary branches [B] and the number of secondary branches present on the right ventral primary branch of the phrenic nerve [C] were quantified on three or four individuals for each genotype. The total distance covered by secondary neurites and the number of secondary branches formed are significantly lower in homozygous mutants. [D] The total length covered by primary branches is not affected in homozygous mutants. [E] The average length of individual secondary branches in the mutant does not differ significantly from those of wild-type and heterozygous littermates. Three diaphragms from wild-type and heterozygous embryos and four diaphragms for homozygous mutants were analyzed. One-way ANOVA with Tukey-Kramer post-hoc test. Error bars indicate standard error of the mean.
counterpart (Fig. 2E; Supplemental Fig. S4E), and motor endplates form in the same numbers in the diaphragm of nSR100 homozygous and heterozygous mutant mice, although at higher density in the homozygous mutant, most likely due to a lack of secondary branching [Supplemental Fig. S4F–I]. The diminished axon sprouting capacities of motor neurons in the diaphragm of nSR100Δ7–8/Δ7–8 mice likely contributes to nSR100-dependent respiratory defects and early postnatal death. These axon guiding or branching defects are not limited to phrenic nerve innervation, as we detected defective formation of the trigeminal, hypoglossal, and spinal nerves in whole-mount staining of E10.5 and E12.5 embryos (Supplemental Fig. S5).

Because nSR100 is highly expressed in the cortex and in utero knockdown of nSR100 resulted in defects in neuronal differentiation in the cortex (Raj et al. 2011), we investigated whether cortical anatomy was altered in nSR100 mutants. The establishment of defined cortical layers is an important and conserved step in mammalian brain development. Immunofluorescence using layer-specific markers revealed that the deep, T-box brain 1 (Tbr1)-positive cortical layer VI is enlarged and comprised of more cells in the homozygous mutant, a phenotype also seen to a lesser extent in heterozygotes (*P* < 0.0001, one-way ANOVA) [Fig. 3A,B]. The definition of the preplate was also altered in homozygotes and heterozygotes. Costaining with antibodies to Tbr1 and Satb2 to highlight superficial layers II–V revealed a decrease in the number of superficial neurons in nSR100 mutants (*P* < 0.05, one-way ANOVA) [Fig. 3A,C]. The total number of post-mitotic neurons was also reduced in the mutants, as highlighted by NeuN staining (*P* < 0.0001, one-way ANOVA) [Fig. 3A,D].

We postulated that the increase in the number of early-born neurons and decrease in the number of late-born neurons observed in nSR100 mutants may be due to the premature commitment of neural progenitors to a post-mitotic fate. Indeed, Pax6 staining at E18.5...
confirmed that the pool of neural progenitors is depleted in nSR100Δ7-8/Δ7-8 mice at the end of embryogenesis (P < 0.05, one-way ANOVA) [Fig. 3A,E]. To verify whether deregulation of neurogenesis was the major cause for the cortical layering phenotype, we performed 5-ethynyl-2′-deoxyuridine [EdU] labeling at E12.5, when early neurons populating deep cortical layers are born, and harvested the labeled brains at E18.5. Costaining for EdU+ and Tbr1+ cells confirmed that a greater number of Tbr1+ cells were born at E12.5 in nSR100 mutants (P < 0.05, one-way ANOVA) [Fig. 3F,G]. The increase in the number of EdU+ cells was specific to the deep cortical layer VI, as a similar number of EdU+ cells was observed in the SVZ and superficial cortical layers across all genotypes (Fig. 3H,I). Finally, we detected an overall thinning of the SVZ in nSR100 mutant brains (P < 0.05, one-way ANOVA) [Fig. 3J] that parallels the early post-mitotic commitment of progenitors and the reduction of their numbers. Taken together, these results indicate that the cortical layering defects in nSR100Δ7-8 mice are primarily caused by deregulation of the timing of early neurogenesis.

Interestingly, while analyzing cortical layering, we noticed that the morphology of the rostral part of the corpus callosum in nSR100Δ7-8/Δ7-8 mice differed from its stereotypical shape. The corpus callosum consists mostly of cortical axons crossing the midline to contact neurons of the opposite hemisphere. This interconnection between hemispheres is essential for the fast processing of information and cognition (Paul et al. 2007). Neurofilament immunostaining revealed that several callosal axons are misrouted in the absence of nSR100 and form thick ectopic fascicles similar to Probst bundles, projecting ventrally instead of crossing the midline (Fig. 4A). This phenotype is never observed in wild-type mice but is important enough in homozygous mutants to alter the shape of the corpus callosum. Although the corpus callosum of nSR100+/Δ7-8 mice does not appear grossly misshapen, it also contains ectopic ventrally projecting bundles (Fig. 4B). These observations represent the first example of a midline crossing defect as a consequence of the knockout of an AS regulator. Overall, our phenotypic survey so far shows that nSR100 controls a diverse array of neuronal functions in both the central and peripheral nervous systems, including cortical layering, axon guidance, and midline crossing.

**Figure 4.** Midline crossing defects in nSR100 mutant mice. [A] Negative grayscale images of immunofluorescence microscopy using an antibody to neurofilament on coronal sections of the rostral part of the corpus callosum of E18.5 embryos. Dashed lines with arrowheads show either the prototypical tracts of callosal axons in the wild-type (+/+ or Δ7-8/Δ7-8) and the ectopic ventral projections in the homozygous mutant (Δ7-8/Δ7-8). Arrows indicate ectopic bundles in the heterozygous and homozygous mutants. Bar, 100 μm. [B] The thickness of ventrally projecting bundles was measured at three levels on each side of the corpus callosum for three (+/Δ7-8) or four (Δ7-8/Δ7-8) individuals per genotype and on three sections for each individual. Whiskers indicate the 10th and 90th percentiles. One-tailed Mann-Whitney test.
recent results from analyzing nSR100-dependent, neural-regulated exons in cell lines (Raj et al. 2014), nSR100-regulated microexons show very strong enrichment for UGC motifs in the first several nucleotides upstream of microexons regulated by nSR100 in vivo (Fig. 5D). Of 22 analyzed differential splicing events involving alternative cassette and microexons, which were detected by RNA-seq to undergo reduced inclusion as a consequence of the loss of nSR100, all were validated by semiquantitative RT–PCR assays, including 3- and 6-nt microexons (Fig. 5E; Supplemental Fig. S6). Expression analysis based on cRPKMs (corrected reads per kilobase per million mapped reads) revealed only nine genes, other than nSR100, with an average mRNA expression difference of \( \geq 1.5 \)-fold between both replicates of wild-type and nSR100\(^{Δ7-8}\) tissues and \( P < 0.05 \) (paired t-test) (Supplemental Table S3). Analysis of genes with alternative cassette exons and microexons affected by loss of nSR100 revealed significant enrichment \( [P < 0.01] \) for gene ontology [GO] terms essential to many aspects of neuronal cell biology, such as “vesicle-mediated transport,” “neurotransmitter secretion,” “synaptosome,” and “cell projection morphogenesis” (Supplemental Fig. S7). Collectively, these observations suggest that multiple neural cassette exons—in particular, highly conserved microexons that display marked decreases in inclusion levels as a consequence of the loss of nSR100—may underlie mutant phenotypes detected in nSR100\(^{Δ7-8}\) mice.

Functions of nSR100-regulated microexons

Based on our previous and present analyses of the in vivo mutant phenotypes of zebrafish and mice lacking nSR100 and also the known functions of genes that harbor nSR100-dependent exons, a major function of the nSR100-regulated splicing program is likely to control different aspects of neurite outgrowth. Consistent with this proposal, we found that hippocampal neurons cultured from nSR100\(^{Δ7-8}\) mice have significantly shorter neurites compared with neurons from wild-type animals.
(P < 0.0001, two-tailed Mann–Whitney test) [Fig. 6A,B]. To investigate whether nSR100-regulated microexons may be responsible for neurite outgrowth, we focused on a previously uncharacterized, highly conserved nSR100 target microexon of 6 nt in Unc13b/Munc13 [Fig. 5E], a gene that has previously been shown to contribute to early neuritogenesis in primary mouse neurons [Broeke et al. 2010]. Since our RNA-seq analysis can only locate this microexon in the context of its immediate flanking constitutive exons due to short read length, Sanger sequencing of RT–PCR products from mouse brains was performed. This revealed that the Unc13b microexon, located between exons 13 and 14, is spliced in transcripts that contain at least exons 5–14 and exons 11–20.

To address whether increased skipping of the Unc13b microexon may contribute to the neuritogenesis defect in nSR100Δ7–8/Δ7–8 neurons, we harvested cortical neurons from wild-type and mutant E18.5 embryos, transfected them with red fluorescence protein [RFP]-Unc13b expression constructs that either include [Unc13b-inc] or skip [Unc13b-skp] the microexon [Fig. 6C,D], and plated them at low density. At day 2 in vitro [DIV2], the cellular distribution and expression levels of Unc13b-inc-RFP and Unc13b-skp-RFP appear similar [Fig. 6D,E, Supplemental Fig. S8A]. Both Unc13b isoforms are distributed over the entire length of the axons, thereby facilitating the visualization of transfected neurons and quantification of individual neurites. Control RFP-expressing mutant cortical neurons display the same neuritogenesis defect as hippocampal neurons (P < 0.001, Kruskal-Wallis test) [Fig. 6F]. Remarkably, wild-type neurons expressing Unc13b-skp-RFP produce neurites that are as short as mutant neurons expressing control RFP and significantly shorter than control wild-type neurons (P < 0.05, Kruskal-Wallis test) [Fig. 6E,F]. Expression of the skipped Unc13b isoform does not further affect neurite growth in mutant neurons. Strikingly, however, while expression of Unc13-inc-RFP does not affect neurite growth in wild-type neurons, inclusion of the microexon in mutant cells restores the lengths of neurites to levels observed in wild-type neurons [Fig. 6E,F]. Transfection of an nSR100-RFP expression vector in mutant neurons also results in longer neurites than in wild-type neurons expressing Unc13b-inc-RFP.
mutant cells expressing Unc13b-skp-RFP \([P < 0.05\), Kruskal-Wallis test\] [Fig. 6E,F]. Furthermore, it is noteworthy that untransfected wild-type neurons cultured in parallel with neurons transfected with any construct produced neurites of similar length, whereas shorter neurites were consistently detected in untransfected nSR100\(^{-/-}\) Δ7–8/Δ7–8 neurons \([P < 0.0001\), one-way ANOVA\] [Supplemental Fig. S6B]. This confirms that the differences in neurite length observed here are dependent on the specific construct that is expressed and are not a consequence of variance between cultures. These results thus reveal that the inclusion of a single nSR100-dependent microexon of 6 nt can positively stimulate neurite formation and rescue a neurite growth defect in primary nSR100\(^{-/-}\) Δ7–8/Δ7–8 neurons. The phenotypes that we observed in nSR100\(^{-/-}\)/Δ7–4 mice may therefore be attributed, at least in part, to the reduced inclusion of neuronal microexons.

Discussion

In this study, we generated and characterized mice deficient of nSR100/SRRM4, a vertebrate- and neural-specific splicing factor that regulates \(~30\%\) of alternative exons with increased neural inclusion, including a large number of highly conserved 3- to 27-nt microexons. We showed that the loss of nSR100 protein in vivo results in numerous neurodevelopmental defects during mouse embryogenesis that lead to early postnatal mortality in the majority of animals. We further linked these neurodevelopmental deficiencies to the loss of microexon regulation.

nSR100 regulates multiple neurodevelopmental processes

Some of the neurodevelopmental phenotypes observed in mice deficient of nSR100 may relate to altered phenotypes seen in other splicing factor knockouts, while others are unique. Neonatal lethality has been reported as a consequence of loss of the splicing regulator Ptbp2. Ptbp2 is expressed in neurons as well as in skeletal and cardiac muscle [Licatalosi et al. 2012], and mice lacking Ptbp2 are paralyzed at birth [Licatalosi et al. 2012, Li et al. 2014]. However, mice lacking Ptbp2 specifically in neurons [Ptbp2 Nestin knockout mice] die within an hour of birth, similar to nSR100 mutants, and initiate breathing at a greatly reduced rate [Li et al. 2014]. Given that nSR100 promotes the expression of Ptbp2 by activating the inclusion of an alternative exon that prevents nonsense-mediated decay of Ptbp2 transcripts (see above; Calarco et al. 2009), it is possible that the requirement for nSR100 for innervation of the diaphragm may relate, at least in part, to Ptbp2 misregulation. However, we found that only 32.8% of in vivo nSR100-regulated exons overlap with in vivo Ptbp2 targets. Therefore, overlapping and distinct exons targeted by these factors may contribute to breathing defects, paralysis, and early postnatal death. This conclusion is further supported by the observation of phenotypes that are unique to nSR100\(^{7–8/7–8}\) mice.

In addition, as in the case of nSR100\(^{7–8/7–8}\) and Ptbp2 knockout mice, mice deficient in both Nova1 and Nova2 proteins [Nova double-knockout mice] showed respiratory defects at birth [Ruggiu et al. 2009]. While phrenic nerve branching appeared normal, NMJs in E18.5 Nova double-knockout mice had few acetylcholine receptors [AChRs] and only rarely contacted motor axon terminals. In contrast, in nSR100\(^{7–8/7–8}\) mice, motor endplates [sarclemma folds in which AChRs concentrate] are of an abundance similar to those of nSR100\(^{7–8/7–8}\) heterozygous littermates, which are fully viable. However, the distribution of AChRs is altered due to the phrenic nerve deficits, which may be a consequence of altered axon branching and/or growth. While it is not clear why a small number of homozygous mutant mice are capable of surviving into adulthood, it is possible that partial and variable reductions in the levels of nSR100 target AS events may afford sufficient degrees of diaphragm innervation in some individuals. It is also known that inconsistent breathing can be regulated by an entrainment contribution from the respiratory center in the brainstem [Champagnat et al. 2011; Giraudin et al. 2012, Mellen and Thoby-Brisson 2012]. It is therefore possible that sufficient entrainment may occur early enough in a small fraction of the homozygous mutants to allow for their survival. In any case, to our knowledge, the deficit in phrenic nerve branching in nSR100 mutants has not been described previously in mice deficient of other splicing regulators.

Another neurodevelopmental aberration in nSR100\(^{7–8/7–8}\) mice that has not been previously observed in other splicing factor knockouts is the axon midline crossing defect in the corpus callosum. Approximately 60 mouse genes are known to be required for the formation of the corpus callosum [Paul et al. 2007; Donahoo and Richards 2009]. Interestingly, our RNA-seq analysis revealed that transcripts from one of these genes, Slit2, contain one of the most strongly nSR100-dependent exons. Slit2 is secreted by distinct cell populations located at or near the midline. It binds Robo receptors expressed in growing axons to help mediate midline crossing. Its function has been extensively studied in vivo [Chedotal 2007], and a Slit2 knockout mouse displays a midline crossing defect that is strikingly similar to the one that we observed in nSR100\(^{7–8/7–8}\) mice, with bundles of callosal axons projecting ventrally along the midline [Unni et al. 2012]. The nSR100-dependent Slit2 microexon adds nine amino acids to the fifth EGF domain in the secreted N-terminal portion of the protein that is responsible for its repulsive activity during axon guidance. The differential activities of the resulting Slit2 isoforms have not been previously investigated, although an AS event in Robo3 that switches the axonal response to Slit proteins from attraction to repulsion has been reported [Chen et al. 2008]. It is interesting to consider that the nSR100-dependent regulation of the alternative Slit2 exon represents a complementary mechanism for controlling axon guidance and may contribute to the midline crossing defect observed in nSR100 mutant mice. However, because the loss of
nSR100 also affects axon growth and the number and distribution of neurons that project callosal axons, we cannot exclude the possibility that a combination of these mechanisms may contribute to this anomaly.

In addition to the differences observed in the corpus callosum of nSR100 heterozygotes and homozygotes, we discovered nSR100 dosage-dependent cortical deficits. It is noteworthy that subtle defects in the corpus callosum and cortical layering have been linked to impaired cognitive and behavioral function in humans (Paul et al. 2007). For example, disruption of cortical layer formation has been observed in individuals with schizophrenia and autism (Akbarian et al. 1993; Ross et al. 2006; Stoner et al. 2014). While cell migration defects often result in the aberrant positioning of cortical layers (Caviness 1982, Kwan et al. 2008), early production of post-mitotic neurons by cortical progenitors has been shown to result in an expansion of deep cortical layers (Feng and Walsh 2004). Premature production of neurons depletes the pool of progenitors and results in fewer late-born neurons being generated. In nSR100Δ7–8/Δ7–8 mice, we found that layer VI is significantly expanded in mutant brains at E18.5, whereas the total number of neurons [including the number of superficial neurons] and neural progenitors is decreased. These phenotypes suggest that loss of nSR100 causes neural progenitors to fail to accomplish asymmetric division early during brain development and commit prematurely to a post-mitotic fate. Further supporting this proposal is our finding that more neurons are produced during the first steps of cortical development when nSR100 is lost. Finally, although the preplate was poorly defined in the brains of mutant mice, our fate-mapping experiments provide evidence that early-born, nSR100-depleted neurons migrate correctly to populate deep cortical layers. These observations thus establish nSR100 as an important regulator of neurogenesis timing.

nSR100 regulates AS events in genes with important neuronal functions

Our previous studies directed at the identification of nSR100 targets in cell lines focused on cassette alternative exons and revealed that nSR100 predominantly promotes neural exon inclusion (Calcaro et al. 2009; Raj et al. 2011; Irimia et al. 2014). The RNA-seq analyses performed here confirmed this property of nSR100 and further encompassed a comprehensive survey of all classes of nSR100-regulated AS events detected in vivo. Notably, 65 of the 138 (47%) longer cassette exons and microexons that are promoted by nSR100 in vivo had not previously been reported as nSR100 targets. Many of these events occur in genes with essential roles in neuronal biology and/or nervous system development (e.g., Ahi1, Camk2b, Dvl1, Itsn1, Shank1, and Unc13b). Moreover, in addition to changes in the inclusion levels of a large number of neural cassette exons, of which many are microexons [see below], the present work also revealed that the nSR100 regulatory program extends to all classes of AS events. For example, many retained introns are misregulated in developing nSR100Δ7–8/Δ7–8 mouse brains. Although a subset of the retained introns introduce premature termination codons, it appears that in most cases, the corresponding transcripts are not subject to nonsense-mediated mRNA decay, as their steady-state levels were not appreciably affected in nSR100Δ7–8/Δ7–8 brain tissue [data not shown]. We also identified a small number of nSR100-dependent alternative 5’ and 3’ splice site selection events, most of which are frame-preserving.

Collectively, AS events misregulated in nSR100Δ7–8/Δ7–8 mouse brains are enriched in genes involved in neuronal functions, such as genes associated with neuronal differentiation (Zmynd8 and Ahi1), neurite outgrowth (Zfyve27 and Clasp2), and axon guidance (Slit2, Ncam, and MycBp2). Many of these genes possess pivotal roles as scaffolding proteins for endocytosis, exocytosis, cytoskeleton remodeling, and vesicle transport and are associated with defects similar to the ones that we observed in our mouse model.

Functional impact of nSR100-regulated microexons

Among genes that contain microexons regulated by nSR100, several encode proteins that are known to interact. These proteins form a network that is involved in the trafficking and recycling of vesicles, including I1s1n1, Ppia2, Rims2, Dmm2, Nbea, Abi1, Ptprd, and Vav2. Strikingly, 65 of the 72 nSR100-activated microexons are frame-preserving and have the potential to result in the insertion of one to nine amino acid residues in the corresponding protein products. These seemingly modest changes to coding sequence raise interesting questions as to the functional roles of microexons.

Recently, we and others have observed that amino acid residues encoded by microexons are almost invariably surface-accessible and enriched within—or immediately adjacent to—domains involved in protein–protein or protein–lipid interactions (Irimia et al. 2014; Li et al. 2015). Consistent with these observations, deletion of microexons reduces interactions with partner proteins. For example, a microexon in the SH3 domain of the Down syndrome-associated gene I1s1n1, which we show here is strongly regulated by nSR100 [Supplemental Fig. S5], promotes interactions with multiple partners (Tsyba et al. 2008). A recent report demonstrated that an nSR100-regulated microexon in the Zfyve27 transcript [Fig. 5E] promotes interactions with partner proteins VAP-A and VAP-B [Ohnishi et al. 2014]. Furthermore, we showed recently that neural microexons in the API endocytic transport complex subunit Ap1s2 and the amyloid β precursor protein-binding family B member 1 (Abpb1), which is also associated with neurogenesis [Cheung et al. 2014], promote interactions with their respective partner proteins [Irimia et al. 2014]. In the present study, we extend these findings by demonstrating that nSR100-dependent inclusion of a 6-nt microexon in Unc13b transcripts is sufficient to promote the increased length of neurites and rescue a neurogenesis defect in nSR100 mutant neurons. This microexon has the potential to add two amino acids adjacent to a predicted MAPK-docking site in the Unc13b protein. It is therefore interesting to consider that the
nSR100-dependent regulation of this microexon affects the phosphorylation status of Unc13b in ways that alter interactions required for neurite formation.

Finally, it is intriguing to note that most (~76%) of the microexons affected by the in vivo loss of nSR100 in the present study are conserved in humans, and many (~46%) of these display loss of inclusion in the brain cortices of subjects with ASD [Irimia et al. 2014]. Furthermore, this altered pattern of inclusion in ASD subjects affects genes enriched in known genetic associations with ASD and is also highly correlated with reduced expression of nSR100 [Irimia et al. 2014]. Additional studies have linked microexon misregulation to schizophrenia and epilepsy [Ovadia and Shifman 2011; Rusconi et al. 2014]. In the future, it will be of considerable interest to address whether partial alteration of nSR100 levels in developing mice may contribute to phenotypes associated with human neurological disorders.

Materials and methods

nSR100 mutant mouse generation

Stem cells containing the conditional nSR100lox allele were obtained from EUCCOM [project no. 71507, clones EPD0538_3_A08 and EPD0538_3_A09] (Friedel et al. 2007) and aggregated with outbred ICR morula. Following confirmation of germline transmission, mice bearing the nSR100lox allele were maintained on a C57BL/6N background and crossed with the B6.C-Tg(CMV-cre)1Cgn/J line from the Jackson Laboratory. Excision of exons 7 and 8 in the resulting nSR100Δ7-8 allele was confirmed by PCR and sequencing.

We attempted to generate mice lacking the partial (25-kDa) polypeptide detected in the nSR100Δ7-8lox allele [Fig. 1] by removing both the Frt site-flanked LacZ reporter and exons 7 and 8 by sequential Flpase and Cre recombinase crosses [Supplemental Fig. S9A]. However, the resulting nSR100Δ7-8lox allele produced an nSR100 protein migrating at ~90 kDa, which is consistent with the predicted size of a partial nSR100 protein lacking only exons 7 and 8 [Supplemental Fig. S9B]. We therefore focused our analyses on the nSR100Δ7-8 allele.

Southern blotting

Southern blotting was performed as described elsewhere [Sambrook and Russell 2001]. Briefly, 60 μg of mouse genomic tail DNA was digested with Ascl and loaded on a 0.8% agarose gel for each genotype. DNA was transferred to a Hybond XL membrane (GE Healthcare Life Sciences) and hybridized with a 32P-dCTP-labeled probe encompassing 456 base pairs [bp] of nSR100 intron 3 upstream of the S′ homology arm used for homologous recombination of the nSR100lox allele.

RT–PCR

Semi-quantitative RT–PCR was performed using the Qiagen one-step RT–PCR kit according to the manufacturer’s instructions using 15 ng of total RNA template per 10-μL reaction and run on 2% or 4% agarose gels. Radiolabeled reactions included 0.05 μCi of 32P-dCTP and were run on 6% Sequalag urea gels [National Diagnostics]. Bands were quantified using Image Lab (Bio-Rad) or ImageJ. Primer sequences are available on request.

Antibodies

For immunoblotting, a polyclonal rabbit antibody [Calarco et al. 2009] raised against amino acids 1–82 of nSR100 was used at 1:5000. Anti-tubulin (T6074, Sigma) was used at 1:5000. For immunostaining, mouse monoclonal anti-neurofilament (2H3 conditioned medium, Iowa Developmental Studies Hybridoma Bank) was diluted to 1:50 for whole-mount diaphragm staining and 1:100 for brain section staining. Mouse anti-NeuN (mab377, Millipore), mouse anti-Satb2 [ab51502, Abcam], rabbit anti-Tbr1 [ab31940, Abcam], and chicken polyclonal anti-β-galactosidase [ab9361, Abcam] were all diluted to 1:500. Chicken anti-MAP2 [ab5392, Abcam] was diluted to 1:10,000, mouse anti-Tuj1 [MRB-435P, Covance] was diluted to 1:750, and rabbit anti-Pax6 [PRB-278P, Covance] was diluted to 1:1500. For in situ hybridization, an anti-DIG antibody conjugated to alkaline phosphatase [Roche] was diluted to 1:5000.

In situ hybridization

In situ hybridization was essentially performed as previously described [Sambrook and Russell 2001]. Twenty-micrometer brain sections were post-fixed in 4% formaldehyde for 10 min at room temperature. Sections were then prehybridized for 3–6 h at room temperature followed by hybridization with sense or antisense DIG-labeled probes to nSR100 exons 9–13 diluted to 200 ng/mL overnight at 60°C. Alkaline-phosphatase-conjugated anti-DIG antibody was added to slides for 1 h at room temperature and washed, and an NBT/BCIP solution [Roche] was applied for 1 h to overnight at room temperature. Sections were cleared in xylene and mounted in Cytoseal XYL [Thermo Scientific].

Immunofluorescence

For whole-mount diaphragm staining, diaphragms were dissected from E16.5 or E18.5 embryos and fixed in 2% formaldehyde overnight at 4°C. Diaphragms were washed in 0.1 M glycine in PBS and blocked overnight at 4°C in 0.5% Triton X-100, 3% BSA, and 5% donkey serum with Alexa-594-coupled α-bungarotoxin diluted at 1:1500 [Life Technologies]. Diaphragms were then further permabilized briefly in 100% methanol, fixed again in 0.2% glutaraldehyde and 4% formaldehyde for 20 min at room temperature, and then incubated overnight at 4°C in blocking buffer with a monoclonal anti-neurofilament antibody diluted to 1:100. After extensive washes, samples were incubated overnight at 4°C in blocking buffer with an Alexa-488 anti-mouse antibody diluted to 1:500 [Life Technologies]. For immunofluorescence microscopy of cortical sections, embryonic mouse brains were dissected and fixed in 4% formaldehyde for 60 min at 4°C. Brains were then sucrose-protected, OCT-embedded [Tissue-Tek], frozen, and coronally sectioned to 16 μm. Sections were blocked and permeabilized in 5% BSA and 0.3% Triton X-100 in PBS with or without M.O.M. blocking reagent [Vector Laboratories] for 60 min at room temperature. All primary antibodies were incubated overnight at 4°C with or without M.O.M. protein concentrate. Secondary antibodies were goat anti-mouse, anti-rabbit, or anti-chicken coupled to Alexa-488, Alexa-568, or Alexa-594 dyes, all diluted to 1:500 and incubated for 60 min at room temperature with or without M.O.M. protein concentrate. Stained sections were mounted in VectaShield mounting medium with DAPI [Vector Laboratories].

EdU labeling

Pregnant dams were injected with 0.1 mg/g EdU at E12.5. Embryos were harvested at E18.5, and brains were dissected, fixed, and...
cryosectioned as described above. Immunostaining using an antibody to Tbr1 was first performed as described above. EdU+ cells were then stained using the Click-IT Alexa Fluor-488 kit (Life Technologies) according to the manufacturer’s protocol.

Quantification of phenotypic data

For neurite length measurements on whole-mount diaphragms, tracings were generated with the NeuroJ plug-in for ImageJ. Right branches on the ventral and dorsal parts of the diaphragm were measured and counted. For NMJs, a 475-μm-long region was selected over the ventrally projecting left primary branch, and NMJs were quantified using the ITCN plug-in in ImageJ. The dispersion of NMJs was measured as the average width of the NMJ band at three different levels of the same region of interest. Numbers of cells were counted using the ITCN plug-in over 300-μm radial unit regions for Tbr1+, Satb2+, NeuN+, and Pax6+ cells and a 750-μm radial unit region for EdU+ cells. Cells and layers were quantified using three sections from each brain. SVZ thickness was measured relative to the total thickness of the cerebrocortex from the lateral ventricle to the surface of layer 1.

RNA-seq analysis

A first replicate consisted of total RNA extracted from cerebral cortices and hippocampi dissected from five wild-type and five nSR10047–8/47–8 mutant brains at E18.5. RNA was pooled by genotype and prepared using the Illumina TruSeq mRNA kit, and cortical and hippocampal samples were sequenced on different runs of Illumina HiSeq2500 (average of ~93 million 100-nt single end and 100-nt paired-end reads for each run, respectively) (see Supplemental Table S4 for details). A second replicate was processed as above and consisted of total RNA pooled from three wild-type or three mutant brains at E18.5. An average of ~90 million 100-nt paired-end reads was sequenced for each sample (Supplemental Table S4).

Transcriptome-wide AS and gene expression profiling was performed using our recently described pipeline [vast-tools] (Irimia et al. 2014: vast-tools uses reads mapping to exon–exon (or exon–intron) junctions (EE) or EEJ) only to accurately detect and quantify all types of AS events, including 3- to 27-nt microexons. Gene expression levels were measured using the cRPKM metric (Labbe et al. 2012).

PSI/PIR/PSU of AS events for the eight samples were paired and four replicates [wild-type and nSR10047–8/47–8 for two cortex and two hippocampus samples], and a paired t-test was performed for AS events with enough read coverage in all eight samples. A given AS event was considered to have sufficient read coverage in a particular RNA-seq sample according to the following criteria (Irimia et al. 2014):

- For cassette exons (except for those quantified using the microexon pipeline), [1] ≥10 actual reads (i.e., before mappability correction) mapped to the sum of exclusion EEJ or [2] ≥10 actual reads mapped to one of the two inclusion EEJs and five or more mapped to the other inclusion EEJ.
- For microexons, [1] ≥10 actual reads mapped to the sum of exclusion EEJs or [2] ≥10 actual reads mapped to the sum of inclusion EEJs.
- For IR, [1] ≥10 actual reads mapped to the sum of skipping EEJs or [2] ≥10 actual reads mapped to one of the two inclusion EEJs and five or more mapped to the other inclusion EEJ.
- For Alt3 and Alt5, ≥10 actual reads mapped to the sum of all EEJs involved in the specific event.

For an AS event to be considered differentially regulated between wild-type and nSR10047–8/47–8 brains, we required a P-value of <0.05 in the t-test and an average ΔPSI (between the four paired replicates) of at least 10%.

All RNA-seq data have been deposited in the Gene Expression Omnibus database under accession number GSE65818.

Functional enrichment analyses

Ensembl gene IDs for the cassette exons and microexons that showed significantly increased skipping in nSR10047–8/47–8 brains (137 genes in total) were uploaded to DAVID ([http://david.abcc.ncifcrf.gov, Huang et al. 2009]) to perform functional enrichment analyses using a stringent background consisting of 10,968 genes with expression of at least cRPKM >2 in one of the brain samples. Only GO_FAT terms and KEGG (Kyoto Encyclopedia of Genes and Genomes) pathways were used for clustering analyses.

Primary neuronal cultures

Protocols for culturing primary mouse neurons were kindly provided by Dr. Antony Bourcud and Dr. Thomas Sudhof (Stanford University) (Bourcud et al. 2005). Briefly, hippocampal or cortical neurons were harvested from wild-type or nSR10047–8/47–8 mice at E18.5 and plated on glass coverslips coated with 2% Matrigel (Corning) in plating medium consisting of MEM (51200-038, Life Technologies) supplemented with 0.5% glucose, 0.2 mg/mL NaHCO3, 0.1 mg/mL transferrin (616420, Calbiochem), 10% fetal bovine serum (FBS) (SH30093-03, GE Life Sciences), 2 mM L-glutamine (12403-010, Life Technologies), and 25 μg/mL insulin (I-6634, Sigma). Plating medium was changed at DIV1 to growth medium consisting of MEM supplemented with 0.5% glucose, 0.2 mg/mL NaHCO3, 0.1 mg/mL transferrin, 5% FBS, 0.5 mM L-glutamine, and 2% B-27 supplement (17504-044, Life Technologies). Dissociated neurons were transfected prior to plating using the Axama Nucleofector kit (VPG-1001, Lonza) using ~5 x 10⁴ cells and 10 μg of plasmid DNA per transfection. Cells from the same embryos were aliquoted and transfected individually with each construct. Unc13 skp, Unc13 inc, and nSR100 were cloned upstream of the RFP coding sequence and placed under the control of the CAGGS promoter. The length of neurites was quantified using the NeuroJ plug-in for ImageJ.

Acknowledgments

We thank Marina Gertsenstein, Sandra Tondat, Monica Pereira, Christina Dalrymple, Jorge Cabezaz, Jessica Raponi, and Amanda Leonelli at the Toronto Centre for Phenogenomics for assistance with the generation and maintenance of mutant mouse strains. Dax Torti and Danica Leung of the Donnelly Sequencing Centre are gratefully acknowledged for sequencing samples. We also thank Jonathan Ellis for generating constructs used in primary neuronal cultures, and members of the Blencowe and Cordes laboratories for helpful discussions and comments on the manuscript. This research was supported by grants from the Canadian Institutes of Health Research (CIHR; MOP-67011, MOP-14609, and MOP-111199) to B.J.B. and S.P.C. M.L. was supported by a long-term Fellowship from the Human Frontiers Science Program Organization. M.Q.-V. was supported by CIHR Banting and Best Scholarship and Ontario Graduate Scholarship. B.J.B. holds the Banbury Chair of Medical Research at the University of Toronto.
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Essential roles for the splicing regulator nSR100/SRRM4 during nervous system development

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*Genes Dev.* 2015 29: 746-759
Access the most recent version at doi:10.1101/gad.256115.114

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